

Fourth Symposium on Harmful Algae in the U.S.

Symposium Agenda, Abstracts and Participants



**October 28 - November 1, 2007
Woods Hole, Massachusetts**

Fourth Symposium on Harmful Algae in the U.S.

Symposium Director:

Donald M. Anderson

Symposium Coordinator:

Judy Kleindinst

Steering Committee:

Don Anderson	Woods Hole Oceanographic Institution
Darcie Couture	Maine Department of Marine Resources
Deana Erdner	University of Texas at Austin
Stacey Etheridge	Center for Food Safety and Applied Nutrition, US FDA
Chris Gobler	Stony Brook University
Jan Landsberg	Florida Fish and Wildlife Commission
Cary Lopez	NOAA Coastal Ocean Program
Chris Scholin	Monterey Bay Aquarium Research Institute
Pete Stratton	Oregon State University
Marc Suddleson	NOAA Coastal Ocean Program
Peter Tango	U.S. Geological Survey / Chesapeake Bay Program

Session Chairs:

Bloom Ecology & Dynamics	Pete Stratton, Pete Tango, Deana Erdner
Toxins and Toxin Detection:	Stacey Etheridge
Fisheries and Food Webs:	Chris Gobler
Communication, Outreach and Education:	Darcie Couture
Human Health:	Darcie Couture
Emerging Technologies:	Chris Scholin

Sponsor:

NOAA / Center for Sponsored Coastal Ocean Research / Coastal Ocean Program

Student support:

NOAA / Center for Sponsored Coastal Ocean Research / Coastal Ocean Program
Miami Center for Oceans and Human Health
Woods Hole Center for Oceans and Human Health

Other support:

Fluid Imaging Technologies

Fourth Symposium on Harmful Algae in the U.S.

October 28 – November 1, 2007

Woods Hole, Massachusetts



**Symposium Agenda, Abstracts
and Participants**

**Fourth Symposium on Harmful Algae in the U.S.
October 29 – November 1, 2007**

Schedule	Mon., Oct. 29	Tues., Oct. 30	Wed., Oct. 31	Thu., Nov. 1
7:00 – 8:30	Breakfast	Breakfast	Breakfast	Breakfast
8:30	Welcome and Introduction			
8:40-10:00	<u>Session 1</u> Bloom Ecology & Dynamics	<u>Session 5</u> Toxins & Toxin Detection	<u>Session 9</u> Bloom Ecology & Dynamics	<u>Session 13</u> Emerging Technologies
<i>Break 10:00 - 10:30</i>				
10:30-11:50	<u>Session 2</u> Toxins & Toxin Detection	<u>Session 6</u> Communication, Outreach & Education	<u>Session 10</u> Emerging Technologies	<u>Session 14</u> Bloom Ecology & Dynamics
<i>Lunch 12:00-1:30</i>				
1:30-2:30	<u>Session 3</u> Fisheries & Foodwebs	<u>Session 7</u> (Bloom Ecology & Dynamics)	<u>Session 11</u> Fisheries & Foodwebs	<u>Session 15</u> Human Health
<i>Break 2:30-3:00</i>				
3:00-4:00	<u>Session 4</u> (Bloom Ecology & Dynamics)	<u>Session 8</u> Human Health	<u>Session 12</u> Bloom Ecology & Dynamics	<u>Session 16</u> Bloom Ecology & Dynamics
4:30-5:30				Community Meeting – HABHRCA update & site selection for next meeting
<i>Dinner 6:00-7:30</i>				
Poster Session & refreshments 4:00 – 6:00	<u>Poster Session 1</u> Bloom Ecology & Dynamics; Communication, Outreach & Education	<u>Poster Session 3</u> (Toxins & Toxin Detection; Human Health)	<u>Poster Session 4</u> Fisheries & Foodwebs; Emerging Technologies	
Evening Sessions 7:30 – 9:30	<u>Poster Session 2</u> Bloom Ecology & Dynamics	Discussion sessions	Banquet and party (till 11:00 p.m.)	

**Fourth Symposium on Harmful Algae in the U.S.
October 28 – November 1, 2007
Woods Hole, Massachusetts**

Please note that all meals (breakfast, lunch and dinner) will be served in the MBL Dining Hall, Swope upstairs.

Breakfast	7 – 8:30 each day (Monday through Friday)
Lunch	12 – 1:30 each day (Monday through Thursday)
Dinner	6 – 7:30 each day (Sunday through Thursday)

All talks will be in the Lillie Auditorium; am and pm breaks will be there as well.

Poster sessions will be held in the Swope upstairs lobby and Meigs Room (also located in Swope upstairs).

Locations for discussion sessions (Tuesday evening) will be announced at the meeting. Current plans are for four concurrent discussion sessions:

- operational HAB forecasting system
- HAB genomics
- Identifying barriers to HAB technology utilization
- volunteer monitoring networks

The registration desk will also be open each day from approximately 8:00 am to 8:30 am.

SUNDAY, OCTOBER 28, 2007

4:00 - 6:00	Registration, Swope Lobby (downstairs)
4:00 – 6:00	Mixer, Meigs Room (Swope Building upstairs)
6:00 – 7:30	Dinner, Swope Dining Hall (upstairs)
7:30 – 9:30	Reception, Meigs Room (Swope Building upstairs)

MONDAY, OCTOBER 29, 2007**Session 1: Bloom Ecology and Dynamics - Chair: P. Strutton (Lillie Auditorium)**

Time	Presenter	Page #	Title
8:30	Don Anderson		Welcome and Opening Remarks
8:40	Barbara Hickey	45	Regional oceanography leading to toxic <i>Pseudo-nitzschia</i> events on beaches in the Northern California Current
9:00	Evelyn Lessard	53	Seasonal and interannual variability of <i>Pseudo-nitzschia</i> and domoic acid in the Juan de Fuca eddy region and its adjacent shelves
9:20	Bill Cochlan	31	Silicic acid limitation is not a trigger for domoic acid production by <i>Pseudo-nitzschia</i> blooms in the Pacific Northwest
9:40	Katherine Hubbard	48	Temporal and spatial variability in Pacific Northwest <i>Pseudo-nitzschia</i> populations

Break 10:00– 10:30**Session 2: Toxins and Toxin Detection - Chair: S. Etheridge (Lillie Auditorium)**

10:30	Al Place	62	Scrambled modules, spliced leaders, cap dependent translation control – what next in dinoflagellate polyketide toxin synthesis?
10:50	Emily Monroe	59	Novel structure of polyketide synthase gene transcripts in the Florida red tide dinoflagellate, <i>Karenia brevis</i>
11:10	Gerry Plumley	57	A genomic approach for identifying the saxitoxin (STX) synthesis genes
11:30	Orlando Sarnelle	68	Relative importance of zebra mussel invasion, phosphorus and other environmental factors on microcystin concentrations in lakes

Lunch 12:00 – 1:30**Session 3: Fisheries and Food Webs – Chair: C. Gobler (Lillie Auditorium)**

1:30	Roz Jester	49	Recent ecosystem shift in central California alters harmful algal bloom patterns
1:50	Porter Hoagland	46	The New England 2005 <i>Alexandrium</i> bloom: Estimates of the economic effects on commercial shellfisheries
2:10	Pat Glibert	40	Impacts of eutrophication-related blooms of <i>Prorocentrum minimum</i> and <i>Karlodinium veneficum</i> on early life stages of oysters in Chesapeake Bay

Break 2:30 – 3:00**Session 4: Bloom Ecology and Dynamics – Chair: P. Strutton (Lillie Auditorium)**

3:00	Jason Adolf	20	Cryptophytes in Chesapeake Bay and their potential relationship to mixotrophic harmful algal blooms
3:20	Wayne Litaker	56	Development of a toxic dinoflagellate (<i>Karlodinium veneficum</i>) bloom in a shallow, eutrophic, lagoonal estuary
3:40	Ted Smayda	72	Harmful algal blooms and the 15°C barrier

4:00 – 6:00 **Poster Session 1: Bloom Ecology & Dynamics; Communication, Outreach & Education (Swope Upstairs Lobby and Meigs Room)**

Dinner 6:00 – 7:30 (Swope Dining Hall)

7:30 – 9:30 **Poster Session 2: Bloom Ecology & Dynamics**

TUESDAY, OCTOBER 30, 2007**Session 5: Toxins and Toxin Detection – Chair: S. Etheridge (Lillie Auditorium)**

Time	Presenter	Page #	Title
8:40	Mark Poli	38	Determination of paralytic shellfish poisoning toxins using the Lawrence method: Application to human urine and serum
9:00	Greg Boyer	47	New tricks with old toys: Application of mass spectrometry to the analysis of peptide toxins
9:20	Steven Plakas	63	Monitoring of brevetoxins in <i>Karenia brevis</i> bloom-exposed eastern oyster
9:40	Faisal Radwan	65	A potent effect of in vitro gastric digestion on the overall toxicity of brevetoxin-laden Atlantic menhaden (<i>Brevoortia tyrannus</i>)

Break 10:00– 10:30**Session 6: Communication, Outreach and Education – Chair: D. Couture (Lillie Auditorium)**

10:30	Lora Fleming	39	Florida aquatic toxins hotline: Formal evaluation of HAB outreach and educational activities
10:50	Sparkle Roberts	67	A comparative study of perceived risk from two coastal communities: Implications for communication and education
11:10	Cliff Scherer	25	Using social science to develop communication messages that facilitate public trust and understanding regarding harmful algal bloom control
11:30	Pat Tester	74	Red tide related losses and small business administration loans: A 20 year retrospective

Lunch 12:00 – 1:30**Session 7: Bloom Ecology and Dynamics – Chair: P. Tango (Lillie Auditorium)**

1:30	Kristy Lidie	54	Characterization and regulation of gene expression networks in response to acute stress in <i>Karenia brevis</i>
1:50	Emily Prince	64	Chemically-mediated competition: Interactions between the red tide dinoflagellate, <i>Karenia brevis</i> , and co-occurring phytoplankton
2:10	Geoff Sinclair	71	Can benthic-pelagic coupling by <i>Karenia brevis</i> support perennial offshore seed populations for coastal blooms?

Break 2:30 – 3:00**Session 8: Human Health – Chair: D. Couture (Lillie Auditorium)**

3:00	Lynn Grattan	42	Domoic acid neurotoxicity in Native Americans in the Pacific Northwest: Human health project methods and update
3:20	Kathi Lefebvre	52	Gene expression in zebrafish after acute and sub-acute exposure to the marine neurotoxin domoic acid
3:40	Lori Backer	23	Recreational exposure to microcystins during a <i>Microcystis aeruginosa</i> bloom in a small lake

4:00 – 6:00 **Poster Session 3: Toxins & Toxin Detection; Human Health (Swope Upstairs Lobby and Meigs Room)**

Dinner 6:00 – 7:30 (Swope Dining Hall)

7:30 – 9:30 **Concurrent Discussion Sessions (topics & locations to be announced)**

WEDNESDAY, OCTOBER 31, 2007**Session 9: Bloom Ecology and Dynamics – Chair: D. Erdner (Lillie Auditorium)**

Time	Presenter	Page #	Title
8:40	Timothy Davis	32	The effects of temperature and eutrophication on toxic and non-toxic strains of <i>Microcystis</i> within New York lakes
9:00	Juli Dyble	35	Assessing the role of environmental stressors and genetic composition on microcystin production in Lake Erie <i>Microcystis</i> populations
9:20	David Avery	22	The evolution of toxin resistance in copepods: How do copepods respond to blooms of toxic <i>Alexandrium fundyense</i> ?
9:40	Andy Juhl	50	Development of quantitatively PCR-based techniques for assessing zooplankton grazing on harmful algae: A tale of two species

Break 10:00– 10:30**Session 10: Emerging Technologies – Chair: C. Scholin (Lillie Auditorium)**

10:30	Sonya Dyhrman	36	Monitoring toxic <i>Alexandrium catenella</i> in the Puget Sound using real-time quantitative PCR (qPCR)
10:50	Greg Doucette	34	Autonomous, sub-surface detection of the algal toxin domoic acid onboard the environmental sample processor
11:10	Senjie Lin	55	Genetic network regulating cell division and toxin production in <i>Karlodinium</i> and <i>Amphidinium</i> : A genomic approach
11:30	Chris Gobler	41	Preliminary insight from the first genome-sequence of a harmful algal bloom species, the brown tide alga, <i>Aureococcus anophagefferens</i>

Lunch 12:00 – 1:30**Session 11: Fisheries and Food Webs – Chair: C. Gobler (Lillie Auditorium)**

1:30	Lihua Chen	30	Isolation of the sodium channel gene from the copepod <i>Acartia hudsonica</i> and its potential link to saxitoxin resistance
1:50	Monica Bricelj	27	Applications of video-endoscopy to study the effects of HAB species on suspension-feeding bivalves
2:10	Hélène Hégaret	44	Effect of the harmful alga <i>Prorocentrum minimum</i> on the hemocyte response of quahogs <i>Mercenaria mercenaria</i> with various levels of QPX infection

Break 2:30 – 3:00**Session 12: Bloom Ecology and Dynamics – Chair: D. Erdner (Lillie Auditorium)**

3:00	Rick Stumpf	73	Blending of observations and models in forecasting transport of harmful blooms
3:20	Mike Twiner	75	Gene expression profiles of <i>Karenia brevis</i> during lysis by algicidal bacteria
3:40	Mike Parsons	60	Observations on the epiphytic relationship between <i>Gambierdiscus</i> spp. and several macroalgal host species

4:00 – 6:00 **Poster Session 4: Fisheries & Food Webs; Emerging Technologies****Lobster Dinner 6:00 – 7:30 (Swope Dining Hall)**8:00 – 11:00 **Halloween Party (Meigs Room, Swope Building)**

THURSDAY, NOVEMBER 1, 2007**Session 13: Emerging Technologies – Chair: C. Scholin (Lillie Auditorium)**

Time	Presenter	Page #	Title
8:40	Christopher Brown	29	Implementation of a harmful algal bloom prediction system in Chesapeake Bay
9:00	Steve Wilhelm	77	Molecular characterization of toxic cyanobacterial communities in the lower Great Lakes: A seven year synopsis
9:20	Gary Kirkpatrick	51	The optical-based HAB detection observatory: Lessons learned during 4 years of implementation
9:40	Dianne Greenfield	43	Applications of the Second-Generation Environmental Sample Processor (2G ESP) for remote detection of harmful algae: 2007

Break 10:00– 10:30**Session 14: Bloom Ecology and Dynamics – Chair: P. Tango (Lillie Auditorium)**

10:30	Don Anderson	21	<i>Alexandrium fundyense</i> cyst dynamics in the Gulf of Maine
10:50	Deana Erdner	37	Population genetics of toxic <i>Alexandrium</i> blooms in the Gulf of Maine
11:10	Dennis McGillicuddy	58	Observations and models of <i>Alexandrium fundyense</i> blooms in the Gulf of Maine and Georges Bank: From climatology to forecasting
11:30	Mario Sengco	70	The fate of saxitoxins in <i>Alexandrium tamarense</i> during infection by <i>Amoebophrya</i> sp., and initial observation of host-parasite dynamics from field studies in a small Cape Cod embayment

Lunch 12:00 – 1:30**Session 15: Human Health – Chair: D. Erdner (Lillie Auditorium)**

1:30	Tracy Villareal	76	Ciguatotoxicity in the northern Gulf of Mexico
1:50	Bob Dickey	33	Formulation of advisory levels for Caribbean and Pacific ciguatoxins and tiered methods for their determination
2:10	Andy Reich	66	Features of neurotoxic shellfish poisoning from recreationally harvested clams in Florida, 2006: Epidemiologic and clinical factors

Break 2:30 – 3:00**Session 16: Bloom Ecology and Dynamics – Chair: P. Strutton (Lillie Auditorium)**

3:00	Sibel Bargu	24	<i>Pseudo-nitzschia</i> and domoic acid in naturally iron-enriched and iron-poor areas of the Gulf of Alaska
3:20	Lisa Pickell	61	Dissolved domoic acid: A competitive advantage for <i>Pseudo-nitzschia</i> in coastal and offshore HNLC waters
3:40	Nick Adams	19	The use of microsatellite markers to compare the population structure of <i>Pseudo-nitzschia pungens</i> from the Pacific Northwest and the North Sea
4:00	Astrid Schnetzer	69	Toxic blooms of <i>Pseudo-nitzschia</i> spp. and their impact on coastal marine life in the Southern California bight area near Los Angeles
4:20	Don Anderson, Quay Dortch & Pat Glibert		Community Meeting – HABHRCA update; NHC update; selection of next site

Dinner 6:00 – 7:30 (Swope Dining Hall)**Mixer - 7:30 – 8:30**

BLOOM ECOLOGY AND DYNAMICS – POSTERS

*(odd-numbered posters will be presented in Poster Session 1, Monday, October 29th, 4 - 6 pm;
even-numbered posters will be presented in Poster Session 2, Monday, October 29th, 7:30 – 9:30 pm)*

First Author (presenting author, if different)	Title	Poster #
Anderson, Jon	A fuzzy logic approach to predicting <i>Prorocentrum minimum</i> blooms in the Chesapeake Bay	B-1
Auro, Maureen	Growth, toxicity and nitrogen uptake capabilities of the toxigenic diatom <i>Pseudo-nitzschia cuspidatae</i> from the Pacific Northwest	B-3
Beall, Benjamin	Low sinking rates of <i>Pseudo-nitzschia</i> : A competitive feature contributing to the development and maintenance of toxic blooms	B-5
Bill, Brian	Puget Sound, Washington: An emerging hotspot for <i>Pseudo-nitzschia</i> blooms and domoic acid toxic events	B-7
Borkman, David	Long-term (1992-2006) patterns in <i>Phaeocystis</i> bloom magnitude and bloom duration in Massachusetts Bay, USA	B-9
Brosnahan, Michael	Evidence of a self-recognition system in the sexual life cycle of <i>Alexandrium tamarense</i>	B-11
Burson, Amanda	The nutritional ecology of the harmful dinoflagellate blooms caused by <i>Cochlodinium polykrikoides</i> in Long Island bays (NY, USA)	B-13
Carter, Melissa	Coastal bloom dynamics in southern California - Have there been any changes since 1917?	B-17
Errera, Reagan	Pigment composition of the Texas strain of <i>Prymnesium parvum</i> during log, stationary, and senescent growth phases	B-19
Goleski, Jennifer	Harmful cyanobacterial bloom proliferation in Florida Bay, FL, USA: Zooplankton grazing and the role of salinity	B-21
Griffith, Jennifer	Development and validation of a novel in vivo nitrate reductase activity assay	B-23
Hall, Emily	Distribution of nutrient data in relation to <i>Karenia brevis</i> cell counts along the west central Florida coast	B-25
Hattenrath, Theresa	Growth and toxicity of <i>Alexandrium sp.</i> on the north shore of Long Island, NY, USA: Dynamics and interactions with nutrients	B-27
Hayashi, Kendra	Long-term, temporal variability in <i>Pseudo-nitzschia</i> population dynamics and domoic acid toxicity in Monterey Bay, CA	B-29
Henrichs, Darren	Evaluating genetic diversity of <i>Karenia brevis</i> blooms in the western Gulf of Mexico	B-31
Horner, Rita	<i>Alexandrium catenella</i> cysts and environmental conditions in Puget Sound, WA: Results of a cyst survey	B-33
Hubert, Jeff (Greengrove, Cheryl)	Do sediment conditions affect the incidence of <i>Alexandrium catenella</i> and paralytic shellfish poisoning? A study of sites from Puget Sound	B-35
Johns, Desmond	Characterization of nitrogen uptake by <i>Heterosigma akashiwo</i> grown in turbidostat culture under two light intensities	B-2
Kulis, David	A mesocosm study examining the influence of nutrients on <i>Alexandrium tamarense/fundyense</i> toxin concentration and composition	B-4

First Author (presenting author, if different)	Title	Poster #
Laine, Edward	Sediments and cysts on the floor of Harpswell Sound: Is there a local cyst bed for the high toxicities in Lombos Hole?	B-6
Lehman, Peggy	The influence of environmental conditions on the seasonal variation of <i>Microcystis aeruginosa</i> cell density and microcystins concentration in San Francisco Estuary	B-8
Lekan, Danelle	Influence of temperature, salinity and nutrient ratios on toxin profiles of <i>Karenia brevis</i>	B-10
Lovko, Vincent	Pathogenicity, prey availability and functional types in <i>Pfiesteria</i> and <i>Pfiesteria</i> -like species: The role of micropredatory feeding	B-12
Mangum, Alicia	The effect of carbon dioxide (CO ₂) on grazing activities for <i>Karlodinium veneficum</i>	B-14
Mazzillo, Fernanda	Spatial distribution of phytoplankton groups and toxic species in a nearshore frontal zone system in Monterey Bay, California	B-16
McCauley, Linda (Libera, Katie)	A study of <i>Pseudo-nitzschia</i> in the Gulf of Maine: Diversity and toxicity	B-18
Millie, David	Bloom-forming phytoplankton in Saginaw Bay (Lake Huron) and western Lake Erie: Abundance, distribution, and cyanobacterial toxicity during late summer	B-20
Myers, Tracey	Gulf of Mexico phytoplankton affect fate of red tide toxin	B-22
Novoveska, Lucie	Seasonal and inter-annual changes in dinoflagellates community composition in nearshore Alabama waters	B-24
Peña, Angelica (Callendar, Wendy)	Biophysical modeling of the Juan de Fuca eddy in the Pacific Northwest	B-15
Radan, Regina	Ammonium surge uptake and inhibition of nitrate uptake by small and large cell-sized <i>Pseudo-nitzschia</i> species from the Pacific Northwest	B-26
Richlen, Mindy	Ecology and biodiversity of toxic benthic dinoflagellates at Johnston Atoll, Pacific Ocean	B-28
Schoener, Donald	Grazing, growth, and behavioral reactions of a ciliate fed <i>Alexandrium</i> spp: apparent lack of response to saxitoxin	B-30
Senft, Christina	Phenotypic variations in ingestion of toxic algae within populations of a marine copepod	B-32
Silver, Mary	Domoic acid in oceanic <i>Pseudo-nitzschia</i> : Is it an issue?	B-34
Skelton, Hayley	Axenic cultivation of the heterotrophic dinoflagellate <i>Pfiesteria shumwayae</i> on a semi-defined medium	B-36
Stauffer, Beth	Small scale bloom dynamics of raphidophyte and dinoflagellate populations in King Harbor, California	B-38
Strutton, Pete	New satellite data products for the detection of Oregon HABs	B-40
Tang, Ying Zhong	Characterization, dynamics, and ecological impacts of harmful <i>Cochlodinium polykrikoides</i> blooms on eastern Long Island, NY, USA	B-44
Tango, Peter	Three year assessment of CyanoHAB forecasting on the tidewaters of the Potomac River, Chesapeake Bay, USA	B-46
Teegarden, Gregory	The importance of surface currents in high PSP toxicity in Lombos Hole, Harpswell Sound, Maine	B-48

