Lecture = Classroom instruction (unless otherwise noted, all lectures in MBL's Speck Auditorium)
Demo = Instructor physically demonstrates lecture concepts
Hands-on = students practice concepts with computers
Lab = Students participate in hands-on lab activities
Discussion = Group discussion

Tour = Visit site at MBL or WHOI, but no hands-on lab work

| Yellow = Segment 1 (Carbonate system measurments)
| Blue = Segment 2 (OA experimental set-up: biology/chemistry)
| Orange = Segment 3 (Biogeochemical modeling)
| Green = Segment 4 (Data reporting)

WEEK 1	NOVEMBER 2 MONDAY	NOVEMBER 3 TUESDAY	NOVEMBER 4 WEDNESDAY	NOVEMBER 5 THURSDAY	NOVEMBER 6 FRIDAY	NOVEMBER 7 SATURDAY	NOVEMBER 8 SUNDAY
Morning Period 1 (8:30-10:00)	Lecture. Carbonate system overview (Sabine/Dickson)	Lecture. pH and Alkalinity (Dickson)	control and advantages/disadvantages of	Lecture. Overview on approaches and tools to manipulate seawater carbonate chemistry (Gattuso)	Lecture. Algal culturing overview (Iglesias- Rodriguez)	Lab. T2 sampling for corals, pH monitoring for mollusk exp't (Holcomb)	Free
Morning Period 2 (10:30-12:00)	Lecture/Hands-on. Intro to COZSYS and seacarb (Sabine and Gattuso)	Lecture. Overview on biogeochemical feedbacks (Hutchins)	Lecture/Discussion. Implications of uncertainties in equilibrium constants and analytical measurements and choosing optimal parameters to measure for an experiment (Dickson/Sabine)	Lecture/Demo. Moored CO2 instrument demo (Sabine)	Lecture. Calcification overview (bkgd for coral calcification and larval mollusk exp'ts), methods for measuring coral and bivalve calcification (Cohen/Miller)	Lecture. Radioisotope Techniques for measuring coccolithophore calcification (Balch) and PIC methods for quantifying calcification (Iglesias-Rodriguez)	
Afternoon Period (13:30-17:30)	Lecture/Lab. Lab safety presentations followed by sampling instruction (movie plus hands-on) and underway pCO2 introduction and demo (Groups A/B/C)	Lab. pH and infrared DIC measurements, coulometric DIC setup demo (Groups D/E/F)	Lab. TA and DIC measurements (Groups D/E/F)	Lab/Hands-on. Lecture/Demo/Hands-On. Theoretical CO2SY'S (Yates) experiments and setup for larval mollusk experiments (McCorkle)	Lab. Coral calcification setup (Holcomb - Groups G/H/I)	Lab. T3 sampling for corals (Holcomb)	Lab. T4 sampling for corals and coral exp't breakdown, pH monitoring for mollusks. (Holcomb)
					Tour. Bernhard lab	Lecture. Measuring calcification in the field (Langdon)	
Evening Period (19:00-22:00)	WELCOME RECEPTION AND POSTER SESSION, WHOI Clark 507		Lecture. Overview onf ocean acidification experimental design (Langdon)		Lab. Larval mollusk inoculations (White/McCorkle). Coral T1 sampling (Holcomb)		

WEEK 2	NOVEMBER 9 MONDAY	NOVEMBER 10 TUESDAY	NOVEMBER 11 WEDNESDAY	NOVEMBER 12 THURSDAY	NOVEMBER 13 FRIDAY
Morning Period 1 (8:30-10:00)	Lab. Larval mollusk harvesting (Cohen/Rose/White)	Lecture. Biogeochemical modeling oveview (Doney)	Lecture/Hands-on. Large databases (e.g., GLODAP), etc. (Key)	Lecture. Introduction to Data Management and Better Practices for Shipboard Data Management (Chandler)	Lecture. Data reporting overview (Kozyr)
Morning Period 2 (10:30-12:00)		Lecture. Physiology (Seibel)	Lecture. Genomics applications to ocean acidification research (Fangue)	Lecture. Biogeochemical Modeling Part II (Doney)	Course wrap-up: Open discussion and course evaluations
Afternoon Period (13:30-17:30)	Lecture/Hands-on. Ocean Data View (Schlitzer)	Lab. Larval mollusk microscopy (Cohen/Rose) Cellular pH setups and initial pH measurements (Seibel)	Lab/Hands-On. Cellular pH final measurements and respirometry demo (Seibel)	Synthesis and discussion	Departures
	Tours. MBL Marine Resources Center and Waterbury algal culturing lab		Lab. Modeling exercises (Doney)		
Evening Period (19:00-22:00)				FAREWELL DINNER, MBL	