First Author (presenting author, if different)	Title	Poster #
Teegarden, Gregory	The importance of surface currents in high PSP toxicity in Lumbos Hole, Harpswell Sound, Maine	B-48
Tobin, Elizabeth	Germination of <i>Alexandrium catenella</i> cysts from surface sediments in Quartermaster Harbor, WA	B-37
Trick, Charlie (Wells, Mark)	<i>Pseudo-nitzschia</i> growth and toxin production in the Juan De Fuca eddy in the Pacific Northwest – Environmental stimulators of a toxic bloom	B-50
Tweddle, Jacqui	The relationship between coastal ocean dynamics and shellfish closures: A satellite based study of Oregon HABs	B-42
Wang, Bin	Effect of cadmium, copper, nickel, and zinc on a minute golden brown alga <i>Aureococcus anophagefferens</i>	B-52
Wei, Liping	Growth response and glutathione production of brown tide bloom alga (<i>Aureococcus anophagefferens</i> , CCMP 1984) upon different salinity, metals, nitrogen source, sewage and herbicide metolachlor exposure: lab culture studies and in situ incubation studies	B-54
Wrabel, Michele	Specificity of bacterial assemblages associated with the toxin-producing diatom, <i>Pseudo-nitzschia</i>	B-56
Wurch, Louie	Nutrient-regulated transcriptome profiling in the brown-tide forming alga <i>Aureococcus anophagefferens</i>	B-58

COMMUNICATION, OUTREACH, AND EDUCATION – POSTERS
(Poster Session 1, Monday, October 29th, 4 - 6 pm)

First Author (presenting author, if different)	Title	Poster #
Ammerman, James	Applications of the ocean biogeographic information system (OBIS) to HAB data	C-1
Cross, Scott (Morton, Steve)	Data management supporting regional volunteer phytoplankton monitoring efforts	C-2
Day, Sheryl	The Pacific Northwest HAB Bulletin pilot project: Technical development of an ocean observing information system for the protection of human health	C-3
Haley, Sheean	Volunteer-driven science: mapping harmful algal blooms in Puget Sound, Washington	C-4
Morton, Steve	Identification of <i>Pseudo-nitzschia</i> and domoic acid from a North Carolina coastal bloom: Linkage between volunteer observations and biotoxins research	C-5
Trainer, Vera (Muir, Christine)	ECOHAB Pacific Northwest (ECOHAB PNW) Outreach: Opening the scientific journey to the world	C-6
Zimmermann, Leigh	Florida's red tide control & mitigation grant program: The beginning	C-7

HUMAN HEALTH – POSTERS
 (Poster Session 3, Tuesday, October 30th, 4 - 6 pm)

First Author (presenting author, if different)	Title	Poster #
Calkins, Julie (Tomlinson, Michelle)	Using a novel low-cost monitoring array for validation of brevetoxin-induced respiratory impact in Southwest Florida	H-1
Gold, Elena	Potential good in a “SEA” of harm: Characterization of a mammalian receptor associated with voltage-sensitive sodium channels using brevenal produced by <i>Karenia brevis</i>	H-2
Kirkpatrick, Barbara	Ocean observing systems and public health: The Florida Beach conditions reporting system to minimize exposure to <i>Karenia brevis</i> aerosols	H-3
LePrell, Rebecca	Development and initiation of HABISS; CDC’s Multi-state HAB surveillance system	H-4
Margot, Kelli	Immunological response of distal lung cell lines to brevetoxins	H-5
Naar, Jerome	The mitigating properties of cysteine on the harmful effects of red tide	H-6
Nierenberg, Kate	Reported respiratory symptom intensity in asthmatics during exposure to aerosolized Florida red tide toxins	H-7
Polansky, Lara (Hoagland, Porter)	Determining the role of <i>Karenia brevis</i> blooms in emergency room visits due to respiratory ailments in Sarasota, Florida	H-8
Shehee, Mina	North Carolina harmful algal bloom events 2005 – 2007	H-9
Tomlinson, Michelle	The development of a beach impact model for Florida <i>Karenia brevis</i> blooms	H-10

TOXINS AND TOXIN DETECTION – POSTERS
(Poster Session 3, Tuesday, October 30th, 4 - 6 pm)

First Author <i>(presenting author, if different)</i>	Title	Poster #
Bottein, Marie-Yasmine	Evaluation of short and long lasting neurological response to single and repeated ciguatoxin exposure in mice	T-1
Fahnenstiel, Gary	Factors affecting microcystin concentrations and cell quotas in the Great Lakes	T-2
Hotto, Amber	Relationship between microcystin potential and environmental variables in lakes across New York state	T-3
Lane, Jenny	Degradation of domoic acid under common storage conditions	T-4
Lee, Jungju	Advanced treatment processes for the removal of cyanotoxins from Lake Erie drinking water	T-5
Mooney, Ben	Distribution of karlotoxins among Australian and North American ichthyotoxic Gymnodinoid dinoflagellates (Kareniaceae)	T-6
Pierce, Richard	Accumulation and depuration of brevetoxins and major metabolites in shellfish exposed to recurring <i>Karenia brevis</i> blooms	T-7
Ramsdell, John	Diel synchronization of embryonic diapause aids prediction of fetal toxicity of California sea lions to domoic acid-producing harmful algal blooms	T-8
Schultz, Irvin	Domoic acid uptake and excretion in fish, Dungeness crabs, razor clams and mussels	T-9
Sinclair, James	Survey of algal toxins in source and finished drinking waters	T-10
Smith, Juliette	Development of an internal standard for the measurement of free microcystins in fish tissue and sediments	T-11
Stewart, Thomas	Rapid enzyme-linked immunosorbent assay for the detection of domoic acid	T-12
Sutherland, Cristy	<i>Dinophysis</i> species and diarrhetic shellfish toxins in Monterey Bay, CA	T-13
Tatters, Avery	Hemolytic activity of selected HAB flagellates: A toxin complex strategy	T-14
Thessen, Anne	The effect of salinity on domoic acid production by the diatom <i>Pseudo-nitzschia multiseriis</i>	T-15
White, Kristy	Studies on the enzymatic hydrolysis of DSP esters to produce the toxic, okadaic acid, in three strains of <i>Prorocentrum lima</i>	T-16
Yang, Xingye	Comparative reactivity of different cysteine congeners as detoxifying agents of brevetoxins	T-17

EMERGING TECHNOLOGIES – POSTERS
(Poster Session 4, Wednesday, October 31st, 4 - 6 pm)

First Author (presenting author, if different)	Title	Poster #
Bhattacharya, Debashish and Moustafa, Ahmed	Phylosort: A Program for detecting endosymbiotic and horizontal gene transfer and its application to understanding STX gene origin in <i>Alexandrium tamarense</i>	E-1
Bowers, Holly	Using Real-time PCR to demonstrate co-occurrence of <i>Karlodinium veneficum</i> and the parasitic dinoflagellate <i>Amoebophyra</i> sp. ex. <i>Karlodinium veneficum</i>	E-2
Campbell, Lisa	Genetic variation among isolates of <i>Karenia brevis</i> as measured with microsatellite markers	E-3
Carvalho, Gustavo	Long-term evaluation of a satellite ocean color dataset to detect blooms of the toxic dinoflagellate <i>Karenia brevis</i> off the west Florida shelf	E-4
Chrest, Francis and Daniel Terlizzi	Measurement of apoptosis in phytoplankton using flow cytometry and Annexin V binding	E-5
Dash, Padmanava	Quantitative mapping of cyanobacterial blooms from Oceansat-1 OCM satellite data: An empirical approach	E-6
Donovan, Chelsea (Younan, Lawrence)	New performance data for in situ and simulated experiments using the phytoflash submersible active fluorometer	E-7
Duy, Janice	Fast and accurate detection of <i>Alexandrium</i> species using peptide nucleic acid probes and surface plasmon resonance	E-8
Fisher, Kathleen	Assessment of an operational harmful algal bloom forecast system for the eastern Gulf of Mexico: A comparative analysis of success and utilization through two bloom seasons	E-9
Freitag, Michael	Coupling physiological responses of the toxic haptophyte <i>Prymnesium parvum</i> to patterns in gene expression	E-11
Gray, Cynthia	Glycolipid families in peridinin-containing dinoflagellates	E-12
Kelly, Vince (Glibert, Pat)	<i>In situ</i> nutrient monitoring: An example of research, development, demonstration and technology transfer	E-13
Langer Atkinson, Heidi	Assessment of microcystin production in cyanobacteria using a novel whole cell fluorescent immunologicalization method, flow cytometry and the enzyme-linked immunosorbant assay (ELISA)	E-10
Love, Rebecca	Integrating novel data sources to improve the Gulf of Mexico harmful algal bloom forecast system	E-14
Miller, Peter	California program for regional enhanced monitoring of phycotoxins (Cal-PReEMPT)	E-15
Miranda, Lilibeth	<i>Alexandrium fundyense</i> cDNA microarray: The hunt for growth-related genes	E-16
Truxal, Laura	Characterization of novel compounds found in <i>Karenia brevis</i> cultures	E-17
Walker, Elyse	Identification and enumeration of <i>Pseudo-nitzschia</i> in Pacific Northwest coastal waters using the Flowcam® continuous imaging particle analyzer	E-18
Wynne, Timothy (Stumpf, Rick & Tomlinson, Shelly)	Detecting cyanobacteria blooms using MERIS	E-19

FISHERIES AND FOOD WEBS – POSTERS
(Poster Session 4, Wednesday, October 31st, 4 - 6 pm)

First Author (presenting author, if different)	Title	Poster #
Coblentz, Francie	Single laboratory validation of the brevetoxins ELISA for shellfish contamination assessment	F-1
Connell, Laurie	Population structure of Paralytic Shellfish Poisoning (PSP) resistant softshell clams, <i>Mya arenaria</i> , in eastern Maine, USA	F-2
Fire, Spencer	Unusually high levels of domoic acid identified in Minke whale (<i>Balaenoptera acutorostrata</i>) stranding during California <i>Pseudo-nitzschia</i> bloom	F-3
Ford, Susan (Bricelj, Monica)	Deleterious effects of a non-PSP bioactive produced by <i>Alexandrium tamarense</i> on bivalve hemocytes	F-4
Garcia, Ana	Bioaccumulation of cyanobacterial <i>Cylindrospermopsis</i> toxin in Louisiana blue crab, <i>Callinectes sapidus</i>	F-5
Garcia, Suzanne	Domoic acid in benthic communities of the Santa Cruz municipal wharf in Monterey Bay, California	F-6
Giner, José	Metabolism of algal sterols by bay scallops and brine shrimp	F-7
Hamilton, Scott	New techniques for non-lethal DNA Extraction from, and Passive Integrated Transponder (PIT) tagging of, the soft-shell clam <i>Mya arenaria</i>	F-8
Jin, Di	The value of HAB predictions to the commercial shellfish industries in the Gulf of Maine	F-9
Landsberg, Jan	The emerging risks of cyanobacteria for fish and wildlife in Florida	F-10
Shumway, Sandra	Mitigating the risk of introduction of harmful algae via transfer of bivalve mollusks	F-11
Wikfors, Gary	Variable expression of toxicity in <i>Prorocentrum minimum</i> , and possible relationships with trophic status	F-12

ABSTRACTS OF ORAL PRESENTATIONS

THE USE OF MICROSATELLITE MARKERS TO COMPARE THE POPULATION STRUCTURE OF *Pseudo-nitzschia pungens* FROM THE PACIFIC NORTHWEST AND THE NORTH SEA

Nicolaus G. Adams¹, Lorenz Hauser², Russell P. Herwig², Gabrielle Rocap³, and Vera L. Trainer¹

¹NOAA-Fisheries, Northwest Fisheries Science Center, Seattle, WA, 98112, USA

²University of Washington, School of Aquatic and Fishery Sciences, Seattle, WA, 98195, USA

³University of Washington, School of Oceanography, Seattle, WA, 98195, USA

Pseudo-nitzschia pungens is a pennate diatom commonly found in the waters of the Pacific Northwest and the North Sea. Due to the frequency of domoic acid events in the Pacific Northwest an understanding of the population and bloom dynamics of *Pseudo-nitzschia* species in this region is needed. Our research compared various population genetic parameters of *P. pungens* isolated from the Juan de Fuca eddy region in the Pacific Northwest and from the North Sea. These data along with population differentiation statistics indicated that *P. pungens* from the Pacific Northwest had a different population structure than *P. pungens* from the North Sea. Genetic data from a North Sea *P. pungens* sample provided evidence for the presence of a single unstructured population while a more complex population structure was found in Pacific Northwest *P. pungens* samples. Microsatellite data indicated that two genetically distinct populations were present in all Pacific Northwest *P. pungens* samples. These results implied that either the two populations of *P. pungens* could have recently mixed in the Juan de Fuca eddy region but had not exchanged genetic material by sexual reproduction, or that there may be cryptic species (morphologically identical but reproductively isolated species). The detection of multiple populations or cryptic species of a potentially toxic diatom suggests a more complex cause of HABs in the Pacific Northwest than was hitherto assumed, and calls for additional studies investigating physiological and genetic differentiation between the two strains.

CRYPTOPHYTES IN CHESAPEAKE BAY AND THEIR POTENTIAL RELATIONSHIP TO MIXOTROPHIC HARMFUL ALGAL BLOOMS

Jason E. Adolf¹, Holly A. Bowers¹, Tsvetan R. Bachvaroff¹, and Allen R. Place¹

¹UMBI Center of Marine Biotechnology, 701 E. Pratt St., Baltimore, MD USA 21202

Cryptophytes represent a substantial portion of phytoplankton biomass in eutrophic coastal estuaries such as Chesapeake Bay, but their ecology is poorly understood due in part to difficulty in identifying species by light microscopy. We hypothesize that cryptophyte abundance drives the formation of mixotrophic harmful algal blooms (HABs) in eutrophic environments. *Karlodinium veneficum* grows 2-3 fold faster when feeding on cryptophytes and produces an allelochemical, karlotoxin (KmTX), that allows it to effectively compete for cryptophytes, even in the presence of another heterotrophic predator, *Oxyrrhis marina*. *Karlodinium veneficum* feeds on a broad array of cryptophyte species, but other mixotrophic bloom formers such as *Myrionecta rubra* (Gustafson et al. 2000) and *Dinophysis acuminata* (Nishitani et al. 2005), show a high degree of specificity in prey selection. We have begun to investigate the *in situ* diversity of cryptophytes in Chesapeake Bay by generating clone libraries from sub-populations of phytoplankton sorted by flow cytometry to enrich for phycoerythrin-containing cryptophytes. Preliminary results from one sample collected during spring 2007 from the Patapsco River, MD showed a high abundance of *Amoebophrya* (Dinophyceae) sequences and one sequence identified as *Teleaulax acuta* based on a BLAST search. This survey will result in the development of molecular tools for identifying and tracking cryptophyte species *in situ* and will play a role in a broader project aimed at improving our ability to predict mixotrophic HABs, including attempting to forecast *K. veneficum* blooms by using phycoerythrin fluorometers on Maryland Department of Natural Resources real-time continuous monitoring platforms to detect potential prey populations in locations where *K. veneficum* blooms have occurred annually for the last two years (Corsica R., MD).

GUSTAFSON JR., D. E., STOECKER, D.K., JOHNSON, M.D., VAN HEUKELEM, W.F., AND SNEIDER, K., 2000. Cryptophyte algae are robbed of their organelles by the marine ciliate *Mesodinium rubrum*. *Nature* **405**: 1049-1052.

NISHITANI, G., YAMAGUCHI, M., ISHIKAWA, A., YANAGIYA, S., MITSUYA, T., IMAI, I. 2005. Relationships between occurrences of toxic *Dinophysis* species (Dinophyceae) and small phytoplankton in Japanese coastal waters. *Harmful Algae* **4**: 755-762.

***Alexandrium fundyense* CYST DYNAMICS IN THE GULF OF MAINE**

Donald M. Anderson¹, Bruce A. Keafer¹, Kerry Norton¹, Dennis J. McGillicuddy¹, Ruoying He², Cynthia H. Pilskaln³, and Darcie Couture⁴

¹Woods Hole Oceanographic Institution, Woods Hole, MA

²North Carolina State University, Raleigh NC

³University of Massachusetts, Dartmouth, MA

⁴Maine Department of Marine Resources, W. Boothbay Harbor, ME

Resting cysts play important roles in bloom initiation, termination, and species dispersal for several important HAB species. Several studies have examined linkages between cyst abundance and bloom timing and location, but these efforts have typically been over relatively small areas due to the difficulties and expense of mapping cysts over larger regions. Here we present the results of four cyst mapping surveys for *Alexandrium fundyense* in the Gulf of Maine, each covering hundreds of km in the alongshore direction, and 50 – 100 km in the offshore. The first survey (in 1997 (augmented with Bay of Fundy data from 1982, 1983 & 1984) revealed a widespread cyst distribution, with two centers of abundance, one in the Bay of Fundy, and a second offshore of Casco and Penobscot Bays in western Maine. The second survey (2004) shows a much larger cyst population, with approximately 10x as many cysts in the western Maine seedbed area. This was the cyst distribution that existed prior to, and undoubtedly contributed to, a massive 2005 *A. fundyense* bloom in southern New England. The third survey was conducted after the 2005 bloom, and reveals levels of cysts in western Maine that are ~5X the 1997 levels, but ½ of the 2004 levels. No significant geographic expansion of cysts into southern waters was seen, as had been feared based on the high motile cell concentrations that occurred in those waters during the 2005 bloom. The 2006 cyst map showed similar patterns to other years, with approximately 30% fewer cysts than observed in 2005. The overall pattern is thus one of large interannual variability, with a gradual decrease in cyst abundance in recent years. This presentation will discuss these interannual changes in the context of the blooms that occurred before and after the mapping efforts and in the temporal trends in PSP toxicity in the region over the last several decades as well. Newly acquired data on cyst deposition fluxes from sediment traps, and the abundance of resuspended cysts in the benthic nepheloid layer will also be discussed.

THE EVOLUTION OF TOXIN RESISTANCE IN COPEPODS: HOW DO COPEPODS RESPOND TO BLOOMS OF TOXIC *Alexandrium fundyense*?

David E. Avery^{1,2}, Hans G. Dam¹, Kristajoy Altland^{1,3}

¹. Department of Marine Sciences, University of Connecticut, Groton, CT 06340-6048, USA

². davery@uconn.edu

³. Current address: Dept. Ecol. Evol. Biology, Yale University, New Haven, CT, USA

We have been using the calanoid copepod *Acartia hudsonica* and the dinoflagellate *Alexandrium fundyense* as a model system to study the interactions of planktonic grazers and toxic algae. The foundation for these studies was laid when it was discovered that some copepods appeared to be resistant to toxins while others were susceptible (Colin and Dam 2002, 2004). Since then, we have made several advances understanding the mechanism and the consequences of resistance. First, life history experiments revealed discrete reproductive phenotypes related to resistance (Avery and Dam 2007). When we used fitness as an index of resistance, it was apparent that the most resistant copepods were not extreme members of the population with respect to various characters, but middling members. As a result, the discrete phenotypes have been hypothesized to represent a simple genetic system of one or two genes showing heterozygote advantage. An immediate consequence is that heterozygote advantage would prevent the fixation of resistance alleles in a population so it appears that no population would become fixed for resistance. This hypothesis is supported by results of experiments lasting three generations, in which resistant and nonresistant alleles would appear to alternate through generations due to segregation and selection. Hence, there is no guarantee that evolution of resistance would necessarily lead to bloom control. If our model applies to other copepods co-occurring with *Alexandrium* sp., then we can expect measures of population growth rates, ingestion rates, and egg production rates to reflect this polymorphism for resistance.

References:

- Colin, S.P. and H.G. Dam. 2002. Latitudinal differentiation in the effects of the toxic dinoflagellate *Alexandrium* spp. on the feeding and reproduction of populations of the copepod *Acartia hudsonica*. Harmful Algae. 1: 113-125.
- Colin, S.P. and H.G. Dam. 2004. Testing for resistance of pelagic marine copepods to a toxic dinoflagellate. Evol. Ecol. 18: 355-377.
- Avery, D.E. and H.G. Dam. 2007. Newly discovered reproductive phenotypes of a marine copepod reveal the costs and advantages of resistance to a toxic dinoflagellate. Limnol. Oceanogr. In press,

RECREATIONAL EXPOSURE TO MICROCYSTINS DURING A *Microcystis aeruginosa* BLOOM IN A SMALL LAKE

Lorraine C. Backer¹, Wayne Carmichael², Barbara Kirkpatrick³, Christopher Williams⁴, Mitch Irvin⁵, Yue Zhou⁵, Trisha B. Johnson⁶, Kate Nierenberg³, Vincent R. Hill⁶, Stephanie M. Kieszak¹, and Yung-Sung Cheng⁵.

¹National Center for Environmental Health, Centers for Disease Control and Prevention, Chamblee, Georgia 30341, ²Wright State University, Dayton, Ohio 45325, ³Mote Marine Laboratory, Sarasota, Florida 34236, ⁴Greenwater Laboratories, Pakatka, Florida 32177, ⁵Lovelace Respiratory Research Institute, Albuquerque, New Mexico 87285, ⁶Center for Infectious Diseases, Centers for Disease Control and Prevention.

Evidence of adverse human health events from exposure to freshwater cyanobacterial blooms is primarily anecdotal. Our objective was to document recreational exposure by measuring microcystins in blood samples from people at risk for swallowing water or inhaling spray (e.g., water skiers, jet skiers) during a *Microcystis aeruginosa* bloom. We found very low levels of microcystins (1 µg/L to 6 µg/L) in the water. Blood levels of microcystins were below the limit of detection (0.147 µg/L) for 97 people who did recreational activities on the lake with the *M. aeruginosa* bloom (exposed participants) and 7 people who did recreational activities on a different lake with no algal bloom (unexposed participants). Using technology developed for the assessment of aerosols containing brevetoxins, we detected microcystins (<0.1 ng/m³) in aerosol samples (both personal samplers and ambient environmental samplers), indicating that recreational activities are potential sources of exposure to these potent toxins. We plan to conduct a similar study in a lake where microcystin concentrations are at least 20 µg/L to assess whether we can detect the toxin in blood samples from people with higher exposures.

***Pseudo-nitzschia* AND DOMOIC ACID IN NATURALLY IRON-ENRICHED AND IRON-POOR AREAS OF THE GULF OF ALASKA**Sibel Bargu¹ and Mary Silver²¹Department of Oceanography and Coastal Sciences, Louisiana State University, 1235 Energy, Coast & Environment Building, Baton Rouge, Louisiana 70803, USA²Department of Ocean Sciences, University of California at Santa Cruz, 1156 High Street, Santa Cruz, California 95064, USA

Although a number of cosmopolitan species of *Pseudo-nitzschia* are now known to produce the neurotoxin domoic acid (DA), very little is known about the ability of the truly oceanic species to produce DA *in situ*. There is considerable literature available to show that *Pseudo-nitzschia* are a common component of subarctic and subtropical oceanic systems, including ones in iron-limited HNLC (high nutrient, low iron) regions of the world ocean, as reviewed in the first talk in this 2-part series. With recent concerns about global warming and discussions of iron enrichment of HNLC regions of the oceans to remedy the warming, the possibility of promoting the growth of a known toxin producing genus needs to be further explored. This presentation describes some new data we acquired on a recent expedition to the Gulf of Alaska, which includes an HNLC area known to contain members of this pennate diatom genus.

Here we present data from the northern Gulf of Alaska, an oceanic system that has both iron limited and naturally iron enriched (from major eddies that contain river discharge) regions, obtained on a cruise devoted to the study of the various forms of iron and their concentrations in seawater. Our studies in the Gulf of Alaska are focused on determining whether the naturally occurring *Pseudo-nitzschia* in this area are producing DA under natural, *in situ* conditions both in the iron rich and iron poor (HNLC) areas. We will present some data on the cellular DA quotas in the field relative to various forms of iron in those waters. If successful, we will also present results from iron fertilization experiments conducted aboard ship during grow-out experiments. Field data will come from stations along north-south transects in the Alaska region where *Pseudo-nitzschia* cells are sufficiently abundant to measure DA using highly-sensitive enzyme-linked immunosorbent assays (ELISA), with corresponding cell numbers being obtained by epifluorescence microscopy aboard ship. Species of *Pseudo-nitzschia* present in the field and in grow-out experiments are being identified using suitably fixed or prepared materials from stations and grow-out experiments. Species designations will rely on SEM and TEM analysis for morphological discrimination of the species in the Gulf of Alaska. This type of field analysis of *Pseudo-nitzschia* populations, we suggest, is needed to determine whether local populations of oceanic *Pseudo-nitzschia* are already producing domoic acid in HNLC areas and whether iron fertilization, as proposed as a solution to global warming, may not only produce phytoplankton blooms but also may inadvertently increase the abundance of neurotoxin-producing species of *Pseudo-nitzschia*.

USING SOCIAL SCIENCE TO DEVELOP COMMUNICATION MESSAGES THAT FACILITATE PUBLIC TRUST AND UNDERSTANDING REGARDING HARMFUL ALGAL BLOOM CONTROL

Marybeth Bauer¹, Charles M. Adams², Linda L. Lampl³, Sherry L. Larkin², Chris Pettit⁴, Clifford W. Scherer⁵, Mario R. Sengco⁶, John M. Stevely², Patricia A. Tester⁷

¹NOAA, National Ocean Service, National Centers for Coastal Ocean Science, Silver Spring, MD 20910

²University of Florida, Gainesville, FL 32611

³Lampl-Herbert Consultants, Tallahassee, FL 32302

⁴New College of Florida, Sarasota, FL 34243

⁵Cornell University, Ithaca, NY 14853

⁶Smithsonian Environmental Research Center, Edgewater, MD 21037

⁷NOAA, National Ocean Service, National Centers for Coastal Ocean Science, Center for Coastal Fisheries and Habitat Research, Beaufort, NC 28516

Harmful algal bloom (HAB) control encompasses mechanical, biological, chemical, genetic, and environmental strategies that kill HAB organisms, limit their growth and proliferation, or remove them from the water column. Despite studies suggesting that clay dispersal is a particularly promising method for controlling HABs (Sengco, 2001; Sengco and Anderson, 2004), public attitudes toward the strategy range from reasonably doubtful to strongly negative. At a 2006 public forum on red tide research in the Gulf of Mexico, for example, stakeholders voiced concern that clay dispersal (a) introduces an “unnatural” substance into coastal waters and (b) may have unforeseen environmental and public health impacts that are worse than the blooms themselves.

In many ways, societal decisions and public reactions will influence whether and how promising research on HAB control will move beyond an experimental scale to field demonstration and implementation. For example, societal decisions weighing the risks and expected benefits of alternative control options will influence the level of funding investment and design of permitting processes. In addition, public outcry stemming from misperceptions and mistrust could directly impede field demonstration and implementation. Public trust, understanding, and informed participation in decisionmaking are critical to ensure the responsible development and effective implementation of HAB control. Public education and outreach are critical toward this end.

A report by the National Science and Technology Council, *Grand Challenges for Disaster Reduction*, emphasizes that social science research is needed to design communication messages appropriate for public education and outreach. Specifically, designing effective communications requires an understanding of diverse stakeholder audiences – e.g., their knowledge of HABs, concerns about HAB control, perceptions of risks, and level of trust in scientists and decisionmakers. It also requires understanding and improvement of the institutional capacity of the HAB science-management community to communicate with stakeholders (e.g., interagency communications and media relations). Systematic study of public audiences and institutional capacity is needed to design effective communication messages and strategies related to HAB control.

This presentation reports on a nine-member interdisciplinary project to develop communication messages and strategies that help resource managers (e.g., Florida Fish and Wildlife Conservation Commission), citizens groups (e.g., Solutions to Avoid Red Tide), scientists, and others foster public trust, understanding, and meaningful participation in decisionmaking regarding HAB control in Florida. Communication design will be informed by social science research (e.g. interviews and surveys) to better understand stakeholders. The project will involve HAB scientists and managers, and investigate the impact of the research process on their knowledge, attitudes, and perceptions related to the human dimensions of coastal management.

- Sengco, M.R. 2001. The aggregation of clay minerals and marine microalgal cells: Physicochemical theory and implications for controlling harmful algal blooms. Ph.D. Dissertation, Massachusetts Institute of Technology/Woods Hole Oceanographic Institution. 237 pp.
- Sengco, M.R. and D.M. Anderson. 2004. Controlling harmful algal blooms through clay flocculation. *J. Eukaryot. Microbiol.* 51(2): 169-172.

APPLICATIONS OF VIDEO-ENDOSCOPY TO STUDY THE EFFECTS OF HAB SPECIES ON SUSPENSION-FEEDING BIVALVES

V. Monica Bricelj¹, J. Evan Ward², Heather Robbins^{1,3}, Luiz Mafra^{1,3}

¹National Research Council, Institute for Marine Biosciences, NS B3H 3Z1, Canada; ²Department of Marine Sciences, University of Connecticut, Groton, CT 06340, USA; ³Biology Department, Dalhousie University, Halifax, NS B3H 4H6, Canada

Bivalve suspension-feeding is a complex process involving particle capture and ciliary transport on the gills, and pre-ingestive particle selection at the gills and/or labial palps via rejection of pseudofeces. *In vivo* observations of bivalve feeding, using video-endoscopy, have been used in the past decade to elucidate mechanisms of particle processing in adult bivalves (Ward et al., 1998), but have rarely been applied to study the effects of harmful algae (Bricelj et al., 1998). This work demonstrated a novel mechanism of particle rejection of toxic *Alexandrium tamarense* cells by oysters which did not result in the formation of consolidated pseudofeces, but rather the loss of free cells from the gill dorsal tract, typically considered an acceptance tract. The present study extends the work of Bricelj et al. (1998) by applying video-endoscopy to study the effects of two non-dinoflagellate HAB species: *Aureococcus anophagefferens*, the causative agent of brown tide (BT), and *Pseudo-nitzschia multiseriis*, a domoic acid (DA)-producer.

Hard clams, *Mercenaria mercenaria*, were exposed to non-toxic algae, *Isochrysis galbana*, and then supplemented with bloom levels of either a toxic or non-toxic isolate of *A. anophagefferens* for 2h. Our results indicate that clams exposed to the toxic BT (in contrast to the non-toxic BT) showed intermittent cessation and reversal of the inhalant flow above the frontal surfaces of the gills, and intermittent convulsions/twitching of the gills within ~4-60 min of exposure. Such effects are likely due to inhibition and lack of coordination of the gill lateral cilia, which create the inhalant feeding current, as well as effects on the gill musculature. During exposure to toxic BT, while major disturbance effects were occurring at the level of the gills, clams showed fully extended siphons with dilated apertures and extended guard tentacles, generally associated with active feeding. Therefore, the siphon aperture cannot be used as a reliable index of feeding activity. Our results support *in vitro* observations of dissected gills in which toxic BT caused dopamine-mimetic inhibition of lateral ciliary activity (Gainey and Shumway, 1991), yet suggest a more complex mechanism of action whereby BT also caused convulsions of gill musculature that will contribute to reduced feeding efficiency. The *in vivo* response was more rapid than that reported *in vitro*, as ~7h of exposure to comparable BT levels were required for maximal reduction of the rate of ciliary beat in excised gills of this species.

Recent advances in endoscopy-guided sampling of gill tracts can also answer key questions about the site and mechanism of particle selection, thus contributing to our understanding of species-specific differences in toxin uptake capacity under varying particle scenarios. Most prior video-endoscopy studies used relatively small algae ($\leq 15 \mu\text{m}$ diameter). Less is known about bivalve feeding on large, non-spherical particles such as *P. multiseriis*, that attain up to $\sim 150 \mu\text{m}$ cell length and can vary greatly in size due to asexual division. It is also known that oysters, *Crassostrea virginica*, accumulate significantly lower DA levels than mussels, *Mytilus edulis*, when exposed to toxic *Pseudo-nitzschia* spp. under both field and laboratory conditions. We therefore used endoscopy-guided sampling to test whether oysters, which possess a highly complex, evolved gill, can selectively reject larger *P. multiseriis* cells from mixed phytoplankton assemblages, thus providing a mechanistic explanation for their low DA accumulation.

BRICELJ, V.M., J.E. WARD, A.D. CEMBELLA, and B.A. MACDONALD, 1998. Application of video-endoscopy to the study of bivalve feeding on toxic dinoflagellates. In: Harmful Algae, B. Reguera, J. Blanco, M.L. Fernández & T. Wyatt, Eds., Xunta de Galicia and IOC of UNESCO Publishers, pp. 453-456.

- GAINEY, L.F., and S.E. SHUMWAY, 1991. The physiological effect of *Aureococcus anophagefferens* ("brown tide") on the lateral cilia of bivalve mollusks. *Biological Bulletin* **181**: 298-306.
- WARD, J.E., L.P. SANFORD, R.I.E. NEWELL and B.A. MACDONALD, 1998. A new explanation for particle capture in suspension-feeding bivalve mollusks. *Limnology and Oceanography* **43**: 741-752.

IMPLEMENTATION OF A HARMFUL ALGAL BLOOM PREDICTION SYSTEM IN CHESAPEAKE BAY

Christopher Brown¹, Thomas Gross², Raleigh Hood³, Wen Long³, Bruce Michael⁴, Michael Naylor⁴, Douglas Ramers⁵, Peter Tango⁶, Jerry Wiggert⁷, and Jiangtao Xu⁸

¹National Oceanic and Atmospheric Administration, College Park, MD 20742, USA

²UNESCO, Paris, France

³University of Maryland Center for Environmental Science - Horn Point Laboratory 21613, USA

⁴Maryland Department of Natural Resources, Annapolis 21401, USA

⁵University of Evansville, Evansville 47722, USA

⁶United States Geological Survey, Annapolis 21401, USA

⁷Old Dominion University, Norfolk, VA 23529, USA

⁸National Oceanic and Atmospheric Administration, Silver Spring, MD 20910, USA

A variety of harmful algal blooms (HABs) impact Chesapeake Bay, degrading the bay's health and jeopardizing the viability of this important natural resource. We are developing an operational system that will predict the likelihood of three important HAB species in Chesapeake Bay and its tidal tributaries: the dinoflagellates *Karlodinium veneficum* and *Prorocentrum minimum* and the cyanobacteria *Microcystis aeruginosa*. Warnings of these events will aid in mitigating their deleterious effects on human and ecosystem health. The approach uses real-time and 3-day forecast data derived from a variety of sources and techniques to drive multi-variate, empirical habitat models that predict the probability of blooms of these HAB species. A prototype prediction system generates daily nowcasts and 3-day forecasts of *K. veneficum* using environmental conditions simulated by the hydrodynamic Regional Ocean Modeling System (ROMS) configured for Chesapeake Bay and a habitat model constructed using an artificial neural network. The predictions, in the form of digital images, are available via the World Wide Web to individuals and interested agencies to help guide research, recreational and management activities (Fig. 1). In particular, these predictions will be employed by the Maryland Department of Natural Resources to guide their response sampling efforts for HAB monitoring.

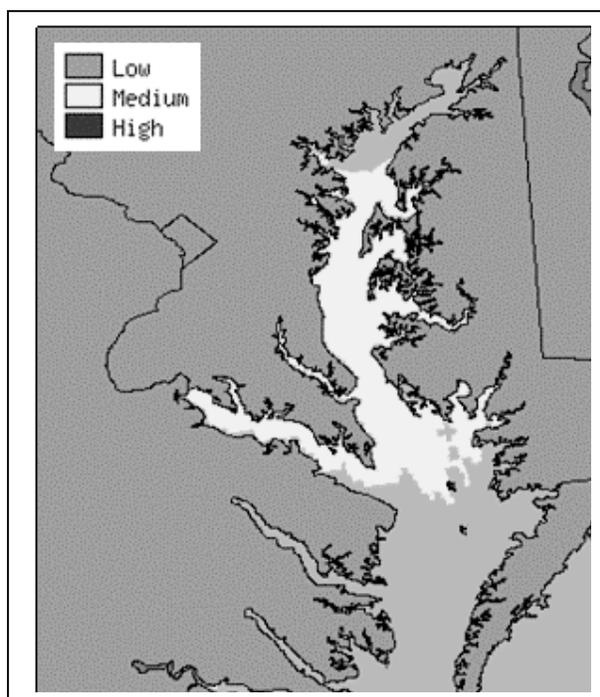


Figure 1. Predicted relative abundance of *Karlodinium veneficum* in Chesapeake Bay for July 11, 2007.

ISOLATION OF THE SODIUM CHANNEL GENE FROM THE COPEPOD *Acartia hudsonica* AND ITS POTENTIAL LINK TO SAXITOXIN RESISTANCE

Lihua Chen¹, Huan Zhang¹, Senjie Lin¹, David Avery¹, and Hans G. Dam¹

¹University of Connecticut, Dept. of Marine Sciences, Groton, CT USA 06340

Coastal environments worldwide are experiencing increases in harmful algal blooms that pose serious threats to human health and fisheries. Several species of the dinoflagellate genus *Alexandrium* produce paralytic shellfish poisoning (PSP) toxins, also known as saxitoxins, which accumulate in shellfish and water column grazers and are then transferred to higher trophic levels. Saxitoxins kill shellfish and finfish, marine mammals and humans by interfering with nerve function via blockage of the sodium channel, resulting in respiratory failure due to muscle paralysis. Mutations in the sodium channel gene, even if only in the form of a point mutation of one nucleotide (e.g., Bricelj et al. 2005), could confer resistance to this toxin. Copepods are the most abundant metazoans in the ocean, and hence a key link between primary producers and higher trophic levels. In previous studies in our laboratory, we used *Acartia hudsonica* as a model to demonstrate the development of resistance to toxic *Alexandrium* in some populations and a fitness advantage for the putatively adapted population (Colin and Dam 2004). Here, we examine whether the mechanism of such resistance may be related to mutations in the sodium channel.

We began with *A. hudsonica* population New Jersey coastal water where there has been no report of *Alexandrium* blooms. cDNA libraries (5'-end cDNA, Hexamer cDNA, regular oligo-dT cDNA, and modified oligo-dT cDNA) were synthesized using reported method (Lin and Zhang 2003, AEM 69: 343-349). Common primers were designed based on the alignment of the amino acid sequence of the sodium channel alpha subunit from other organisms. Using the combination of these primers, several fragments of copepod sodium channel were PCR amplified and sequenced. Specific primers were designed based on these fragments and combined with common primers to obtain the 7.1 kb full-length cDNA of *A. hudsonica* sodium channel alpha subunit (AhSC).

This is the first report of sodium channel alpha subunit from copepods. AhSC shares 50-60% similarity in amino acid level to counterparts in other organisms and has a typical channel structure comprising four domains with six motif[?] in each. Phylogenetic analysis reveals that AhSC is closest to the sodium channel alpha subunit of insects, consistent with taxonomic phylogeny. Strikingly, we found two types of sodium channel alpha subunits in *A. hudsonica*, which are identical **except that one (type 1) has a three-amino-acid insertion** between the third and the fourth functional domains of the protein. This insertion is located very close to an "IFM" or "MFM" [Isoleucine (Methionine, Valine), Phenylalanine and Methionine] motif that acts as an inactivation gate, opening and closing the channel. The insertion may shift the positioning of this motif when it is plugged in the channel, thereby affecting the sodium ion inflow. We hypothesize that this insertion may change the effect of saxitoxin on the sodium channel, and account for saxitoxin resistance in copepods.

Bricelj, V.M. Connell, L. Konoki, K. et al. 2005. Sodium channel mutation leading to saxitoxin resistance in clams increases risk of PSP. *Nature*. 434: 763-767.

Colin, S.P. and Dam H.G. 2004. Testing for resistance of pelagic marine copepods to a toxic dinoflagellate. *Evolutionary Ecology*. 18: 355-377.

Lin, S. and Zhang, H. 2003. Mitogen-activated protein kinase (MAPK) in *Pfiesteria piscicida* and its growth rate-related expression. *Appl. Environ. Microbiol.* 69: 343-349

SILICIC ACID LIMITATION IS NOT A TRIGGER FOR DOMOIC ACID PRODUCTION BY *Pseudo-nitzschia* BLOOMS IN THE PACIFIC NORTHWEST

William P. Cochlan¹, Mark L. Wells², Charles G. Trick³, Vera L. Trainer⁴, Evelyn J. Lessard⁵, and Barbara M. Hickey⁵

¹Romberg Tiburon Center for Environmental Studies, San Francisco State University, Tiburon, CA 94920, USA ²School of Marine Sciences, University of Maine, Orono, ME 04469, USA

³Schulich School of Medicine and Dentistry, University of Western Ontario, London, ONT, N6A 5B7, Canada

⁴NOAA Northwest Fisheries Science Center, Seattle, WA 98112, USA

⁵School of Oceanography, University of Washington, Seattle, WA 98195, USA

A central paradigm in the study of toxigenic diatoms is that concentrations of cellular DA (particulate DA per cell) become elevated as silicic acid (silicate) concentrations become limiting for growth and/or uptake by these diatoms. We have tested this hypothesis during numerous ECOHAB-PNW cruises from 2003-2006 in the coastal waters off Washington State, U.S.A. and British Columbia, Canada where toxigenic blooms of *Pseudo-nitzschia* typically occur. During 2004, we studied a massive toxic bloom measuring up to 48 km in diameter and reaching cell concentrations of 11-13 million cells/liter of *P. cuspidata* - the overwhelmingly dominant *Pseudo-nitzschia* species present. Results from this bloom event demonstrate that the highest levels of cellular toxin (5-64 pg DA/cell) correlate poorly with ambient silicate concentrations, and typically occur where dissolved silicate concentrations were 5-50 μM . None of the ~400 particulate DA (pDA) analyses conducted in 2004 (determined using cELISA) showed elevated cellular toxin concentrations when ambient concentration of silicate were $< 4 \mu\text{M}$, rather elevated pDA was generally associated with ambient silicate levels well above those considered limiting for its uptake and growth by most neritic diatoms. Increased cellular toxin levels also did not correlate with decreased ambient concentrations of nitrate or orthophosphate, indicating that toxin production in this natural *Pseudo-nitzschia* bloom was not governed by macronutrient availability. A similar result was found during the intense toxic *Pseudo-nitzschia* bloom in Monterey Bay, CA in 1998. The most established correlate for elevated domoic acid concentrations in 2004 was low dissolved iron concentrations; a finding consistent with laboratory culture experiments and our field incubation studies in the ECOHAB-PNW study region. These findings provide perhaps the most detailed insight to date into the environmental triggers for toxin production in natural assemblages of *Pseudo-nitzschia*, and demonstrate that commonly implicated macronutrient factors such as silicate limitation are poor predictors of either *Pseudo-nitzschia* dominance or toxicity in the Pacific Northwest.

THE EFFECTS OF TEMPERATURE AND EUTROPHICATION ON TOXIC AND NON-TOXIC STRAINS OF *Microcystis* WITHIN NEW YORK LAKESTimothy W. Davis¹, Christopher J. Gobler¹¹ School of Marine and Atmospheric Sciences, Stony Brook University, Stony Brook, NY 11794, USA

During recent decades, toxic cyanobacteria blooms have been increasing in frequency and intensity across the globe. The cyanotoxin, microcystin, which can be produced by *Microcystis*, has been linked to incidences of hepatic cancer and other grave medical conditions. This study focused on the effects of altered temperatures ($\pm 4^\circ\text{C}$) and nutrient levels (N, P) on toxic and non-toxic strains of *Microcystis* in Missisquoi Bay (MB) of Lake Champlain, which lies between New York and Vermont. Since it is not possible to distinguish between the different strains of *Microcystis* by means of traditional light microscopy, toxic and non-toxic strains were differentiated via quantification of the microcystin synthetase gene (*mcyD*) and ribosomal DNA gene, 16S. During the summer of 2006, Missisquoi Bay hosted chlorophyll concentrations of which ranged from 17.2 to 145 $\mu\text{g L}^{-1}$. The ratio of the cyanobacteria specific pigment phycocyanin-to-chlorophyll *a* within the phytoplankton community peaked in early August, suggesting dominance by cyanobacteria at this time. *Microcystis* was not detectable in this system until mid-July when the non-toxic strain dominated the *Microcystis* community (> 94% of the population). During an August *Microcystis* bloom, non-toxic and toxic populations peaked at $1.5 \pm 0.8 \times 10^7$ cell equivalents L^{-1} and $9.8 \pm 2.3 \times 10^5$ cell equivalents L^{-1} , respectively. In a manner similar to the *Microcystis* population, microcystin was not detectable until early August but remained elevated through October ranging from 0.8 to 2.0 $\mu\text{g L}^{-1}$, with peak concentrations coinciding with the peaks in toxic cell densities. Nutrient manipulation experiments suggested that MB was primarily phosphorus-limited throughout the summer months (May-September) when ambient DIP concentrations were low ($0.2 \pm 0.1 \mu\text{M}$) and water column alkaline phosphatase activities were elevated ($94 \pm 37 \text{ pmol h}^{-1} \text{ ml}^{-1}$). During 60% (3 of 5) of the experiments conducted at this time, increased levels of phosphorus lead to an increase in growth rates of the total phytoplankton community as well as of cyanobacteria (phycocyanin-specific growth rates). However, this system changed to being nitrogen-limited during the fall (October) when ambient DIN concentrations were low ($1.1 \pm 0.1 \mu\text{M}$) and experimental N additions yielded significantly increased growth rates for the total phytoplankton community. In experiments where increased cyanobacterial growth rates were elicited by increasing either N or P, an increase in temperature coupled with elevated nutrient concentrations often lead even higher growth rates. For example, in an experiment conducted during the peak of the August *Microcystis* bloom, increases in P significantly increased the growth rate of non-toxic *Microcystis* cells ($p \leq 0.001$, 2-Way ANOVA). In contrast, growth rates of toxic *Microcystis* cells were significantly enhanced by both N and P additions at this time ($p \leq 0.001$, 2-Way ANOVA). While similar trends were observed at higher incubation temperatures (4°C above ambient), the growth rates for each population associated with the significant treatments (N, P) were significantly greater than they were at ambient temperatures ($p \leq 0.002$, 3-Way ANOVA for both populations). Importantly, temperature also changed the relative abundance of toxic *Microcystis*. At ambient temperatures, the toxic population comprised between 0.45 and 0.96% of the total community in all treatments, whereas at elevated temperatures, toxic cells comprised 5 - 14%. In summary, the results suggest that increases in surface temperature coupled with nutrient loading could initiate a shift in dominance within the *Microcystis* population, causing toxic cells to comprise a greater percentage of the total population. Results on the response of toxic and non-toxic strains of *Microcystis* to nutrients and temperature from other New York lakes will also be presented.

FORMULATION OF ADVISORY LEVELS FOR CARIBBEAN AND PACIFIC CIGUATOXINS AND TIERED METHODS FOR THEIR DETERMINATION

Robert W. Dickey, H. Ray Granade, Edward L.E. Jester, Ann Abraham, Kathleen R. El Said, and Steven M. Plakas

U.S. FDA, Center for Food Safety and Applied Nutrition, Gulf Coast Seafood Laboratory, Dauphin Island, AL 36528, USA

Ciguatera fish poisoning is a seafood hazard caused by consumption of fish that have accumulated lipid-soluble ciguatoxins. Ciguatoxins traverse the food web from the primary producer(s), *Gambierdiscus* spp., to piscine predators in tropical and subtropical regions of the world. Expanding trade in fisheries from these regions gives rise to wider distribution and increasing frequency of ciguatera outbreaks among seafood consumers. Information derived from the study of ciguatera outbreaks has improved clinical recognition, confirmation, and timely treatment of this disease. Such studies are equally important for the differentiation of ciguatoxin profiles from one region to the next, the determination of ciguatoxicity thresholds in humans, and the formulation of policy for public health, regulatory and industry organizations. Review of case studies of the past decade suggests that 1.0 ppb Caribbean and 0.1 ppb Pacific ciguatoxins in finfish are approximate thresholds for adverse effects in seafood consumers. These estimates are derived from analyses of fish tissues implicated in ciguatera outbreaks using a two-tiered protocol to assess ciguatoxicity and confirm molecular presence of ciguatoxins in fish tissues. To these threshold estimates are added safety factors to address 1) individual human risk factors (e.g. pre-existing health status which influence susceptibility to ciguatoxins); 2) variation or uncertainty in the amount of fish consumed; and 3) uncertainty in assay accuracy (e.g. only C-CTX-1 and P-CTX-1 are available as reference standards, yet multiple ciguatoxin congeners are present in toxic fish). Using this approach toxin advisory levels for South Atlantic, Gulf of Mexico, Caribbean, and Pacific regions can be assessed for acceptable levels of consumer protection.

AUTONOMOUS, SUB-SURFACE DETECTION OF THE ALGAL TOXIN DOMOIC ACID ONBOARD THE ENVIRONMENTAL SAMPLE PROCESSOR

Gregory J. Doucette¹, Christopher A. Scholin², Christina M. Mikulski¹, Roman Marin III², Scott Jensen², Brent Roman², Dianne I. Greenfield², Kelly L. Jones¹, Kristen L. King¹, Jason Feldman³, Eugene Massion², Christopher T. Elliott⁴

¹Marine Biotoxins Program, NOAA/National Ocean Service, Charleston, SC 29412 USA

²Monterey Bay Aquarium Research Institute, Moss Landing, CA 95039, USA

³Jet Propulsion Laboratory, Pasadena, CA 91125, USA

⁴Queen's University Belfast, Belfast, BT9 5AG, Northern Ireland

The ability to detect harmful algal bloom (HAB) species and their toxins in real or near-real time has been identified as a critical need in the recently revised U.S. National Plan - Harmful Algal Research and Response: A National Environmental Science Strategy 2005-2015 (HARRNESS). Over the past decade considerable effort has been directed toward development of remote, in-situ diagnostic capabilities for harmful algal bloom (HAB) taxa, their genes, gene products, and metabolites, paralleling the rapid emergence of ocean observing initiatives and infrastructure that will accommodate the resulting technologies in the future. Both researchers studying HAB/toxin dynamics and coastal resource managers responsible for monitoring bloom populations as well as mitigating their socioeconomic impacts eagerly await the transition of such tools to operational status.

The Environmental Sample Processor (ESP; www.mbari.org/microbial/ESP), developed for the autonomous, sub-surface application of molecular diagnostic tests, has successfully detected several HAB species using DNA probe arrays (e.g., Greenfield et al., 2006). Since toxin production and thus the potential for public health and ecosystem effects can vary widely with algal physiological status as influenced by a number of environmental factors, the concurrent detection of algal toxins onboard the ESP is essential. We have developed methods for extracting domoic acid (DA) from toxic *Pseudo-nitzschia* cells (extraction efficiency > 90%) and testing of samples using immuno-based membrane arrays on the ESP platform. Current toxin detection limits in extracts are in the 10's of ng DA per mL and the average repeatability between independent analyses ranges from 15-20%. Through the conduct of species- and toxin-specific arrays, potentially toxic species of *Pseudo-nitzschia* and domoic acid were detected concurrently on a second generation (2G) ESP instrument during 2006 deployments in Monterey Bay, CA, USA, representing the first remote, integrated assessment of algal cell abundance and toxin presence in coastal waters. Efforts are now underway to increase the assay sensitivity and refine its calibration in order to obtain more accurate estimates of a bloom's toxicity and thus its potential impacts. Deployment of the modified DA assay method is planned for August-September 2007 in Monterey Bay. Ultimately, integration of multiple ESPs with ocean observing systems will enhance our monitoring, prediction, and management capabilities for HABs and their adverse effects.

Greenfield, D.I., R. Marin III, S. Jensen, E. Massion, B. Roman, J. Feldman, C.A. Scholin. 2006. Application of environmental sample processor (ESP) methodology for quantifying *Pseudo-nitzschia australis* using ribosomal RNA-targeted probes in sandwich and fluorescent in situ hybridization formats. *Limnol. Oceanogr.: Methods* 4: 426-435.

ASSESSING THE ROLE OF ENVIRONMENTAL STRESSORS AND GENETIC COMPOSITION ON MICROCYSTIN PRODUCTION IN LAKE ERIE *Microcystis* POPULATIONS

Juli Dyble¹, Gary L. Fahnenstiel¹, H.A. Vanderploeg¹, and R.W. Litaker²

¹NOAA, Great Lakes Environmental Research Lab, Ann Arbor, MI 48105, USA

²NOAA, Center for Coastal Fisheries and Habitat Research, Beaufort, NC 28516, USA

Blooms of the cyanobacterial HAB *Microcystis* have recently resurged in some regions of the Great Lakes, coincident with the invasion of zebra mussels to the system. Due to the use of these waters for drinking water and recreation, there is a significant need to understand the factors contributing to bloom toxicity and develop the ability to forecast when the presence of these blooms will be a threat to human health. *Microcystis* bloom toxicity is regulated both by environmental factors and genetic composition of the strains present in the bloom. Some of the environmental factors that have previously been shown in culture experiments to influence HAB toxicity (nutrients, light and grazers) were manipulated in laboratory experiments using natural communities of the cyanobacteria HAB *Microcystis* from western Lake Erie. The response of the *Microcystis* community to these environmental stressors was assessed by measuring changes in growth rates and in the concentration of the toxin microcystin. Additionally, a quantitative PCR assay was used to quantify changes in the number of toxic colonies using the *mcyB* gene, which is involved in the synthesis of the toxin microcystin. The genetic composition of the *Microcystis* community in western Lake Erie was also assessed over the course of a bloom season using both the highly variable phycocyanin intergenic spacer region (PC-IGS) and *mcyB*. Sequence analysis showed that there is shift in community composition over time and that strain composition plays a role in *Microcystis* bloom toxicity in this region.

MONITORING TOXIC *Alexandrium catenella* IN THE PUGET SOUND USING REAL-TIME QUANTITATIVE PCR (QPCR)

Sonya T. Dyhrman¹, Sheean T. Haley¹, Jerry A. Borchert², Bob Lona³, and Deana L. Erdner⁴

¹Woods Hole Oceanographic Institution, Woods Hole, MA 02543, USA

²Shellfish Program, Washington State Department of Health, Tumwater, WA 98501, USA

³Marine Biotxin Unit, Washington State Department of Health, Tumwater, WA 98501, USA

⁴University of Texas at Austin, Marine Science Institute, Port Aransas, TX 78373, USA

Dinoflagellates of the genus *Alexandrium* can produce a suite of potent neurotoxins that cause paralytic shellfish poisoning (PSP) in humans, and can have serious deleterious impacts on public health and economic resources. *Alexandrium* and related PSP-toxicity is a problem of global scale. Within this genus, *Alexandrium catenella* is widespread in the northwestern part of North America, including the Puget Sound, and is responsible for seasonal harmful algal blooms (HAB) in these regions. Even at low cell densities, *A. catenella* toxins can accumulate in shellfish and result in PSP. As a result, accurate measurements of *A. catenella* distributions, particularly at low cell density, are critical to continued PSP monitoring and mitigation efforts. For example, detection of low, but increasing cell densities may trigger increased PSP monitoring, or help to target PSP monitoring to specific locations or time-periods. Towards this end a specific, sensitive, and high throughput real-time quantitative PCR (qPCR) method has been developed to assay the abundance of *A. catenella*.

In this study, Puget Sound surface water samples for qPCR analyses, microscope cell counts, and shellfish for PSP analyses (typically *Mytilus edulis*) were collected every two weeks from April 2006 through October 2006 by community volunteers and local public health organizations from 41 Sentinel Sites distributed throughout the Puget Sound. qPCR amplification of DNA extracted from field samples and standards was performed with a SYBR Green detection system. With the qPCR assay, low water column abundances of *A. catenella* of less than 10 cells per liter were measured. The detection of low cell numbers by qPCR resulted in the ability to report cells at all Sentinel Sites before these sites reached the USDA's regulatory PSP limit. Monitoring cell abundance by qPCR predicted, at times, an increase in PSP toxicity. This was seen for roughly half of the sampled Sentinel Sites. Often the increase in cell abundance occurred a week or two in advance of the increase in PSP toxicity. However, given the variability associated with the sites, qPCR cell counts were unable to define an absolute or threshold cell number necessary to predict PSP toxicity. There is a clear seasonality to *A. catenella* bloom dynamics in Puget Sound, as cell numbers increased substantially in nearly all Sentinel Sites from May to October. This first sampling season has begun to establish the utility of qPCR in providing early warning of PSP toxicity and its utility as a tool for deriving the seasonal bloom patterns of *A. catenella* within the Puget Sound. These bloom patterns are some of the first data for *A. catenella* cell numbers in this region and will ultimately help guide management practices. These results will be coupled with an additional field season to establish a more concrete relationship between cell number, bloom dynamics and PSP toxicity.

POPULATION GENETICS OF TOXIC *Alexandrium* BLOOMS IN THE GULF OF MAINE

Deana L. Erdner¹, Linda A. R. McCauley², Katie Libera² and Donald M. Anderson²

University of Texas Marine Science Institute, Port Aransas, TX 78373, USA

Woods Hole Oceanographic Institution, Woods Hole, MA 02543, USA

Coastal waters of the northeastern U.S. are subject to recurrent outbreaks of paralytic shellfish poisoning (PSP) caused by the toxic dinoflagellate *Alexandrium fundyense*. PSP toxicity shows considerable spatial and temporal variability, due in part to growth and accumulation zones defined by coastal circulation patterns. Another likely contributing factor is phenotypic heterogeneity in the *A. fundyense* population, evidenced by dramatic differences in inherent toxicity amongst clonal *A. fundyense* isolates from the region. Overall, this variability in PSP toxicity represents a significant challenge from public health and fisheries management perspectives. As part of the Woods Hole Center for Oceans and Human Health, one of our aims is to investigate the manner in which genetic diversity affects bloom dynamics. More effective management of shellfish resources threatened by PSP toxins requires an understanding of the population biology of *Alexandrium* species, as well as the manner in which these different genotypes respond to environmental conditions, as this ultimately affects the quantity and types of toxins accumulating in shellfish.

A set of microsatellite markers, developed for *A. tamarense*, has been used to study the genetic diversity of bloom populations of *A. fundyense* throughout the Gulf of Maine. These markers are highly polymorphic in *A. fundyense*, and are able to resolve fine-scale genetic differences between isolates. Toxic *Alexandrium* blooms occurred in the Gulf of Maine in 2005 and 2006, with the 2005 bloom being the largest toxic bloom in at least several decades. Several hundred clonal isolates were established from water samples taken throughout the blooms, in different geographic locations and at different times. The genotypes of these toxic bloom isolates were determined using microsatellite markers and used to assess spatial and temporal changes in the genetic composition of the bloom. In 2005, early-bloom populations from Bay of Fundy, Casco Bay, and MA Bay were not significantly different from one another. However, late-bloom populations collected near Martha's Vineyard (the southern extent of the bloom) were genetically distinct from early-bloom samples. In 2006, populations were collected from across the Gulf of Maine region, and none were significantly different from one another. The bloom that year did not extend to the Martha's Vineyard area. Further, a comparison of 2005 and 2006 samples showed that, in general, populations from the two different years were not genetically distinct. We also sampled a 2006 bloom in an isolated embayment, Salt Pond, MA. Populations in Salt Pond were genetically distinct from those in the wider Gulf of Maine, and they changed over the course of the 3-week bloom. From these two years of data, it appears that overall genetic composition of *Alexandrium* blooms in the Gulf of Maine is not significantly different from year to year. Within a year, however, we did observe changes in bloom populations on the timescale of approximately one month. This could result from the natural progression or 'turnover' of genotypes during a bloom, or from the mixing of genetically distinct cells from other (unknown) sources. Results of the 2006 analysis of the Salt Pond bloom provides support for the former hypothesis, although the mixing of different source populations cannot be discounted.

DETERMINATION OF PARALYTIC SHELLFISH POISONING TOXINS USING THE LAWRENCE METHOD: APPLICATION TO HUMAN URINE AND SERUM

Stacey Etheridge¹, Victor Rivera², Kevin White¹, John Roach¹, and Mark Poli².

¹Center for Food Safety and Applied Nutrition, US Food and Drug Administration, College Park, MD 20740

²Integrated Toxicology Division, US Army Medical Research Institute of Infectious Diseases, Ft Detrick, MD 21702-5011

The AOAC Lawrence HPLC method for determining paralytic shellfish poisoning (PSP) toxins in shellfish was evaluated on the human clinical matrices urine and serum. Initial analysis revealed an interfering, naturally-occurring fluorescent compound in urine. Further analysis by high resolution mass spectrometry identified the compound to be hippuric acid, a major constituent in human urine originating from dietary sources. The hippuric acid was removed from samples by adjusting the pH to 4 prior to sample clean-up and by doubling the SPE cartridge bed volume. Interference by naturally-occurring fluorescent compounds was found to be minimal in the serum matrix. Quantitation of a range of PSP congeners spiked in these matrices will be presented and implications for public health will be discussed.

FLORIDA AQUATIC TOXINS HOTLINE: FORMAL EVALUATION OF HAB OUTREACH AND EDUCATIONAL ACTIVITIES

Fleming LE¹, Jerez E², Stephan W², Cassedy A³, Bean JA³, Reich A⁴, Kirkpatrick B⁵, Backer L⁶, Nierenberg K⁵, Watkins S⁴, Hollenbeck J¹, Weisman R².

¹NSF NIEHS Oceans and Human Health Center, University of Miami, Miami, FL; ²South Florida Poison Information Center, University of Miami, Miami, FL; ³ Biostatistics, Cincinnati Childrens Hospital Medical Center, Cincinnati, OH; ⁴Aquatic Toxins Group, Florida Dept of Health, Tallahassee, FL; ⁵Mote Marine Laboratory, Sarasota, FL; ⁶National Center for Environmental Health (NCEH), Centers for Disease Control and Prevention (CDC), Atlanta, GA

With the increasing number of persons interacting with the coastal areas (both freshwater and marine) and with the apparent increase of harmful algal blooms (HABs) around the world, healthcare providers, public health personnel and coastal managers are struggling to provide scientifically-based appropriately targeted outreach and education to people with possible marine and freshwater toxin diseases and to persons with possible exposure to the marine and freshwater toxins, as well as the media and the general public. A variety of outreach and education materials and services have been created, but there has been almost no formal evaluation to determine whether these materials are effectively reaching their target audience and meeting the audience's expectations.

The Florida Poison Information Center-Miami (FPIC-Miami) has provided 24 hour 365 day/yr toll free Aquatic Toxins Hotline (1-888-232-8635) in several languages which has received over 25,000 calls since its inception in 1998. All calls are answered by highly trained Poison Information Specialists. These calls are reported as a form of passive surveillance of HAB-related illness and information requests to the Aquatic Toxins Program of the Florida Department of Health and to the Centers for Disease Control and Prevention. Recently, the Hotline was expanded to include an automated call processing menu system that allows callers to access information in English or Spanish. Callers can get information about the health effects and locations of the Florida red tide (including the NOAA HAB Bulletin), ciguatera fish poisoning, and blue green algae (cyanobacteria), and resources for learning about general marine toxin issues. Callers also have the opportunity to speak directly with a Poison Information Specialist.



This Pilot Study is the first known evaluation of the use of and satisfaction with outreach and education materials for HABs. Overall, the majority (68%) of callers reported that they were satisfied with the information provided by the Aquatic Toxins Hotline. Most callers were also satisfied with specific services offered by the Aquatic Toxins Hotline automated system (including speaking directly with a Poison Information Specialist), and some callers provided specific suggestions for improvements. This study demonstrated that the new automated system quickly provided useful HAB-related information for the large majority of the callers (78%), thus decreasing the workload of routine informational calls for the Poison Information Specialists and allowing them to focus on the those persons who may truly be ill. The results from this study will lead to the expansion and improvement of this valuable HAB outreach, education and surveillance tool, as well as recommendations for the evaluation of other HAB outreach and educational materials.

IMPACTS OF EUTROPHICATION-RELATED BLOOMS OF *Prorocentrum minimum* and *Karlodinium veneficum* ON EARLY LIFE STAGES OF OYSTERS IN CHESAPEAKE BAY

Patricia M. Glibert¹, Jeffrey Alexander¹, Diane K. Stoecker¹ and Donald Meritt¹

¹University of Maryland Center for Environmental Science, Horn Point Laboratory, PO Box 775, Cambridge, MD 21613, USA

Eutrophication-related algal blooms, especially *Prorocentrum minimum* and *Karlodinium veneficum* are now common in Chesapeake Bay. Of their documented impacts, high biomass leading to low oxygen and fish kills are the most recognized. Here we report on a suite of impacts that these blooms have on early life stages of oysters. Native oyster (*Crassostrea virginica*) populations are now <1% of their historical values in Chesapeake Bay and a major restoration effort is underway using hatchery-produced spat for restocking natural habitat. Furthermore, there is also consideration of introduction of a new oyster species, *Crassostrea ariakensis*, which is thought to be more disease resistant and faster growing than its native counterpart. All phases of hatchery operations depend on ambient water, which can be filtered or treated prior to use. To test the effects of HABs, exposure experiments of various life stages of oysters to cells and their exudates were conducted. When embryos from freshly spawned *C. virginica* were exposed immediately to *K. veneficum* at 10⁴ cells/ml, either alone or in combination with other algae, virtually all of the developed larvae were deformed within 24-48 hours and were dead within 72-96 hours. Similar effects were found for *C. ariakensis* exposed to *K. veneficum* at a single concentration of 10⁴ cells/ml. No deformities, and mortalities of <40%, were observed in controls to which a standard diet of the haptophyte *Isochrysis* was added. Effects of exudates of *K. veneficum* and of whole cells of *P. minimum* have been fewer. Viability increased somewhat when older larvae were exposed to the same HAB. Blooms of *P. minimum* and *K. veneficum* are most prevalent during spring and summer, when spawning of *C. virginica* occurs. Eutrophication and associated harmful algal blooms have the potential to reduce survival of early life history stages of oysters and hence to reduce oyster recruitment. Any reduction in recruitment either spatially or temporally, combined with an overall reduction in sheer numbers of larvae that survive, will make establishment of significant, self-sustaining populations of natural or introduced oyster species more difficult.

PRELIMINARY INSIGHT FROM THE FIRST GENOME-SEQUENCE OF A HARMFUL ALGAL BLOOM SPECIES, THE BROWN TIDE ALGA, *Aureococcus anophagefferens*

Christopher J. Gobler¹, Steven W. Wilhelm², Dianna L. Berry¹, Leo Poorvin², Neha Sarode², Astrid Terry⁴, Igor Grigorev⁴, Mine Berg³, and The *Aureococcus* Genome Consortium.

¹ School of Marine and Atmospheric Sciences, Stony Brook University, Stony Brook, NY 11794, USA

² Department of Microbiology, The University of Tennessee, Knoxville, TN 37996 USA

³ School of Earth Sciences, Stanford University, Stanford, CA 94305 USA

⁴ Department of Energy, Joint Genome Institute-Lawrence Berkley National Laboratory, Walnut Creek, CA 94598 USA

Aureococcus anophagefferens is the causative species of harmful brown tide blooms which have plagued the eastern US seaboard for more than twenty years. While brown tides can be sporadic, the concurrent outbreak of massive *A. anophagefferens* blooms in NY, NJ, and MD during 2007 attested to their persistence and pervasiveness. The sequence of the *A. anophagefferens* genome has been completed by the Department of Energy's Joint Genome Institute, a process which will facilitate a better understanding of this organism's ecology, as well as provide impetus for the development of novel molecular tools. A combination of expressed sequence tags (ESTs, ~ 50,000) and raw genomic data (~ 7-fold coverage of the organism's 56 megabase genome) have been completed and are undergoing analysis. Preliminary bioinformatic predictions suggest that > 11,500 identifiable genes exist within the genome with an average gene length of 2,138 base pairs, an average transcript length of 1601 base pairs, and an average protein length of 523 amino acids. Gene density was estimated to be 205 genes per megabase pair. Many gene models were supported by EST presence and/or putative gene homologs found in public informatics databases. A total of 49% of the genes were predicted by homology, while Ab-Initio prediction and EST clusters were used to predict the remaining 49% and 2%, respectively. Between 68 and 77% of the predicted proteins were homologous with published gene sequences found in the P-FAM and Swissprot databases. This preliminary introduction to the *A. anophagefferens* genome will emphasize the presence and expression of genes involved in nutrient utilization and cycling, as well as photosynthesis.

DOMOIC ACID NEUROTOXICITY IN NATIVE AMERICANS IN THE PACIFIC NORTHWEST: HUMAN HEALTH PROJECT METHODS AND UPDATE

Lynn M. Grattan,^{1,5} Sparkle Roberts,¹ Vera Trainer,² Carol Boushey³, Tom Burbacher⁴, Kimberly Grant⁴, Kate Tracy⁵ and J. Glenn Morris⁵

¹ Department of Neurology, University of Maryland School of Medicine, Baltimore MD, USA 21201

² Northwest Fisheries Science Center, Seattle, WA USA 98112

³ Department of Foods and Nutrition, Purdue University, West Lafayette, IN USA 47907

⁴ Department of Environmental and Occupational Health Sciences, University of Washington, Seattle, WA 98195

⁵ Department of Epidemiology and Preventive Medicine, University of Maryland School of Medicine, Baltimore MD, USA 21201

A five-year epidemiological cohort study is currently underway in the Pacific Northwest to determine whether or not there are human health effects of low-level exposure to Domoic Acid through Razor Clam consumption. The cohort is comprised of a random sample of 653 Native Americans, stratified by age (6 months to 80 years) from three coastal tribes in Washington State. The project is currently in year three of data collection. The methodology includes a broad range of valid and reliable standard behavioral, dietary, medical history and symptom assessment tools. This includes the Occupational and Environmental Neurology Questionnaire, Shellfish Assessment Survey, Block Food Frequency Questionnaire and Daily Food Records. Specialized, state of the art methods for assessing cognition and memory across the lifespan are also being utilized including select subtests of the Wechsler Child and Adult Intelligence Scales, Ravens Progressive Matrices, California Verbal Learning Test, Mullen Scales of Early Learning and the Fagan Test of Infant Intelligence. Domoic Acid levels are being obtained on a regular basis (every two weeks) from razor clams harvested from the regions under study.

Baseline findings indicate that 1) low levels of Domoic Acid have been consistently reported in the regions under study, 2) a significant number of persons in the cohort eat razor clams, 3) the general intellect of the cohort is distributed normally and similar to the overall U.S. population and 4) behavioral variables such as depression and substance abuse are also normally distributed in the study sample. Available data to date suggests that persons under study are at risk of low-level exposure to Domoic acid. Whether or not there is health risks associated with low-level exposure remains to be determined.

APPLICATIONS OF THE SECOND-GENERATION ENVIRONMENTAL SAMPLE PROCESSOR (2G ESP) FOR REMOTE DETECTION OF HARMFUL ALGAE: 2007

Greenfield¹, D.I., Jensen¹, S., Marin¹, R. III, Roman¹, B., Everlove¹, C., Pargett¹, D., Alvarado¹, N., Doucette², G.J. and Scholin¹, C.A.

¹Monterey Bay Aquarium Research Institute, Moss Landing, CA 95039, USA

² Marine Biotoxins Program, NOAA/National Ocean Service, Charleston, SC 29412 USA

Molecular approaches for identifying harmful algal bloom (HAB) species and affiliated toxins are central to research and monitoring, but such methods typically require the return of discrete samples for laboratory analysis. We are working to overcome this impediment by using the Environmental Sample Processor (ESP; <http://www.mbari.org/microbial/ESP>). The second-generation (2G ESP) was deployed for the first time in Monterey Bay, CA during 2006 and has the capacity to develop DNA probe arrays for target HABs, archive samples, and detect the phycotoxin domoic acid (DA). During 2006, the 2G ESP successfully automated detection of a number of harmful species, including diatoms of the genus *Pseudo-nitzschia*, some of which produce DA. The ESP and affiliated assays also detected DA itself.

Deployments of the 2G ESP in Monterey Bay are on-going. At the time of this submission, we completed 2 deployments during April -June 2007. A number of HABs, including members of the genera *Pseudo-nitzschia*, *Alexandrium*, and *Heterosigma* were detected. We are also refining methods that validate instrument performance by periodic water sampling and analyses using laboratory versions of molecular assays that are emulated aboard the ESP. In addition to field studies, we are generating standard curves for a number of HABs as part of an effort to become more quantitative with our DNA arrays. Finally, we are investigating the utility of qPCR for both *Heterosigma akashiwo* and *Alexandrium* sp. for comparisons to instrument results. Here we present findings from the 2007 field sampling season as well as examples of laboratory studies aiming confirm results from the field.

EFFECT OF THE HARMFUL ALGA *Prorocentrum minimum* ON THE HEMOCYTE RESPONSE OF QUAHOGS *Mercenaria mercenaria* WITH VARIOUS LEVELS OF QPX INFECTION

Hélène Hégaret¹, Gary H. Wikfors², Roxanna Smolowitz³, Sandra E. Shumway¹

¹Department of Marine Sciences, University of Connecticut, Groton, CT, USA.

²NOAA-NMFS, Milford, CT, USA

³Marine Biological Laboratory, Woods Hole, MA 02543, USA

Northern quahogs *Mercenaria mercenaria* are regularly exposed to harmful algal blooms (HABs), which can impact the immune system and, thus, susceptibility to infection by parasites. Moreover, quahogs in a given population can be more- or less-severely infected by a parasite, Quahog Parasite Unknown, or QPX. HABs occur in areas where parasite-stressed shellfish are located. As many parasites can modify hemocyte characteristics in affected bivalves, we conducted experiments to assess the combined effects of both stresses on clams.

To evaluate the possible individual or combined effects of the harmful alga *Prorocentrum minimum* and the parasite QPX, on hemocyte characteristics of quahogs *M. mercenaria*, an experiment was conducted exposing quahogs with varying levels of QPX infection to a simulated bloom of cultured *P. minimum* for five days. The simulated bloom was created by adding cultured *P. minimum* to a natural phytoplankton assemblage, and control clams were exposed to just the natural phytoplankton with no amendment. Hemocyte characteristics measured with flow-cytometric analyses (hemocyte concentration, morphology, percentage of phagocytic cells, viability, adhesion, apoptosis and production of reactive oxygen species), histology and prevalence and intensity of QPX were assessed for individual quahogs after microalgal exposure.

The results indicate that an exposure of quahogs to *P. minimum* for 5 days triggers an increase in the size of the hemocytes and in the production of reactive oxygen species of hemocytes as well as a decrease in the percentage of phagocytic hemocytes. Histological observations also shown the presence of some significant inflammations in the tissues of quahogs exposed to *P. minimum*. A lack of infected animals fed the natural plankton did not permit us to assess the effect of QPX alone. But the results indicate that the presence of QPX infection affect the hemocyte parameters of quahogs exposed to *P. minimum* by decreasing the size of the hemocytes.

This study revealed that the harmful alga *P. minimum* can alter hemocyte morphology and functions after only five days of exposure. In contrast, the presence of the parasite modified only the size of the hemocytes, no other responses of the hemocytes of the quahogs to this dinoflagellate. Thus, *P. minimum* effects upon quahogs are consistent, regardless of parasite QPX burden.

REGIONAL OCEANOGRAPHY LEADING TO TOXIC *Pseudo-nitzschia* EVENTS ON BEACHES IN THE NORTHERN CALIFORNIA CURRENT

Hickey, B.M¹, A. MacFadyen¹, V.L. Trainer², E.J. Lessard¹, W.P. Cochlan³, C.G. Trick⁴ and M.L. Wells⁵

¹School of Oceanography, Univ. Washington, Seattle, Washington 98195;

²NOAA, Northwest Fisheries Science Center, Seattle, Washington, 98112;

³Romberg Tiburon Center for Environmental Studies, San Francisco State Univ., Tiburon, California 94920;

⁴Department of Biology, Univ. Western Ontario, London, Ontario, Canada N6A 5B7

⁵School of Marine Sciences, Univ. Maine, Orono, Maine 04469

Recent interdisciplinary studies of *Pseudo-nitzschia* off the Washington/Oregon coasts have improved our understanding of the physical/chemical/biological oceanographic elements leading to significant toxic conditions in razor clams on coastal beaches. First, phytoplankton must become concentrated in a bloom source region such as the Juan de Fuca eddy. ECOHAB PNW studies suggest this requires a period of downwelling-favorable or lightly fluctuating winds. 2) Environmental conditions leading to elevated domoic acid (DA) levels must be present. In this region, cellular DA levels appear to be influenced by iron and copper availability and not by macronutrient levels. 3) The Juan de Fuca eddy region, which has been shown to be a regional source of domoic acid, is located offshore and well to the north of coastal beaches that have experienced toxic outbreaks. Patches of toxic phytoplankton must first escape this eddy-like offshore source region--escape is favored during upwelling-favorable wind conditions, which allow the geostrophic constraint of the eddy circulation pattern to be broken. 4) The patch must then move alongshore to sites with shellfish populations and 5) the cells must retain their cellular toxin content during the time period of transport. For a toxic source in the Juan de Fuca eddy this requires southward advection along the shelf, as occurs during periods of upwelling-favorable winds in summer and early fall. ECOHAB PNW studies show that toxin can be maintained in the 7-14 days required for transport. For an Oregon source of toxin such as Heceta bank to impact the Washington shelf, this requires northward advection along the shelf, as occurs during periods of downwelling-favorable winds in spring. Last, 6) the patch of toxic phytoplankton must move onshore to coastal beaches and/or estuaries and 7) must remain there for a period sufficient for significant ingestion by shellfish. In the northern California Current this requires a period of significantly strong downwelling-favorable winds, as occurs during a storm. The requirement to fulfill each of these several steps illustrates why toxic events occur sporadically on coastal beaches in spite of the frequent observations of toxicity in offshore locations.

THE NEW ENGLAND 2005 *Alexandrium* BLOOM: ESTIMATES OF THE ECONOMIC EFFECTS ON COMMERCIAL SHELLFISHERIES

Porter Hoagland¹, Di Jin¹ and Eric Thunberg²

¹Marine Policy Center, Woods Hole Oceanographic Institution, Woods Hole, MA 02540, USA

²Social Sciences Branch, Northeast Fisheries Science Center, National Marine Fisheries Service, Woods Hole, MA 02543, USA

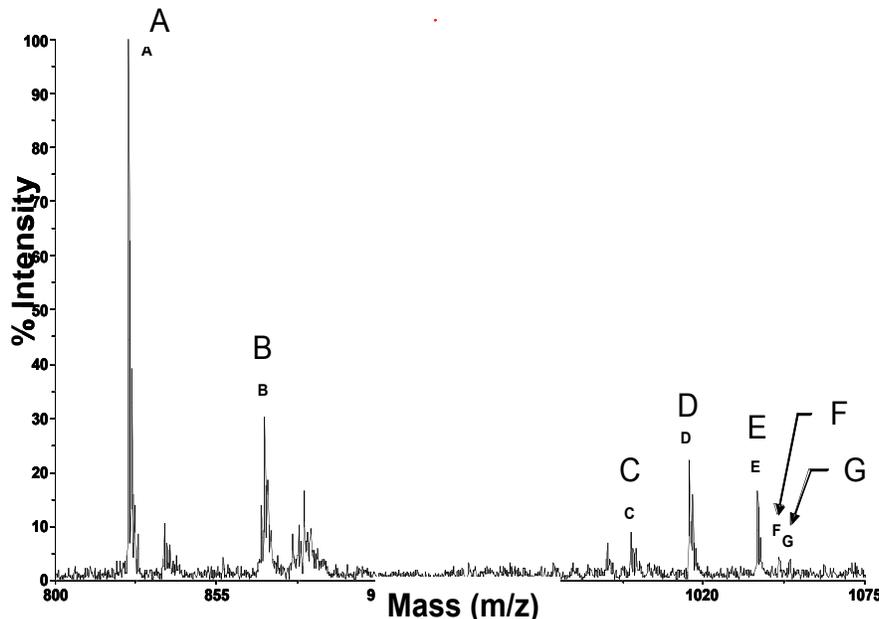
Over the last several decades, harmful algal bloom (HAB) events have been observed in more locations than ever before throughout the United States. The 2005 bloom of *Alexandrium fundyense* was the most widespread and intense in New England waters since a similar event more than three decades ago. Oceanographers have argued that another significant *Alexandrium* bloom could recur in the future due to the deposition of large quantities of algal cysts in New England ocean sediments. If true, then the results of our study should provide useful information for the public and natural resource managers. Using historical data from the National Marine Fisheries Service, the Maine Department of Marine Resources, the Massachusetts Division of Marine Fisheries, and other sources, we develop estimates of the direct economic impacts of the 2005 event on commercial shellfishing and growing industries in Maine and Massachusetts. We identify the effects of the event on market supply channels and prices using empirical data from the shellfishing and growing industries. Results of our analysis suggest that the 2005 event had broad spatial and temporal effects on certain components of the shellfish market. In response to a supply shortage resulting from local closures, there was an increase in shellfish imports to New England during the red tide. Our results indicate that the low-end estimate for total direct impacts in Maine was \$2.4 million, including lost revenues in the softshell clam and mussel fisheries. Shellfish closures in Maine were the most likely cause of observable price changes for softshell clams on the Fulton Fish Market in New York. The total direct impacts on the commercial shellfish industry in Massachusetts may have been as high as \$18 million. (Because of serious data limitations, however, this estimate should be viewed with caution.) To improve estimates of HAB impacts in Massachusetts, the consistent compilation of monthly shellfish landings is essential. We compare these results with estimates of economic impacts from other locations in the nation, which have been compiled in a continuing effort to characterize the economic effects of HABs in the United States.

NEW TRICKS WITH OLD TOYS: APPLICATION OF MASS SPECTROMETRY TO THE ANALYSIS OF PEPTIDE TOXINS

Karen Howard, Michael F Satchwell and Gregory L. Boyer

¹Faculty of Chemistry, State University of New York, College of Environmental Science and Forestry, Syracuse NY 13210 (glboyer@esf.edu)

Microcystins (MCs) are potent toxins produced by some species of cyanobacteria. Rapid analysis of natural waters for MC content is essential to protect the public from exposure through drinking water or recreational contact. Matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS) offers several advantages for analysis of MCs over current methods, including speed, reduced sample handling, and congener identification, but has previously been limited to qualitative analysis of MCs. In this study, a recently developed quantitative MALDI-TOF method for analysis of MCs was used to analyze intact cells and pre-extracted samples, with comparison to the results obtained using PPIA and by HPLC with electrospray ionization mass spectrometry (ESI-MS). Addition of MALDI matrix to intact cell samples provided simple, rapid identification and quantitation of MCs. Consistent results were obtained across a wide range of cyanobacterial and diatom densities, with recoveries from 87.2% (MC-LR) to 113.2% (MC-YR). Samples that were pre-extracted using a traditional approach were analyzed in parallel by PPIA, MALDI, and ESI-MS. The initial correlation between the MALDI and PPIA results ($R^2 = 0.69$) improved significantly ($R^2 = 0.81$) when the MALDI results were adjusted based on the mouse bioassay toxicity of each identified congener. The results confirm the utility of MALDI-TOF MS as a quantitative screening method for MCs, and the potential for its use as a stand-alone technique. Advantages and disadvantages of this technique, the selection of the appropriate internal standard, as well as a comparison with the more traditional LCMS-ESI will be presented.



Maldi TOF Mass spectrum of an intact cell protocol (ICP) field sample from Lake Neatahwanta, NY. Peak identification based on m/z ratio: (A) nodularin (internal standard); (B) phaeophytin α ; (C) MC-LR; (D) desmethyl MC-FR; (E) MC-RR; (F) MC-YR; and (G) MC-(H₄)YR.

TEMPORAL AND SPATIAL VARIABILITY IN PACIFIC NORTHWEST *Pseudo-nitzschia* POPULATIONS

Katherine A. Hubbard¹, E. Virginia Armbrust¹

¹University of Washington School of Oceanography, Seattle, WA 98195, USA

Diatoms in the genus *Pseudo-nitzschia* are a common component of phytoplankton assemblages, although little is known about the distribution of individual species and/or strains in any given location. Automated Ribosomal Intergenic Spacer Analysis (ARISA), a form of DNA fingerprinting, was developed to recognize specifically at least 14 different species of *Pseudo-nitzschia*. This technique allows the rapid determination of species distributions over time and space. Puget Sound, WA is an estuary connected to the Pacific Ocean by the Strait of Juan de Fuca; within the estuary, there are four interconnected but hydrographically distinct basins. We examined species distributions within the different basins of Puget Sound during a two-day window in June, 2006. Overall, six *Pseudo-nitzschia* species were detected throughout the Sound, but species composition varied among the different basins. For example, some species such as *P. multiseriata* and *P. australis/P. seriata* were restricted to a subset of the basins, whereas the distributions of other species such as *P. pungens* and *P. delicatissima* were much more widespread. Preliminary results from a time series collected at single site in Puget Sound indicated that *P. pungens* and *P. delicatissima* persist across multiple seasons, and PCA analyses suggest that these species may have lower environmental constraints than other species. Environmental clone libraries were generated to investigate the role of intraspecific diversity in distribution patterns. Interestingly, *P. pungens* and *P. delicatissima* exhibited the most and least intraspecific diversity, respectively, of the ten *Pseudo-nitzschia* species detected in clone libraries. At this time, 13 distinct genotypes of *P. pungens* have been detected in Pacific Northwest waters, 12 of which appear to be unique to the Pacific Northwest, and 5 of which appear to be unique to Puget Sound. Nucleotide divergence for Pacific Northwest *P. pungens* (4.7%) is higher than other species detected in clone libraries (0-3%), which may be indicative of multiple populations or cryptic or pseudo-cryptic species. Only a single genotype of *P. delicatissima* has been detected in Pacific Northwest samples, and this same genotype has been detected in isolates from Denmark and Portugal. We are currently generating clone libraries for additional basins in Puget Sound, and for samples from toxic *Pseudo-nitzschia* blooms, to further investigate the relationship between genetic composition, distribution, and the environment.

RECENT ECOSYSTEM SHIFT IN CENTRAL CALIFORNIA ALTERS HARMFUL ALGAL BLOOM PATTERNS

Rozalind Jester¹, Veronica Vigilant¹, Gregg Langlois² and Mary Silver¹

¹Ocean Science Department, University of California at Santa Cruz, Santa Cruz, California 95064, USA

²California Department of Health Services, Environmental Management Branch, 850 Marina Bay Parkway, Building G, Room 165, Richmond, California 94804, USA

In California, the toxic species of primary concern are the dinoflagellate *Alexandrium catenella* and members of the pennate diatom genus *Pseudo-nitzschia*, both producers of potent neurotoxins that have sickened and killed marine life and humans. During the summer of 2004 in Monterey Bay, we observed a dramatic change in the taxonomic structure of the phytoplankton community – the typically diatom-dominated community shifted to more of a red-tide, dinoflagellate-dominated community. Here we use a six-year time series (2000-2006) to show how the dominant harmful algal bloom (HAB) species in the Bay up to that point, *Pseudo-nitzschia*, was replaced as a major toxin producer by two genera of toxic dinoflagellates, *Alexandrium* and *Dinophysis*. This change represents a shift from a genus of toxin producers that typically dominate the community during a toxic bloom, to HAB taxa that need only be a minor component of the community to create a toxic event. This has significant implications for monitoring because toxic events are therefore not dictated by the relative dominance of a species. To strengthen that point, this change in the local HAB species was also reflected in the toxins present in higher trophic levels. Despite the small contribution of *A. catenella* to the overall phytoplankton community, the increase in the presence of this species in Monterey Bay was associated with an increase in the detection of paralytic shellfish toxins in shellfish and clupeoid fish. We also provide evidence, based on the statewide biotoxin monitoring program, that this increase in the frequency and abundance of *A. catenella* events occurred not just in Monterey Bay, but also in other coastal regions of California. Our results demonstrate that changes in the taxonomic structure of the phytoplankton community influences the nature of the algal toxins that move through local food webs and also emphasizes the importance of monitoring for the full suite of toxic algae, rather than just one genus or species.

DEVELOPMENT OF QUANTITATIVE PCR-BASED TECHNIQUES FOR ASSESSING ZOOPLANKTON GRAZING ON HARMFUL ALGAE: A TALE OF TWO SPECIES

Andrew R. Juhl¹, Sheean T. Haley², Rebecca R. Kelly¹, and Sonya T. Dyhrman²

¹Lamont-Doherty Earth Observatory, Columbia University, Palisades, NY, 10964, USA

²Woods Hole Oceanographic Institution, Woods Hole, MA, 02543, USA

Quantitative polymerase chain reaction (qPCR) techniques can be used to sensitively detect and quantify DNA of harmful algae. By using the amount of target DNA as a biomarker for cell number, the approach has been successfully used to quantify the concentration of single algal species in mixed field samples. Similarly, it may be possible to use qPCR to enumerate ingested cells of harmful algae in zooplankton gut contents. Eventually, a qPCR-based measure of gut contents could be used to derive predation rates on field populations of harmful algae. Such an approach offers several advantages over current techniques for measuring grazing on harmful algae and could increase sampling frequency in both space and time. However, detailed experimentation is required to ground truth the method prior to its use in the field.

We conducted laboratory grazing experiments using two model grazer species, adult *Acartia hudsonica* copepods, and *Nassarius* sp. veliger larvae, both fed *Alexandrium fundyense* (GTCA 28) as the harmful algal prey. DNA extraction procedures were optimized to ensure quantitative recovery of *Alexandrium* DNA. QPCR primers amplified a 183 base-pair fragment of large subunit rDNA. It has been tested against numerous algae and is specific for North American populations of *Alexandrium fundyense/tamarense/catenella* (Dyhrman et al. 2005). The detection limit is <1 cell and standard curves with known numbers of *Alexandrium* cells showed a linear signal response between 5 – 10,000 cells. *Alexandrium*-free extracts of either predator did not produce false positives when analyzed alone or PCR inhibition when added to *Alexandrium* standards.

When fed *A. fundyense* in the absence of other prey, a positive qPCR signal for *Alexandrium* was found in samples of 5-15 individual predators, indicating that a detectable amount of *Alexandrium* DNA survived ingestion and digestion. The *Alexandrium* signal in veliger larvae increased with the prey concentration, reaching 1-2 ingested cells per animal. In *A. hudsonica*, by contrast, the *Alexandrium* signal was detectable but exceedingly low, generally << 0.1 cell per copepod, even when other measures indicated that the copepods were eating >100 cells indiv.⁻¹ h⁻¹. Microscopic inspection of the copepods confirmed the presence of fecal material in their guts. However, samples of isolated fecal pellets and dissected gut contents also had very low *Alexandrium* signals. The veligers ingested *A. fundyense* at a relatively low rate but had a strong qPCR signal, while the copepods ingested *A. fundyense* at a high rate but had a barely detectable signal. We hypothesize that differences between the predators' ingestion and digestion mechanisms account for the differences in qPCR signal strength from ingested *Alexandrium*. In particular, copepods are known to mechanically damage large algal cells during ingestion, which may promote very rapid digestion of algal DNA. Based these early results, a qPCR-based measure of gut contents offers a specific and rapid approach for evaluating the impact of grazers on harmful algal blooms but it may be best suited for use with predators like veligers and some protists, that ingest algal prey whole.

DYHRMAN, S. T., D. L. ERDNER, J. LA DU, M. GALAC, AND D. M. ANDERSON. 2005. Molecular quantification of toxic *Alexandrium fundyense* in the Gulf of Maine using real-time PCR. Harmful Algae doi:10.1016/j.hal.2005.07.005.

THE OPTICAL-BASED HAB DETECTION OBSERVATORY: LESSONS LEARNED DURING 4 YEARS OF IMPLEMENTATION

Gary Kirkpatrick, Robert Currier, Cory Boyes, Jim Hillier, Alan Hails, Mike Miller, Augie Kotlewski, Karl Henderson and Brad Pederson
Mote Marine Laboratory, Sarasota, Florida, 34236

The first *in situ* BreveBuster, optical-based HAB detector, was deployed May 20, 2003. The first BreveBuster-equipped autonomous underwater vehicles began missions in November 2003. Fixed installations on buoys and channel markers took place in the summer of 2004. Presently, there are four AUVs and one vertical profiler equipped with BreveBusters, and seven regularly instrumented fixed sites. Current plans call for six more fixed installations in the eastern Gulf of Mexico and three near Vera Cruz, Mexico. By the end of the year there will be approximately 23 BreveBusters in existence. Besides the many engineering and funding complications that had to be addressed, there have been and continue to be many issues that must be dealt with on a day by day basis. For instance: Hurricane Charley removed our first fixed installation just weeks after it was deployed; cables, no matter how well secured, work loose on open water installations; satellite phone data telemetry can be interrupted by higher priorities; VHF radio telemetry rarely meets the manufacturers range specifications; installers and operators make mistakes; and of course there is biofouling. In addition to these 'mechanical' issues there are the issues of how to handle the information output. The internet provides a very functional dissemination infrastructure, but there have been many questions about who the end users should be and in what format should the information be presented. Our experiences with these issues will be itemized and our responses discussed.

GENE EXPRESSION IN ZEBRAFISH AFTER ACUTE AND SUB-ACUTE EXPOSURE TO THE MARINE NEUROTOXIN DOMOIC ACID

Kathi Lefebvre¹, Susan Tilton², Theo Bammler³, Richard Beyer³, Pat Janssen³, Fred Farin³, Sengkeo Srinouanprachanh³, Evan Gallagher²

¹Marine Biotoxins Program, NOAA Fisheries/Northwest Fisheries Science Center, 2725 Montlake Blvd. East, Seattle, WA 98125; ²Department of Environmental and Occupational Health Sciences, ³NIEHS Center for Ecogenetics and Environmental Health Functional Genomics and Bioinformatics Core Facilities, University of Washington

Domoic acid (DA) is a neuroexcitatory amino acid that is naturally produced by some species of the diatom genus *Pseudo-nitzschia*. The toxin accumulates in filter-feeding marine shellfish and is transferred through the food web resulting in a severe neurotoxic illness known as amnesic shellfish poisoning (ASP). Acute signs of ASP include vomiting, diarrhea, confusion, disorientation, seizures, memory loss, coma, and death. Dose-response relationships for the acute signs of DA-induced neurobehavioral excitotoxicity have been well defined for primate, rodent, and fish model species. However, little is known about the effects of sub-acute DA exposure (levels below those shown to induce overt toxicity). Furthermore, there is growing concern regarding the potential human health impacts of long-term low-level exposure to DA particularly in Washington State coastal Tribal communities that are dependant on shellfish for subsistence. In the present study, the zebrafish (*Danio rerio*) model was used to identify gene expression effects in the central nervous system (CNS) associated with acute and sub-acute DA exposure. Differential gene expression as evidenced by microarray analysis was observed in the brains of both sub-acute and acute treatments compared to controls. The observed gene expression patterns indicated that sub-acute DA exposure impacted the zebrafish CNS, and also that the mechanisms of DA toxicity may be different under conditions of acute and sub-acute DA exposure. Collectively, the dose-responses of DA-induced behavioral injury coupled with the microarray-generated gene expression data suggest that zebrafish are a useful model for exploring the mechanisms of chronic algal toxin exposure relevant to the vertebrate central nervous system.

SEASONAL AND INTERANNUAL VARIABILITY OF *Pseudo-nitzschia* AND DOMOIC ACID IN THE JUAN DE FUCA EDDY REGION AND ITS ADJACENT SHELVES

Evelyn J. Lessard¹, Barbara M. Hickey¹, William P. Cochlan², Charles G. Trick³, Mark L. Wells⁴, and Vera L. Trainer⁵

¹School of Oceanography, University of Washington, Seattle, WA 98195 USA

²Romberg Tiburon Center for Environmental Studies, San Francisco State University, Tiburon, CA 94920

³Department of Biology, University of Western Ontario, London, ONT N6A 5B7, Canada

⁴School of Marine Sciences, University of Maine, Orono, ME 04469 USA

⁵NOAA, Northwest Fisheries Science Center, Seattle, WA, 98112 USA

The Ecology and Oceanography of Harmful Algal Blooms of the Pacific Northwest (ECOHAB PNW) program was motivated by a hypothesized physical pathway between the Juan de Fuca Eddy, an apparent initiation site for toxigenic *Pseudo-nitzschia* blooms, and coastal clamming beaches on the Washington coast. Data from 6 cruises over 3 summers have shed light on the processes controlling the presence of domoic acid (DA)-producing *Pseudo-nitzschia* in the eddy region, those species responsible for toxic events, and the environmental parameters controlling bloom development. The field program used a combination of synoptic surveys and lagrangian drifter studies. The synoptic surveys provided information on the scales of variability and insights into the factors governing *Pseudo-nitzschia* and toxin occurrence. We identified more than 10 species of *Pseudo-nitzschia* off the WA coast, but those responsible for the most toxic blooms were *P. cuspidata* and *P. australis*. However, the presence of a particular species cannot be used as an absolute indicator of toxicity due to the high level of variability in DA production by field assemblages of *Pseudo-nitzschia*. Although sometimes achieving high abundances ($>10^6/L$), *Pseudo-nitzschia* spp. were always a small component of the total phytoplankton biomass, therefore satellite imagery is not useful for bloom prediction in this region. Throughout the study area, *Pseudo-nitzschia* usually occurred as a minor member of diatom-dominated assemblages; notably, it was often the major diatom taxa present in euglenoid and dinoflagellate blooms that occurred in the eddy region. Over the entire data set, no simple predictive relationship was found between environmental parameters (nitrate, phosphate, silicate, chlorophyll, temperature or salinity) and either *Pseudo-nitzschia* abundance, species or DA. Both *Pseudo-nitzschia* abundance and DA were highly variable in time and space. On a 21 day time scale, measurable toxin was always observed in the eddy region, and the highest level of toxin (>90 nM) and *Pseudo-nitzschia* cell numbers ($>11-13 \times 10^6/L$) were observed in an eddy bloom in September 2004. The field as well as modeling studies clearly demonstrated that toxic blooms can escape the eddy and move southward along the WA coast. On two occasions, toxin found in the coastal region was associated with the presence of Columbia River plume water. Our observations confirm the idea that the eddy region and not recently upwelled coastal water is the primary initiation site for most toxic blooms on the WA coast.

CHARACTERIZATION AND REGULATION OF GENE EXPRESSION NETWORKS IN RESPONSE TO ACUTE STRESS IN *Karenia brevis*

Kristy B. Lidie* and Frances M. Van Dolah

Biotoxins Program, NOAA Center for Coastal Environmental Health and Biomolecular Research

Karenia brevis is a dinoflagellate whose expressed genome is of significant interest because of its role in producing harmful algal blooms. The longevity of a *K. brevis* bloom is dependent on the cells' ability to adapt to changing conditions in the coastal environment. The induction of stress response genes has been shown at the protein level in *K. brevis*. To identify if this induction is controlled at the level of transcription initiation, we used a *K. brevis* microarray to measure changes in transcript abundance in response to acute stresses, including heat and oxidative stress. Consistent with a general stress response that includes a transient shut-off of general mRNA transcription, genes involved in ATP driven processes were downregulated following each of the treatments. However, transcription of stereotypical heat shock proteins and other stress related genes, known to be induced at the protein level in *K. brevis*, were not seen, implicating post-transcriptional regulation of these mRNAs.

There is precedence for widespread post-transcriptional control in dinoflagellates. Recently, we identified an RNA mediated *trans*-splicing mechanism in *K. brevis* that may play a role in this process. Consistent with the hypothesis that the stress response in *K. brevis* under post-transcriptional control, we found the SL sequence on many of the *K. brevis* stress response genes. To investigate control at the level of translation, polysomal fractionation was used to separate polysome bound mRNAs from the translationally inactive pool following peroxide stress. The microarray was then used to interrogate over-representation in the polysome fractions. We found many of the stress response genes, whose transcript levels remained constant, were present in the polysome bound fractions following peroxide treatment. Based on the results at the protein level, this suggests that the rate of protein synthesis may be accelerated via an increase in translational efficiency following environmental insult in *K. brevis*. This study provides the first comprehensive investigation into the stress responses networks present in a dinoflagellate responsible for harmful algal blooms (HABs).

* current affiliation is with Dr. Allen Place of COMB, Baltimore, MD

**GENETIC NETWORK REGULATING CELL DIVISION AND TOXIN PRODUCTION IN
Karlodinium AND *Amphidinium*: A GENOMIC APPROACH**

Senjie Lin¹, Huan Zhang¹, Allen Place², Jason Adolf², Terry Gaasterland³, Yu-Hui Rogers⁴, John Gill⁴ and Bao Tran⁴

¹Department of Marine Sciences, University of Connecticut, Groton, CT 06340, USA; ²Center of Marine Biotechnology, University of Maryland Biotechnology Institute Scripps Institution of Oceanography, Baltimore, MD 21202, USA; ³Scripps Institution of Oceanography, University of California at San Diego, La Jolla, CA 92093, USA; ⁴J. Craig Venter Institute, Rockville, MD 20850, USA

Karlodinium and *Amphidinium* are distinct dinoflagellate lineages but produce similar toxins. To understand what regulates population and toxin dynamics in these species, it is necessary to uncover biochemical or genetic network involved in the regulation. Given that their genomes are too large to sequence currently, analysis of expressed genes is the only way to gain information of this kind. Based on our recent discovery of widespread trans-splicing using a common splice-leader gene (Zhang et al. 2007), we undertook a project to sequence the full-length cDNAs for *Karlodinium veneficum* (CCMP 2778; 60k clones) and *Amphidinium carterae* (CCMP 1314; 30k clones) with a goal to assemble regulatory pathways for cell division and toxin production. Conditions necessary for production of billion cell cultures, optimization of cDNA library construction, and sequencing results will be presented. Insights into features of the dinoflagellate genome and the genetic network of cell division and toxin production will be discussed.

Zhang, H., Hou, Y., Miranda, L., Campbell, D. A., Sturm, N. R., Gaasterland, T. and Lin, S. 2007. Spliced leader RNA trans-splicing in dinoflagellates. *Proc. Natl. Acad. Sci. USA* 104: 4618-4623.

DEVELOPMENT OF A TOXIC DINOFLAGELLATE (*Karlodinium veneficum*) BLOOM IN A SHALLOW, EUTROPHIC, LAGOONAL ESTUARY

R. Wayne Litaker¹, Nathan S. Hall², Elizabeth E. Fensin³, Jason E. Adolf⁴, Allen R. Place⁴, Hans W. Paerl²

¹National Oceanographic and Atmospheric Administration, National Ocean Service, Beaufort, NC 28516

²Institute of Marine Sciences, University of North Carolina at Chapel Hill, Morehead City, NC 28557

³Division of Water Quality, North Carolina Department of Environment and Natural Resources
Raleigh, NC 27699-1621

⁴Center of Marine Biotechnology, University of Maryland Biotechnology Institute, Baltimore, MD 21202

A dense bloom of the ichthyotoxic dinoflagellate *Karlodinium veneficum* was discovered in the Neuse River Estuary, NC on 19 October, 2006 and was associated with four subsequent fish kills. This bloom was fostered by runoff following tropical storm Ernesto which input particulate and dissolved nutrients. Initially, runoff lead to increased flushing, light limited productivity and low algal biomass. As riverine discharge declined and the water column stabilized a surface frontal zone with lowered dispersion rates favorable for biomass accumulation became established. At the same time a prolonged period of low wind allowed vertical stratification and development of hypoxic bottom conditions that produced the among the highest hypolimnetic concentrations of remineralized NH_4^+ ever measured in the estuary. A brief wind event mixed regenerated nutrients throughout the water column. A subsequent period of stable runoff, calm winds and a highly stratified water column provided salinity, light, nutrient and hydrologic conditions ideal for phytoplankton growth. The resultant community became dominated by dinoflagellates, the most successful of which was the mixotroph *K. veneficum* (>200,000 cells/ml, 740 ng / ml karlotoxin). The success of this species is probably due to its ability to produce the karlotoxin KmTx2, which aids in the capture of algal prey during mixotrophic feeding and in deterring microzooplankton grazers. Once the bloom was established, small-scale estuarine physical processes coupled with vertical migration behaviors acted to further concentrate cells. The bloom demise was linked to disruption of an already senescing population by a turbulent wind mixing event. Toxin released from these cells was postulated to be the cause the concurrent fish kills. Data that supports this assumption includes likely movement of the disrupted bloom into the fish kill area, the presence of *K. veneficum* at the kill sites, and the characteristic premortem symptoms of karlotoxin poisoning which

include air gulping and lethargy despite high ambient DO conditions. This bloom underscores the important linkages between meteorological forcing and benthic-pelagic coupling in fostering phytoplankton blooms in shallow estuaries.

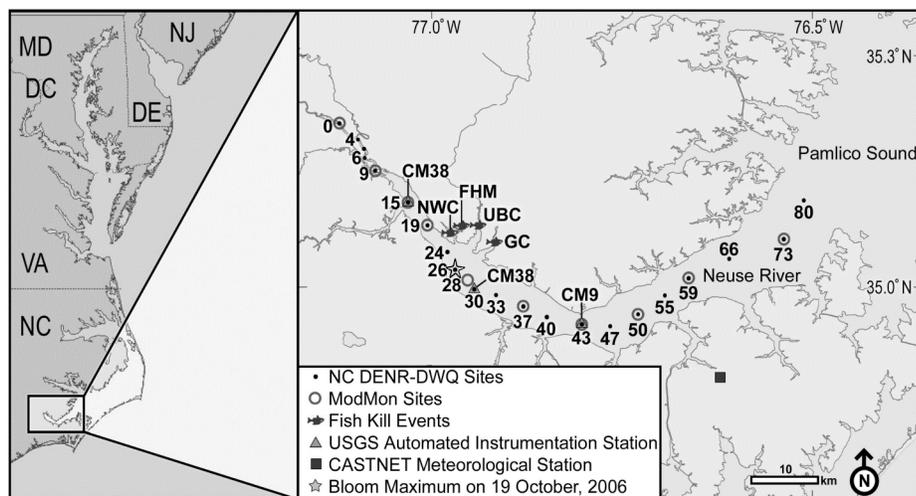


Fig. 1. Map of the Neuse River Estuary showing locations of sampling stations and

documented fish kill events. NWC= North-west Creek. FHM= Fairfield Harbor Marina. BC= Broad Creek. GC= Goose Creek. Water quality station identifiers are expressed as km downstream. USGS automated instrument-ation stations are labeled by the channel marker (CM) to which they are fixed.

OBSERVATIONS AND MODELS OF *Alexandrium fundyense* BLOOMS IN THE GULF OF MAINE AND GEORGES BANK: FROM CLIMATOLOGY TO FORECASTING

Dennis J. McGillicuddy, Jr.¹, Ruoying He², Keston W. Smith¹, Donald M. Anderson¹, and Bruce A. Keafer¹

¹Woods Hole Oceanographic Institution, Woods Hole, MA 02540, USA

²North Carolina State University, Raleigh, NC 27695, USA

Coupled physical-biological models are used to study fluctuations in *Alexandrium fundyense* populations in the Gulf of Maine in a) climatological and b) data-assimilative modes.

Climatological distributions of *A. fundyense* are investigated through simulations of population dynamics of *A. fundyense* within the seasonal mean flow. A model that includes germination, growth, mortality, and nutrient limitation is qualitatively consistent with observations. Cells germinated from cyst beds in the Bay of Fundy and near Penobscot and Casco Bays are advected from east to west in the coastal current. Growth of vegetative cells is limited by temperature from April through June throughout most of the region, whereas nutrient limitation occurs in July and August in the western gulf.

The coupled model is used to hindcast the historic 2005 *A. fundyense* bloom and to investigate the relative importance of factors governing initiation and development. Sensitivity experiments distinguish the roles of three major factors hypothesized to contribute to the bloom: 1) the high abundance of cysts in western GOM sediments; 2) several strong storms with prevailing downwelling favorable winds; and 3) anomalously large fresh water discharge entering the gulf due to abundant rainfall and heavy snowmelt. Our results suggest that high abundance of cysts in western GOM was the main cause of the 2005 bloom. Wind forcing was an important regulator, as episodic bursts of northeast winds caused onshore advection of offshore populations. These downwelling favorable winds also accelerated the along-coast flow, resulting in transport of high cell concentrations into Massachusetts Bay. Anomalously high river runoff in 2005 resulted in stronger buoyant plumes/currents, but had limited impact on the gulf-wide bloom distribution.

Initial results from 2006 and 2007 will also be discussed, emphasizing comparisons between observations and forecast/hindcast predictions.

NOVEL STRUCTURE OF POLYKETIDE SYNTHASE GENE TRANSCRIPTS IN THE FLORIDA RED TIDE DINOFLAGELLATE, *Karenia brevis*

Emily A. Monroe^{1,2} and Frances M. Van Dolah^{1,2}

¹Marine Biotoxins Program, NOAA Center for Coastal and Environmental Health and Biomolecular Research, Charleston, SC 29412

²Marine Biomedicine and Environmental Sciences, Medical University of South Carolina, Charleston, SC 29412

Karenia brevis is the Florida red tide dinoflagellate responsible for detrimental effects on human and environmental health through production of brevetoxins. Brevetoxins are polyketide compounds thought to be synthesized by a modified polyketide synthase (PKS) complex, but the gene cluster for this PKS has yet to be identified. Eight PKS transcripts were identified in *K. brevis* by high throughput screening of two *K. brevis* cDNA libraries. However, because axenic cultures are unavailable, the origin of PKS transcripts remains controversial. Through phylogenetic analysis of PKS transcripts encoding ketosynthase domains and the presence of a dinoflagellate – specific spliced leader sequence, these transcripts have been confirmed to be encoded by *K. brevis*. Identification of the spliced leader at the 5' end of the PKS transcripts indicates that the sequences described are full-length transcripts, which was further confirmed by northern blot analysis. Although most similar to type I modular PKS, sequence analysis determined that seven of the transcripts encode single catalytic domains, six KS domains and one KR domain. This is the first study to describe full-length PKS transcripts in a dinoflagellate and identifies a novel PKS organization, with sequence most similar to type I modular PKS, but structure most similar to type II.

OBSERVATIONS ON THE EPIPHYTIC RELATIONSHIP BETWEEN *Gambierdiscus* SPP. AND SEVERAL MACROALGAL HOST SPECIES

Michael L. Parsons¹, Chelsie J. Settlemier², and Josh M. Ballauer²

1 – Department of Marine and Ecological Sciences, Florida Gulf Coast University, Fort Myers, FL 33928

2 – Marine Science Department, University of Hawaii at Hilo, Hilo, HI 96720

Specimen of twenty six different macroalgal species were collected, rinsed, and placed in petri dishes containing sterile, modified Keller's media and monoclonal *Gambierdiscus* cells (approximately 110 per dish). The dishes were placed in an incubator at 27C, 12 L/D, 100 µE, and examined periodically over a one-month period to count the number of attached, unattached, and dead *Gambierdiscus* cells. *Gambierdiscus* cells displayed one of three attachment modes, depending on the host species: no attachment at all (e.g., *Portieria hornemannii*), attachment in the early stages of the experiment, followed by unattachment (*Jania* sp.), or unattachment followed by later-stage attachment (*Dasya* sp. 1). *Portieria hornemannii* was the least favorable host, with zero *Gambierdiscus* cells attaching over the course of the study, the highest *Gambierdiscus* mortality (68%), the quickest time to 50% mortality (168 hours), and the fewest live cells (maximum of 34 cells). *Chaetomorpha* sp., *Galaxuara marginata*, *Jania* sp., and an unidentified cyanobacteria all exhibited good host characteristics. *Chaetomorpha* sp. had the lowest *Gambierdiscus* mortality rate (0.8%), whereas *G. marginata* had the maximum number of *Gambierdiscus* cells observed (4,532 at the end of the experiment). *Jania* sp. had the highest average number of cells over the course of the study (1,189), and the second highest average and maximum % attached cells (17 and 57%, respectively), with the unidentified cyanobacteria exhibiting the highest average and maximum % attached cells (33 and 73%, respectively). These results demonstrate that 1) some macroalgae species are better hosts than others (corroborating published field results); 2) attachment is not simply a matter of macroalgae morphology, but that chemical exudates likely play a significant role (corroborating published results); and 3) the composition of the chemical exudates likely changes as a macroalgal host proceeds through its life cycle and subsequent death phase.

DISSOLVED DOMOIC ACID: A COMPETITIVE ADVANTAGE FOR *Pseudo-nitzschia* IN COASTAL AND OFFSHORE HNLC WATERS

Lisa D. Pickell¹, Mark L. Wells¹, and Charles G. Trick²

¹Darling Marine Center, University of Maine, Walpole, Me 04573, USA

²Schulich School of Medicine and Dentistry, The University of Western Ontario, London, Ontario, N6A5B7, Canada

The toxigenic diatom genus *Pseudo-nitzschia* is known to release variable amounts of the potent neurotoxin, domoic acid (DA) during both exponential and senescent growth stages. The environmental conditions that regulate DA production and release by *Pseudo-nitzschia* are poorly understood but recent evidence shows that it can be associated with micronutrient (Fe, Cu) limitation. Because DA is a charged molecule, its release from exponentially growing cells implies an active transport process, which in turn, suggests dissolved DA provides a benefit to the cell. We investigated the effects of elevated dissolved DA concentrations on the species composition of natural population cultures in coastal waters of the Pacific Northwest and in offshore high nitrate low chlorophyll waters of the subarctic Pacific. These experiments used a novel sea-going continuous culture system that accentuates differences in growth adaptations of the community by selectively “washing” out those organisms having lower growth rates, thereby providing unique insights to bottom-up control of phytoplankton community trajectories. In both coastal and offshore regimes, overall chlorophyll biomass increased significantly with the addition of either DA or DA bound to Cu, a hypothesized purpose for DA release (Wells et al. 2005). Moreover, community analyses showed that growth of *Pseudo-nitzschia* spp. was strongly selected over that of other diatoms present, resulting in an overwhelming dominance of *Pseudo-nitzschia* in these treatments. This finding was consistent in both coastal and offshore waters indicating it may be a universal pattern. To our knowledge these findings provide the first direct evidence that DA is produced and released by *Pseudo-nitzschia* as a dissolved tool to facilitate its competition for growth. By implication then, it may follow that increased cell toxicity may signal environmental conditions where there is less cellular need to deploy the internal reserves of this competitive tool.

WELLS, M. L., C. G. TRICK, W. P. COCHLAN, P. HUGHES, and V. L. TRAINER. 2005. Domoic Acid: The synergy of iron, copper and the toxicity of diatoms. *Limnology and Oceanography* **50**: 1908-1917.

SCRABBLED MODULES, SPLICED LEADERS, CAP DEPENDENT TRANSLATION CONTROL – WHAT NEXT IN DINOFLAGELLATE POLYKETIDE TOXIN SYNTHESIS?

Allen R. Place¹ and Tsvetan R. Bachvaroff¹

¹UMBI Center of Marine Biotechnology, 701 E. Pratt St., Baltimore, MD USA 21202

Polyketide synthases (PKSs) and nonribosomal peptide synthetases (NRPs) catalyze chain elongation from simple building blocks to create a diverse array of natural products including algal toxins. Modular PKS and NRPS proteins consist of one or more polyfunctional polypeptides, each of which is made up of as few as 1 or as many as 11 modules, with the amino-terminal to carboxy-terminal order of the individual modules paralleling the sequential order of polyketide or nonribosomal polypeptide chain elongation. In contrast, the corresponding structural genes for discrete PKS or NRPS open reading frames may be transcribed in an order that directly parallels the sequence of the eventual biosynthetic steps, as is the case for the erythromycin PKS. Each PKS or NRPS module is made up of a set of three mandatory or core domains, two of which are catalytic and one which acts as a carrier, that together are responsible for the central chain-building reactions of polyketide or polypeptide biosynthesis, as well as a variable set of auxiliary domains that mediate the modification (e.g. reduction, dehydration and methylation) of the newly extended polyketide or polypeptide chain. Based on the recent cloning of the putative PKS genes involved in amphidinolide synthesis by Koboyashi's group, the dinoflagellates break all known PKS paradigms. Not only do the genes not contain three mandatory core domains but the genes are not tandem arrangements but disperse over nearly 10,000 bp of genomic sequence. Our own work with *Amphidinium carterae* found a keto-reductase (KR) module dispersed over nearly 10,000bp of genomic sequence with frequent introns and no apparent nearest neighbor PKS modules. Perhaps an operon arrangement for transcriptional control of these genes is not needed given the recent discovery of trans-splicing in dinoflagellates (Zhang et al. 2007, Lidie and Van Dolah, 2007). All transcripts are treated equally and just need to have the appropriate mRNA CAP structure to be recruited for translation. We will present our current attempt to characterize the 5' CAP structure in dinoflagellates and how this might relate to constitutive expression of PKS genes.

Kubota, T., Iinuma, Y. and Koboyashi, J. 2006. Cloning of polyketide synthase genes from Amphidinolide-producing dinoflagellate *Amphidinium* sp. *Bio. Pharm. Bull.* 29(7): 314-138.

Zhang, H., Hou, Y., Miranda, L., Campbell, D. A., Sturm, N. R., Gaasterland, T. and Lin, S. 2007. Spliced leader RNA trans-splicing in dinoflagellates. *Proc. Natl. Acad. Sci. USA* 104: 4618-4623.

Lidie, K.B and Van Dolah, F.M. (2007) Spliced leader RNA-mediated trans-splicing in a dinoflagellate, *Karenia brevis* *J. Eukaryotic Microbiology* (In Press).

MONITORING OF BREVETOXINS IN *Karenia brevis* BLOOM-EXPOSED EASTERN OYSTER

Steven Plakas¹, Edward Jester¹, Kathleen El Said¹, Hudson Granade¹, Ann Abraham¹, Robert Dickey¹, Paula Scott², Leanne Flewelling², Michael Henry³, Patricia Blum³, Richard Pierce³

¹U.S. FDA, Gulf Coast Seafood Laboratory, Dauphin Island, AL 36528, USA

²Florida Fish and Wildlife Conservation Commission, Fish and Wildlife Research Institute, St. Petersburg, FL 33701, USA

³Mote Marine Laboratory, Sarasota, Florida 34236, USA

Brevetoxins are produced by the marine alga *Karenia brevis* and accumulated in filter-feeding molluscan shellfish. Consumption of brevetoxin-contaminated shellfish causes neurotoxic shellfish poisoning. Previous studies in Eastern oyster (*Crassostrea virginica*) established the extensive metabolism of algal brevetoxins and the persistence of brevetoxin metabolites after controlled exposures to *K. brevis* culture. The present study examines uptake and persistence of brevetoxins and metabolites in Eastern oyster exposed to recurring blooms of *K. brevis* at a single study site in Sarasota Bay, FL, over a three-year period. Brevetoxins were monitored by several methods, both structure-based (LC-MS, ELISA) and pharmacology-based (cytotoxicity assay, receptor binding assay). Toxicity of shellfish was assessed by traditional mouse bioassay. Measurements by all methods reflected well the progression and magnitude of the blooms. Shellfish toxicity above the guidance level (20 mouse units/100 g) was associated with cell densities >5,000 cells/L, and retained for up to two weeks after bloom dissipation. By in vitro methods, brevetoxins were measurable for several months after a bloom. By using LC-MS, we find that brevetoxins persist in the field-exposed oyster mainly as metabolites of the algal brevetoxins PbTx-1 and PbTx-2, as previously established in laboratory studies. Most abundant of the brevetoxin metabolites in shellfish were cysteine and cysteine-sulfoxide conjugates of PbTx-2; lesser amounts of PbTx-1 metabolites reflect the lower abundance of parent toxin in the bloom water. Levels of PbTx-2 conjugates as determined by LC-MS were highly correlated ($r^2 = 0.9$) with composite toxin measurements by ELISA (antibody directed against B-type brevetoxin backbone). LC-MS and ELISA values also correlated well ($r^2 = 0.7$) with those of mouse bioassay. Cytotoxicity and receptor binding assays did not correlate as well. ELISA and LC-MS methods offer rapid screening and confirmation, respectively, of brevetoxin contamination in Eastern oyster, and an alternative to mouse bioassay in assessing toxicity following *K. brevis* blooms.

CHEMICALLY-MEDIATED COMPETITION: INTERACTIONS BETWEEN THE RED TIDE DINOFLAGELLATE, *Karenia brevis*, AND CO-OCCURRING PHYTOPLANKTON

Prince, E.K. Georgia Institute of Technology, Atlanta, GA, gtg982j@mail.gatech.edu

Myers, T.L. Georgia Institute of Technology, Atlanta, GA, tm192@mail.gatech.edu

Naar, J. University of North Carolina at Wilmington, Wilmington, NC, naarj@uncw.edu

Kubanek, J. Georgia Institute of Technology, Atlanta, GA, julia.kubanek@biology.gatech.edu

The red tide dinoflagellate *Karenia brevis* blooms seasonally and often nearly monospecifically in coastal areas of the Gulf of Mexico, producing neurotoxic brevetoxins which kill fish and marine mammals. However, the mechanism that *K. brevis* uses to dominate the phytoplankton community is not well understood. We considered how *K. brevis* interacts with competing phytoplankton species: whether it releases inhibitory chemical compounds to suppress competitors, what effect these compounds have on competitor growth and physiology, and how competitors respond to compounds produced by *K. brevis*. We found that compounds exuded during *K. brevis* blooms and in *K. brevis* cultures inhibited the growth and lowered the photosynthetic efficiency of competing phytoplankton. Exudates of *K. brevis* cultures also increased membrane permeability of three phytoplankton species. Compounds produced during *K. brevis* blooms were allelopathic to the diatom *Skeletonema costatum*, but *K. brevis* bloom samples lost their allelopathic effect when exposed to *S. costatum*, indicating that *S. costatum* possesses a mechanism for undermining *K. brevis* allelopathy. Efforts to isolate and identify the responsible compounds are ongoing. Our results indicate that although *K. brevis* produces potent allelopathic compounds, competitors vary in their susceptibility and resistance strategies, and competitive interactions in the marine plankton appear to be complex, multi-directional, and part of an ongoing co-evolutionary battle over limiting resources.

A POTENT EFFECT OF IN VITRO GASTRIC DIGESTION ON THE OVERALL TOXICITY OF BREVETOXIN-LADEN ATLANTIC MENHADEN (*Brevoortia tyrannus*)

Faisal F. Y. Radwan, M.-Yasmine Bottein Dechraoui, Zhihong Wang, John S. Ramsdell
Marine Biotoxins Program, Center for Coastal Environmental Health and Biomolecular Research,
NOAA-National Ocean Service, Charleston, SC 29412, USA.

Brevetoxins, produced by dinoflagellate *Karenia brevis*, have increasing impact on marine biodiversity, coastal health and economy. These toxins were responsible for severe dolphin fatalities. A large dolphin mortality event was recorded in March and April of 2004, in which 107 bottlenose dolphins found dead along the Florida panhandle, although neither an observed algal bloom nor toxin present in water samples. Analysis of stomach contents, suggested that these animals have died of gorging enormous amount of toxin-exposed Atlantic menhaden fish, *Brevoortia* sp^{1,2}. Although high levels of toxin were reported in undigested menhaden in the dolphin stomach contents, the total toxin load appears substantially less than what would be projected to be a lethal dose, based upon oral toxicity studies of purified toxin in mice. In light of the above, we investigated the effect of gastric digestion on the brevetoxin composition and toxicity of Atlantic menhaden exposed to cultures of *K. brevis*. Viscera were submitted to an *in vitro* digestion experiment in presence of a synthetic or a natural gastric juice collected from a stranded bottlenose dolphin (*Tursiops truncatus*). Brevetoxins were extracted from the gastric suspensions and alterations in toxin composition were examined using liquid chromatography / mass spectrometry (LC/MS) and radioimmunoassay (RIA). Alterations in toxin activity were measured using receptor binding assay (RBA) and Neuro-2A cytotoxicity assay (N2A). Unlike the undigested extracts, *in vitro* digestion produced a dramatic increase in biological activity in parallel with depletion of the predominate brevetoxins and cysteine conjugate metabolites. LC / assay guided fractionation of the post-digestion toxic products revealed that the peaks of biological activity are mostly due to unknown hydrophobic metabolites, which still remain to be determined. Taken together, our report suggests that gastric digestion play a significant role on the quantity and the composition of brevetoxins which are bioaccessible for further intestinal absorption. Digestion may trigger a brevetoxin transformation along with a possible discharge of more potent lipophilic brevetoxin conjugates extracted by the digestion process.

⁽¹⁾ NMFS [National Marine Fisheries Service] Interim Report 2004. <http://www.nmfs.noaa.gov/> ⁽²⁾ Flewelling *et al.* *Nature* **435**, 755-756 (2005).

FEATURES OF NEUROTOXIC SHELLFISH POISONING FROM RECREATIONALLY HARVESTED CLAMS IN FLORIDA, 2006: EPIDEMIOLOGIC AND CLINICAL FACTORS

Andy Reich¹, Sharon M. Watkins¹, Robert South², Robin Terzagian¹, Roberta Hammond¹, Carina Blackmore¹, Jan Landsberg³, Leanne Flewelling³, Steve M. Plaky, Ann Abraham, Robert W. Dickey⁴.
¹Florida Department of Health, Tallahassee, FL 32399; ²Lee County Public Health Department, Fort Myers, FL 33916; ³Fish and Wildlife Research Institute, Florida Fish and Wildlife Conservation Commission, St. Petersburg, Florida 33701, ⁴United States Food and Drug Administration, Center for Food Safety and Applied Nutrition, Gulf Coast Seafood Laboratory, Dauphin Island, Alabama 36528

A series of Neurotoxic Shellfish Poisoning (NSP) cases occurred along the south-west coast of Florida during 2006. NSP is caused by consumption of shellfish contaminated with brevetoxins originating from the marine dinoflagellate, *Karenia brevis*. *K. brevis* blooms cause the phenomenon known as red tide, a recurrent environmental hazard in the Gulf of Mexico. Florida state agencies routinely sample coastal waters for *K. brevis*; testing of regulated shellfish beds has been in place since the 1970's.

Descriptions of NSP cases are rare. In this series, 8 clusters representing 17 individuals were identified. All cases were associated with consumption of recreationally-harvested clams from coastal areas not officially approved or open for harvesting. Most (n=15) cases sought medical attention and treatment at surrounding hospitals. A variety of gastrointestinal and neurological symptoms were reported; neurological symptoms being the predominant clinical presentation. Incubation was from 1-6 hours, symptom duration was a few hours to 2 days, and 7 cases were admitted after emergency room presentation. Available samples of clams from case meals, implicated shellfish beds, and urinalysis from cases were positive for brevetoxins.

Most cases were tourists of Asian ethnicity who were unaware of shellfish bed closures. Increased public health warnings, including public services announcements, radio interviews and door-to-door reverse 911 contacts were initiated, cases abated after July. Despite local knowledge about red tides, routine monitoring of *K. brevis* cell counts in seawater, successful monitoring of state-regulated shellfish beds and local dissemination of shellfish-risk information; education of transient groups such as tourists remains a challenge.

**A COMPARATIVE STUDY OF PERCEIVED RISK FROM TWO COASTAL COMMUNITIES:
IMPLICATIONS FOR COMMUNICATION AND EDUCATION**

Sparkle Roberts¹, Lynn M. Grattan^{1,2}, Kate Tracy,² Jessica Rowe¹, Stephan Parker¹, J. Glenn Morris².

¹ Department of Neurology, University of Maryland School of Medicine, Baltimore, MD USA 21201

² Department of Epidemiology and Preventive Medicine, University of Maryland School of Medicine, Baltimore, MD USA 21201

Risk perception is the characteristic way individuals approach, think about and interpret environmental risks and hazards. An individual's perception of risk guides their response to environmental threats. Moreover, it provides a framework for interpreting medical symptoms or illnesses. Since perceived risk is not always correlated with actual risk, obtaining risk perception data for a community is important for effective public health communication, education and intervention programs. Toward this end, we studied the perceived risk of harmful algal blooms (HABs) and HAB-related illnesses in a coastal community in the Mid-Atlantic States (Maryland Eastern Shore) and the Pacific Northwest (Northwest Washington). Since coastal communities often differ in history, culture and environmental hazards, it was hypothesized that they would differ with respect to the nature and extent of environmental worry, environmental health knowledge, and credible sources for information. Approximately 500 persons from the Maryland Waterman community (MD group) and seafood dependent tribal coastal communities in Washington (WA group) were studied with a standard risk perception questionnaire. The questionnaire was comprised of 17 questions in Likert-type or rank-order format regarding environmental worry, environmental health knowledge and sources for reliable HAB-related information. Findings indicated that although both communities worried about HAB's, there were significant differences in many areas. This included differences in how the groups perceived the effectiveness of scientists and where they would turn for reliable information. The WA group was more confident than the MD group that scientists would find a solution to their HAB problems. Moreover, they were more likely to turn to state and Tribal agencies for accurate HAB-related information. In contrast, the Maryland group had significantly more environmental anxiety and was more likely to turn to the media for reliable HAB-related information. The results support the need to take into consideration specific community perceptions and behavioral tendencies in the development of community and culturally-appropriate communication pathways.

ENVIRONMENTAL FACTORS ON MICROCYSTIN CONCENTRATIONS IN LAKES

Orlando Sarnelle, Howard Wandell, and Jamie Morrison

Department of Fisheries and Wildlife, Michigan State University, East Lansing, MI

Invasion of Midwestern lakes by the zebra mussel (*Dreissena polymorpha*) has altered the prevailing limnological paradigm of a positive influence of nutrient loading on cyanobacterial dominance (Raikow et al. 2004, Sarnelle et al. 2005). A multi-lake survey by our group (Knoll et al. *in press*) has indicated an average 3X elevation in microcystin (toxin) concentrations lakes that have been invaded by the zebra mussel, but this survey was limited to lakes with total phosphorus concentrations less than 20 $\mu\text{g L}^{-1}$. Microcystin concentrations in the mixed layer were highly correlated with the biomass of *Microcystis aeruginosa*, but not with the biomass of *Anabaena*. As a follow up, we enlisted citizen monitors to sample 75 lakes distributed across broad environmental gradients throughout the state of Michigan. Preliminary analysis has revealed a positive influence of total phosphorus concentrations on microcystin concentrations (as measured by ELISA), but only for lakes lacking zebra mussels. The influences of lake morphometry and mean annual air temperature (based on lake location) will also be examined to increase predictive power and suggest the impacts of global warming.

Knoll, L. B., O. Sarnelle, S. K. Hamilton, C. E. H. Scheele, A. E. Wilson, J. B. Rose, and M. R. Morgan. *in press*. Invasive zebra mussels (*Dreissena polymorpha*) increase cyanobacterial toxin concentrations in low-nutrient lakes. *Canadian Journal of Fisheries and Aquatic Sciences*.

Raikow, D. E., O. Sarnelle, A. E. Wilson, and S. K. Hamilton. 2004. Dominance of the noxious cyanobacterium *Microcystis aeruginosa* in low nutrient lakes is associated with exotic zebra mussels. *Limnology and Oceanography* **49**:482-487.

Sarnelle, O., A. E. Wilson, S. K. Hamilton, L. B. Knoll, and D. E. Raikow. 2005. Complex interactions between the zebra mussel, *Dreissena polymorpha*, and the harmful phytoplankter, *Microcystis aeruginosa*. *Limnology and Oceanography* **50**:896-904.

TOXIC BLOOMS OF *Pseudo-nitzschia* spp. AND THEIR IMPACT ON COASTAL MARINE LIFE IN THE SOUTHERN CALIFORNIA BIGHT AREA NEAR LOS ANGELES

Astrid Schnetzer¹, Lauren Palmer², Susan Kaveggia³, Richard Evans⁴, Michele Hunter⁴, Burton Jones¹, Ivona Cetinic¹, Elizabeth Fitzpatrick¹, Gloria Song¹, Peter E. Miller⁵, Rebecca Schaffner⁶, Steve Weisberg⁶ and David A. Caron¹

¹University of Southern California, Los Angeles, CA 90089

²Marine Mammal Care Center at Fort McArthur, San Pedro, CA 90731

³California Council for Wildlife Rehabilitators, Santa Rosa, CA 95402

⁴Pacific Marine Mammal Center, Laguna Beach, CA 92651

⁵University of California Santa Cruz, Santa Cruz, CA 95064

⁶Southern California Coastal Water Research Project, Costa Mesa, CA 92626

Blooms of *Pseudo-nitzschia* spp. and domoic acid concentrations in the plankton have been documented in coastal waters near the Los Angeles metropolitan area on an annual basis since 2003. Field observations have demonstrated that major bloom characteristics such as temporal bloom dynamics or level of toxicity varied considerably among years, but general trends were also apparent for coastal waters along Los Angeles and Orange County shorelines: 1) Particulate domoic acid concentrations commonly exceeded those reported from other geographical areas (ie, 2003, 2006 & 2007) indicating that regional environmental conditions particularly favor *Pseudo-nitzschia* and domoic acid production. 2) Highest concentrations of domoic acid within the study area ($\approx 400 \text{ km}^2$) repeatedly occurred inside or immediately adjacent to the breakwater of the Los Angeles harbor (2003, 2006 & 2007). 3) Local *Pseudo-nitzschia* blooms impacted a wide array of species through food web interactions. The latter observation was based on domoic acid detection and quantification in a large number of animal samples collected from marine mammals and seabirds (body fluids, stomach contents and feces). In 2006 and 2007 eleven coastal species that had not been implicated previously tested positive for the algal toxin. The spatiotemporal relationships between bloom dynamics and physiochemical parameters were examined.

THE FATE OF SAXITOXINS IN *Alexandrium tamarense* DURING INFECTION BY *Amoebophrya* sp., AND INITIAL OBSERVATION OF HOST-PARASITE DYNAMICS FROM FIELD STUDIES IN A SMALL CAPE COD EMBAYMENT

Mario R. Sengco¹, David M. Kulis², D. Wayne Coats¹ and Donald M. Anderson²

¹Smithsonian Environmental Research Center, Edgewater, MD 21037, USA

²Woods Hole Oceanographic Institution, Woods Hole, MA 02540, USA

Laboratory studies were conducted to examine the fate of saxitoxins in *Alexandrium tamarense* (strain SPE10-1) during the course of infection by the parasitic dinoflagellate, *Amoebophrya* sp. Host cultures were grown in f/2-Si medium until they reached late exponential phase. Parasite dinospores of known age were harvested through gravity filtration. Small volumes of host culture were placed in duplicate scintillation vials to which dinospores were added at a 10:1 parasite-to-host ratio. A parallel set of duplicate vials were used as controls (i.e. no parasites). Subsamples were taken every 12 hrs from 0 (initial conditions) to 96 hrs. The samples were processed to determine host abundance, dinospore abundance, parasite prevalence and toxin content. For the infected cultures, host abundance decreased steadily over the course of the study. Dinospore abundance initially decreased as the cells entered the host. This was followed by a small increase in dinospore abundance at 36 hrs and large increase at 72 hrs. Parasite prevalence reached 49% at 36 hrs. Host abundance in the controls was unchanged during the experiment. There was no significant difference in total toxin content per cell between the infected cultures and controls. Similarly, there were no significant differences in the toxin profiles per cell (i.e. quantity and quality of each toxin) between infected and control cultures during the course of the study. There was a decrease, however, in the total amount of toxin in the infected *culture* relative to the uninfected controls, which corresponded to the decrease in the number of hosts as the infection proceeded. The fate of these toxins – whether they were destroyed by the developing parasite, remained intact within the emerging dinospores, and/or released into the medium following host lysis – is currently under investigation.

To understand the role of *Amoebophrya* sp. in controlling an *Alexandrium* sp. population in the field, studies were conducted in Salt Pond (Eastham, MA) during the spring bloom in 2006 and 2007. Integrated water samples were collected weekly prior to the bloom to monitor host abundance. As host density increased to > 1,000 cells/L, integrated water samples were collected every two or three days. For greater spatial resolution, water samples were also collected at three locations across the length of the pond from the surface down to depth at 1-m intervals using a submersible pump. The water samples were preserved and analyzed for host abundance. Parasite prevalence was examined using microscopic observations, fluorescent in situ hybridization (FISH) probes, and quantitative protargol staining. Preliminary results from the 2007 study showed the initiation, development and decline of the host population during which three peaks in host abundance were seen. The second and third peaks were followed by two peaks in parasite prevalence based on FISH probing and microscopic observations. An initial peak in parasite prevalence following the first peak in host abundance was found only in microscopic observation, which suggests the presence of another parasite that affects *Alexandrium* sp. cells that was not detected by the probes. Protargol staining and other analyses are currently being applied to characterize this further. The potential implications of these data on the role of *Amoebophrya* sp. in controlling *Alexandrium* sp., naturally or as a possible control agent, will be discussed.

CAN BENTHIC-PELAGIC COUPLING BY *Karenia brevis* SUPPORT PERENNIAL OFFSHORE SEED POPULATIONS FOR COASTAL BLOOMS?Geoffrey A. Sinclair¹, Daniel Kamykowski¹¹ North Carolina State University, Raleigh, NC, 27695, USA

Multiple hypotheses have been presented to explain how nutrients are delivered to populations of *Karenia brevis* in oligotrophic water columns in the Gulf of Mexico. Vertical migration behavior coupled with the physiology of *K. brevis* may help alleviate bottom-up controls and permit populations to persist as vegetative cells near the sediment-water interface throughout the year. Aggregations of natural *Karenia brevis* populations near the sediment-water interface suggest that cells may derive nutrients from the sediment in oligotrophic water columns. How cells interact with the sediment, however, remains uncertain. Video of cells near the sediment-water interface suggest that cells may either access nutrients that flux out of the sediment or migrate into the sediment pores where higher nutrient concentrations exist. Experiments to test the ability of *K. brevis* to migrate into the sediment were devised using chambers divided by a 100 μm mesh overlain with a thin layer of sediment collected from the Gulf of Mexico. Since the diel vertical migration of *K. brevis* typically displays a nocturnal descent, experiments tested migration response during the day and night and with and without a sub-sediment nutrient source. In order to determine the ability of *K. brevis* to exploit elevated nutrients associated with the sediment, we examined the diel rates of uptake and assimilation of different nitrogen substrates (NH_4^+ , NO_3^- , Urea). Uptake and assimilation rates were measured during the day with the light intensity under which the cells were grown ($< 30 \mu\text{mol quanta m}^{-2} \text{sec}^{-1}$) and in the dark at night. Growth under a 12:12 light/dark cycle produced average growth rates of ~ 0.15 divisions day^{-1} for all the experiments. The chamber experiments suggest that while the sediment affects the progress of descending cells, migration occurs through thin layers of sediment and increases in response to elevated nutrient concentrations below the sediment. Since all cells found below the sediment had significantly higher C/N ratios than those remaining above the sediment, the motivation for migration appears related to a cell's internal biochemical state. The flexible exploitation of sediment-derived nutrients combines with the diel uptake and assimilation of a variety of N substrates to support low growth rates. The ability to maintain low growth rates in low light environments while accessing sediment derived nutrients permits *K. brevis* to persist as vegetative cells near the sediment-water interface. This ability to maintain slow growing populations associated with the sediment may substitute for life cycle strategies that involve encystment under unfavorable conditions that are observed in other dinoflagellates. Populations of vegetative cells associated with the sediment may provide seed populations that are advected onshore, under upwelling favorable conditions, to environments that promote cell aggregation and growth on higher light and opportunistic nutrient sources. The combination of cell aggregation and growth would result in the near-surface harmful algal blooms that are observed near-shore.

HARMFUL ALGAL BLOOMS AND THE 15°C BARRIER

Ted Smayda, Graduate School of Oceanography, University of Rhode Island, Kingston, RI 02881, USA

Experimental data on the temperature-cell division relationships of ca. 45 dinoflagellate, raphidophyte and other flagellate species representing various toxicity modes, bloom behavior and Life-form Types recognized by Smayda and Reynolds (2001, 2003) were analyzed. HAB and red tide species exhibit an unexpected, general sensitivity to temperature - an ignored factor in HAB ecology. A 15°C barrier to bloom development of the major, harmful flagellates and red tide species is evident, with two distinct thermal groups separated by this "bloom threshold temperature" recognizable. Temperatures below, or near 15°C suppress cellular growth of the raphidophytes, other "naked" flagellates, and almost all toxic dinoflagellates examined - the optimal growth temperature for these species is usually $\geq 20^\circ\text{C}$. The dinoflagellate species that are inhibited at, or below 15°C cluster into "mixing-drift" Life-form Types IV,V and VI, while those species that can grow below 15°C, and often bloom close to this temperature, belong primarily to Life-form Types I, II and III. The latter species - "cold water tolerant" - tend to be non-toxic or, if toxic, ichthyotoxic. *Alexandrium tamarense* is a conspicuous transitional species among the PST-producing *Alexandrium* spp. in bridging the 15°C barrier. The ecological consequences of the 15°C barrier and the multiple effects of temperature on motility and other behavior are considered, including the potential impacts of global warming on HABs. The relevance of the findings to Margalef's Mandala is discussed, and the hypothesis developed that it is temperature - not biophysics or ecology - which generally constrains flagellate blooms to periods when stratified waters prevail. The ecological corollary that HAB flagellates exploit, rather than require stratified waters for their growth, life cycle completion, and in meeting their nutritional (energetic) needs is considered

Smayda, T.J. and C.S. Reynolds 2001. Community assembly in marine phytoplankton: Application of recent models to harmful dinoflagellate blooms. *J. Plankton Res.* **23**: 447-461.

Smayda, T.J. and C.S. Reynolds 2003. Strategies of marine dinoflagellate survival and some rules of assembly. *J. Sea Res.* **49**: 95-106.

BLENDING OF OBSERVATIONS AND MODELS IN FORECASTING TRANSPORT OF HARMFUL BLOOMS

Richard P. Stumpf¹, Matthew Stumbaugh², Michelle C. Tomlinson¹, Timothy T. Wynne¹, Richard Patchen³, Glenn Watabayashi²

¹NOAA, National Ocean Service, Silver Spring, MD USA

²NOAA, Office of Response and Restoration, Seattle, WA USA

³NOAA, Coast Survey Development Lab, Silver Spring, MD USA

Forecast systems for harmful algal blooms (HABs) have a key objective of identifying where a HAB will have an impact. This requires coupling of HAB locations with forecasted currents in a transport model. Assimilation of HAB fields is the most problematic part of forecasted locations. Water samples are point measurements describing only a part of a HAB concentration field. Satellite imagery, when usable, is constrained by both clouds and the need for interpretation. Other data types, such as gliders and moored instruments, also are limited in coverage. As a result of these limitations, the location data will include a range of points in space and time, which points to a need for interpreted HAB fields, rather than automated assimilation. A transport model must then merge the interpreted fields and the forecast circulation.

One modeling environment, the General NOAA Operational Modeling Environment (GNOME) provides a tool and potential capability to examine this assimilation problem. GNOME was designed for horizontal two-dimensional transport, and has been configured for integration of modeled currents and fields. The transport can be examined, including consideration of the influence of characteristics and uncertainty of the HAB location field on the forecasts. Examples of transport using GNOME with fields derived from satellite and observations, and model-derived currents are made for *Microcystis aeruginosa* in Lake Erie and *Karenia brevis* in the Gulf of Mexico. The results are examined to capture the influence of uncertainties in the HAB field on the forecasts, and lead toward broadly applicable capabilities.

**RED TIDE RELATED LOSSES AND SMALL BUSINESS ADMINISTRATIONS LOANS:
A 20 YEAR RETROSPECTIVE**

Patricia A. Tester, R. Wayne Litaker, Jill Sullivan National Ocean Service, Beaufort, NC 28516, USA

An unusual sequence of events in the fall of 1987 lead to the transport of a Florida red tide (*Karenia brevis* bloom) to the coast of North Carolina (Tester et al. 1991. Limnol. Oceanogr. 36:1053-1061). Before the end of this red tide event in May 1988 more than 145,280 hectares of shell fishing waters were closed leaving 5,000 commercial fishermen were without a resource to harvest. The estimated loss of revenue to the coastal community was estimated to be \$24.7 million. In an attempt to mitigate the economic damages to the local community the Small Business Administration (SBA) was requested to lend aid. It was then North Carolina shell fishermen and other small business owners discovered that loss of revenue from red tides was not covered in the SBA’s definition of disaster. Testimony from NOAA staff and the combined efforts of the North Carolina congressmen resulted in changes in the SBA’s definition of “disaster” to include ocean conditions that resulted in the closure of customary fishing waters as stated in the Small Business Administration Reauthorization and Amendment Act of 1988. The bill amended the Small Business Act and the Small Business Investment Act of 1958. This study reports the SBA assistance for red tide related disasters under this bill. Disasters due to red tides have been declared only 7 times since the legislation was changed in 1988. Businesses in four states have been provided loans by the SBA for red tide related losses. Forty-one SBA loans were made to North Carolina businesses in 1988 for a total of \$1,334,526. In 1996, 1999, 2001 and 2002 a total of 32 loans were made to Florida businesses for a combined total of \$850,515. During the 2005 *Alexandrium* bloom from Maine through Massachusetts the fishery was declared a “failure” and shell fishermen were eligible for grants. The SBA legislation has never been invoked to assist with losses in west coast states of the US or in states bordering the Gulf of Mexico west of Florida.

Table 1. Small Business Administration loans for losses related to Red Tides or harmful algal blooms summed over all businesses from 1988 to 2005 by declaration date and by state. Loan values are given in two formats. The original loan amount is given (unshaded) and the loan amounts converted to 2006 dollars (shaded).

State	Date	Number of Loans	Max	Max	Min	Min	Total	2006 \$
			2006 \$	2006 \$	2006 \$			
NC	1988	51	\$114,500	\$189,394	\$1,200	\$1,985	\$806,800	\$1,334,526
FL	1988	4	\$28,700	\$35,177	\$4,900	\$6,006	\$61,700	\$75,625
FL	1996	16	\$73,500	\$81,912	\$4,900	\$5,461	\$390,700	\$435,414
FL	1999	12	\$589,000	\$64,682	\$4,400	\$4,832	\$174,500	\$191,630
FL	2001	4	\$61,300	\$66,885	\$5,000	\$5,456	\$135,500	\$147,845
MA	2002	3	\$46,500	\$46,500	\$5,000	\$5,000	\$67,200	\$67,200
ME	2005	6	\$26,000	\$26,000	\$2,500	\$2,500	\$67,600	\$67,600
Totals		96					\$1,704,000	\$2,319,840

GENE EXPRESSION PROFILES OF *Karenia brevis* DURING LYSIS BY ALGICIDAL BACTERIA

Michael J. Twiner¹, Patricia B. Roth¹, Kristy Lidie¹, James Ryan¹, Fran Van Dolah¹, and Gregory J. Doucette¹

Marine Biotoxins Program, NOAA/National Ocean Service, Charleston, SC 29412, USA

Flavobacterium sp. (strain S03) and *Cytophaga* sp. (strain 41-DBG2) are algicidal bacteria active against the brevetoxin (PbTx)-producing dinoflagellate, *Karenia brevis*. Both algicidal bacteria cause lysis of *K. brevis* (Fig. 1), but *Flavobacterium* sp. requires physical contact with the target algal cells (i.e., direct attack), whereas *Cytophaga* sp. releases a dissolved algicidal agent (i.e., indirect attack). However, little else is known about the specific mechanisms by which these algicidal bacteria target and lyse *K. brevis*. Time-course experiments involving exposure of this dinoflagellate to algicidal strains S03 and 41-DBG2 were conducted, utilizing an 11000 feature *K. brevis* microarray (Lidie et al., 2005) to ascertain transcriptional profiles prior to algal cell lysis. Algal RNA was collected at 12 and 36 h following the inoculation of individual algicidal bacteria into exponentially growing cultures of bacteria-free *K. brevis* (C2 isolate). RNA was amplified and fluorescently labeled prior to hybridization with time-matched control RNA collected from *K. brevis* cultures exposed to the non-algicidal bacterium *Stanierella latercula*. Relative to control cultures, algicidal strain S03 induced 625 and 366 differentially expressed *K. brevis* genes at 12 and 36 h, respectively, following inoculation. Similarly, strain 41-DBG2 induced 736 and 675 genes at 12 and 36 h, respectively. In each algicidal treatment, the majority of differentially expressed *K. brevis* genes were up regulated. Genes with known annotations (~35%) were compared based on functional groups between exposures to the two algicidal bacteria. A consistent transcriptional response induced by both algicidal strains was the up regulation of chloroplastic and photosynthetic genes (i.e., light harvesting proteins, flavoproteins) prior to cell lysis. In addition, other genes involved in translation, protein recycling, and ion signaling were up regulated by both algicidal treatments. In contrast, strains 41-DBG2 and S03 also elicited a response from functionally distinct gene sets. The indirect attacking *Cytophaga* sp. (strain 41-DBG2) induced various stress response and signaling genes consistent with an antimicrobial response (i.e., permease, neomycin fusion protein, integrin complex binding). Actin-related genes involved in motility (i.e., various flagellar proteins) and dormancy or encystment were also identified. The direct attacking *Flavobacterium* sp. (strain S03) induced more structurally-related (i.e., ankyrin, myosin heavy chain) and cell adhesion/interaction genes. Our findings suggest that algicidal bacteria induce an early transcriptional response consistent with algal cell lysis and apparently unique to the mechanism of attack. These data will be useful for assessing natural causes of *K. brevis* bloom termination and for evaluating HAB control/mitigation strategies leading to lysis of *K. brevis* cells.

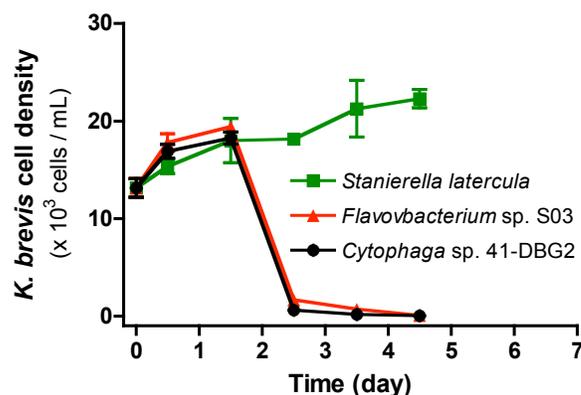


Figure 1. Growth of *K. brevis* in controls and algicidal bacteria treatments. Algal cell counts in bacteria-free cultures of *K. brevis* (C2) inoculated with the non-algicidal bacterium *Stanierella latercula* (100 cells/mL), algicidal strain S03 (10 cells/mL), or algicidal strain 41-DBG2 (100 cells/mL). Values are mean \pm SE (n=3).

Lidie, K.L., J.C. Ryan, M. Barbier and F.M. Van Dolah. 2005. Gene expression in the Florida red tide dinoflagellate *Karenia brevis*: analysis of an expressed sequence tag (EST) library and development of a DNA microarray. *Mar. Biotechnol.* **7**:481-493.

CIGUATOXICITY IN THE NORTHERN GULF OF MEXICO

T. A. Villareal¹, R. W. Dickey², G. Luber³

¹Marine Science Institute, The University of Texas at Austin, Port Aransas, Texas 78373, USA. ²FDA, CFSAN, Office of Food Safety Gulf Coast Seafood Laboratory, Dauphin Island, AL 36528. ³National Center for Environmental Health, Centers for Disease Control and Prevention, Atlanta, GA 30341

The northern Gulf of Mexico has historically been virtually free from ciguatera. Although included within maps for regions that are potentially ciguatoxic, there is little historical data to suggest a problem. Ciguatoxic barracuda were caught from the continental shelf of Texas in 1988 and 1998, but there is no public perception of ciguatera and no public health warnings have been issued. No other fish has been linked to a confirmed case of ciguatera, although there is a 20+ year old report of neurological symptoms from a grey snapper caught on the coral reefs at the Flower Garden Banks National Marine Sanctuary. A follow up study noted the presence of *Gambierdiscus* on oil platforms and *Sargassum* along the Texas coast as well as low levels of toxin in a small percentage of barracuda (Villareal et al. 2007).

In 2006-2007, an unusual number of ciguatera cases were noted. At the time of this writing, there are 9+ cases linked to both grouper and barracuda landed in Florida and Texas that are traceable to fish caught either at the Flower Gardens or along the Florida/Alabama section of the northeastern Gulf. Where available, fish has been sampled and the presence of ciguatoxin confirmed by LC-MS. A follow-up survey of 31 fish collected at the Flower Gardens found toxin levels greater than 0.2 ppb in 2 of 31 fish with trace levels reported in two more. *Gambierdiscus* has been cultured from the Flower Gardens as well, extending the range of this genus to the most northerly coral reef in U.S. waters.

These cases may represent the beginnings of a broad incursion of ciguatera into the northern Gulf of Mexico. The sudden increase in ciguatera in general and presence of toxin in a previously safe guild of fish (grouper) suggests significant changes are occurring in the Gulf of Mexico. In the Gulf of Mexico, barracuda migrate extensively following isotherms; however, grouper are generally more territorial except when assembling for spawning aggregations. The hypothesis that an endemic ciguatera food-web is present, as opposed to a migratory introduction, cannot be rejected and is consistent with predictions that the increase in artificial reefs and oil platforms, as well as warmer sea-surface temperatures could lead to increased ciguatera (Villareal et al. 2007).

Villareal, T.A., S. Hanson, S. Qualia, and R.L. Dickey. 2007. Petroleum production platforms as sites for the expansion of ciguatera in the northwestern Gulf of Mexico. *Harmful Algae* 6:253-259

MOLECULAR CHARACTERIZATION OF TOXIC CYANOBACTERIAL COMMUNITIES IN THE LOWER GREAT LAKES: A SEVEN YEAR SYNOPSIS.

Steven W. Wilhelm¹ and Gregory L Boyer²

1. Department of Microbiology, The University of Tennessee, Knoxville TN 37996 USA
2. Department of Chemistry, State University of New York - Environmental Science and Forestry, Syracuse NY 13210 USA

Lab experiments as well as field surveys and experiments have been ongoing in the lower Great Lakes (Lakes Erie and Ontario) since 1999 to gain some insight into the processes associated with the proliferation of toxic cyanobacteria. Cyanobacteria of the genus *Microcystis* have reoccurred at bloom levels in most years since 1995 with cell densities exceeding 1 million cells per liter and toxin concentrations above suggested World Health Organization standards. As better management of these events can only occur with a better understanding of their ecology, molecular biological tools, designed to phylogenetically characterize and quantify all cyanobacteria, all *Microcystis* or potentially toxigenic *Microcystis* have been developed and employed in combination with analytical chemistry and standard limnology to ascertain why blooms may occur and how environmental parameters may influence toxin production. Phylogenetic analyses of toxin genes indicate that *Microcystis* spp. are the primary producers of the hepatotoxin in microcystin in this region, although other toxin producing cells (*e.g.*, *Planktothrix*) proliferate in some regional embayments, and genetic indicators of an as-of-yet unknown to science population of cells are also present. Quantitative analyses further suggest that only a subset of the total *Microcystis* community (mean ~ 10 %, range ~ 5 - 50% across all years and stations) is potentially toxic, implying that these populations are diverse and that toxin production may carry some costs (and potential benefits) that affect competition and proliferation within the microcystis-producing community. When combined with data on environmental physiochemistry, our observations suggest a disconnect between the causes of blooms and toxin production in natural systems. These observations will be presented in the context of future management and research directions that are needed for this and other HAB-impacted aquatic systems.

ABSTRACTS OF POSTERS

APPLICATIONS OF THE OCEAN BIOGEOGRAPHIC INFORMATION SYSTEM (OBIS) TO HAB DATA

James W. Ammerman¹, Edward Vanden Berghe¹, Y. Zhang¹, and J. Fred Grassle¹
Institute of Marine and Coastal Sciences, Rutgers University, New Brunswick, NJ 08901-8521, USA

The Ocean Biogeographic Information System (OBIS) is the information component of the Census of Marine Life (CoML), an international discovery program representing a network of more than 1700 researchers in 73 nations engaged in a 10-year initiative to assess and explain the diversity, distribution, and abundance of life in the oceans - past, present, and future. OBIS is the most authoritative web-based provider of global geo-referenced information on marine species and currently includes 13.2 million records of 82,000 species from 216 databases. In addition to gathering and maintaining marine species-level and habitat-level databases, it provides a variety of spatial query tools for visualizing geographical relationships among species, and between species and their environment. OBIS is growing rapidly to become the national, regional, and international infrastructure for information on marine species and their distribution and abundance, and playing a central research role in ocean biodiversity informatics (Costello and Vanden Berghe 2006).

The current number of HAB-related databases around the world demonstrates both the interest in and the fragmentation of HAB data. There is currently a concerted effort at the international level to consolidate HAB databases. Because it is a worldwide distributed database, OBIS should be an excellent vehicle for this consolidation, though it has not been specifically used for HABs in the past.

OBIS is looking for opportunities to work with HAB scientists and other interested groups to incorporate HAB data sets into OBIS and develop targeted products for end users. We hope to increase the incentives for investigators to supply their research and monitoring data to databases like OBIS, by facilitating data entry and then making this data available in more useful formats that can be compared with others. This should help to improve linkages of HAB data to related environmental data as well as improve taxonomic identification and standardization. A new HAB portal will be created on the OBIS Home Page including useful links to other HAB resources.

On the technical level, the schema specifying the information shared within OBIS could be extended with elements specific for HABs. These extensions could then be incorporated in the database structure and portal site of OBIS. On the community level, the first step is an analysis of needs and expectations of the HAB scientists; results will feed into the technical part and guide development. The new technological platform should provide an incentive for HAB scientists to share their data in the OBIS community; this would bring them several advantages: first of all, their data would become available in a standardized format, together with data from other providers - both within the U.S. and outside. Finally, analytical tools developed by OBIS will automatically be available, such as tools to correlate physical oceanographic measurements with species distributions.

Costello, M. J., and E. Vanden Berghe. 2006. 'Ocean biodiversity informatics': a new era in marine biology research and management. *Mar. Ecol.-Prog. Ser.* **316**: 203-214.

A FUZZY LOGIC APPROACH TO PREDICTING *Prorocentrum minimum* BLOOMS IN THE CHESAPEAKE BAYJon T. Anderson¹¹Estuarine Research Center, Morgan State University, Saint Leonard, Maryland 20686, USA

The dinoflagellate, *Prorocentrum minimum*, forms seasonal blooms in the mesohaline portions of the Chesapeake Bay and its tributaries. Often, these blooms cause hypoxic conditions, which can result in fish kills. Furthermore, certain strains can produce toxins that directly affect the survivorship or behavior of finfish and bivalves. Understanding the environmental conditions preceding a bloom can help scientists understand the mechanisms behind bloom formation. Fuzzy logic provides a novel, flexible solution for predicting blooms, since it incorporates the uncertainty of categorizing and defining a bloom concentration. The Chesapeake Bay and its tributaries are an ideal location for developing predictive models because of the multi-decade, spatially extensive water quality monitoring and phytoplankton datasets available. The results of predictive models at three representative water quality monitoring stations in the Chesapeake Bay will be presented, along with an examination of the sensitivity of the models to the definition of a “bloom” concentration. The usefulness of the model in illustrating potential mechanisms of bloom formation, as well as in guiding efficient best management practices (BMPs), will also be explored.

GROWTH, TOXICITY AND NITROGEN UPTAKE CAPABILITIES OF THE TOXIGENIC DIATOM *Pseudo-nitzschia cuspidata* FROM THE PACIFIC NORTHWESTMaureen E. Auro¹, William P. Cochlan¹, and Vera L. Trainer²¹Romberg Tiburon Center for Environmental Studies, San Francisco State University, Tiburon, CA, 94920-1205²NOAA Northwest Fisheries Science Center, Seattle, WA, 98112

The toxigenic diatom, *Pseudo-nitzschia cuspidata* (Hasle) has recently been found in bloom concentrations in the Juan de Fuca Eddy region off the coasts of Washington State, U.S.A. and British Columbia, Canada. Isolates collected during the September surveys of the ECOHAB-PNW project during 2004 and 2005 were examined for their growth, toxicity and nitrogen uptake response to inorganic (nitrate, ammonium) and organic (urea) nitrogen. The kinetics of N uptake as a function of substrate concentration were estimated from short (20-min) incubations using the ¹⁵N-tracer technique, and are compared to the long-term exponential growth rates of *P. cuspidata* determined in semi-continuous, batch cultures grown on the various nitrogen substrates. Based on the estimated maximum specific uptakes rates (V_{max}), nitrogen preference follows the order: ammonium > nitrate > urea, whereas the nutrient affinity indices - half saturation constants (K_s) and alpha parameter ($\alpha = V_{max}/K_s$), indicate that N affinity follows the order: urea > nitrate > ammonium. Long-term growth experiments conducted at saturating ($120 \mu E \cdot m^{-2} \cdot s^{-1}$) and sub-saturating ($40 \mu E \cdot m^{-2} \cdot s^{-1}$) photosynthetic photon flux densities (PPFDs), demonstrate that *P. cuspidata* grows significantly faster at the higher PPFD for all N substrates, but there are substantial differences in the growth rates achieved on the various N substrates. The exponential growth rate (determined using raw fluorescence units) at high PPFD is slower for cells grown on urea ($0.84 \pm 0.03 d^{-1}$) compared to the cells maintained on nitrate ($0.88 \pm 0.01 d^{-1}$) or ammonium ($0.91 \pm 0.02 d^{-1}$), whereas at low PPFD, urea supported faster growth ($0.65 \pm 0.003 d^{-1}$) than either nitrate ($0.55 \pm 0.01 d^{-1}$) or ammonium ($0.51 \pm 0.02 d^{-1}$).

Recently, Armstrong-Howard *et al.* (2007) showed that both field assemblages and laboratory cultures of *P. australis* produce more domoic acid (both cellular and dissolved DA) when grown on urea versus nitrate or ammonium. In our study, the cellular DA content (determined using cELISA) for *P. cuspidata* did not significantly differ as a function of the nitrogen substrate used for growth at saturating PPFD, but at sub-saturating PPFD, nitrate-grown cells produced 74% and 78% more pDA per cell than ammonium- and urea-grown cells. In contrast to other *Pseudo-nitzschia* species where cellular domoic acid is generally enhanced during stationary phase, the cellular DA concentrations for *P. cuspidata* were always greater during exponential growth compared to stationary growth, regardless of the N substrate or PPFD used for growth. These laboratory results demonstrate the capability of this small, toxigenic diatom to grow and produce DA on both oxidized and reduced N substrates supporting field observations that *Pseudo-nitzschia* species bloom during both upwelling and non-upwelling conditions off the west coast of North America where substantial differences in the nitrogenous nutrition of *P. cuspidata* can be expected.

ARMSTRONG-HOWARD, M. D., COCHLAN, W. P., LADIZINSKY, N. L. AND KUDELA, R. M. 2007. Nitrogenous preference of toxigenic *Pseudo-nitzschia australis* (Bacillariophyceae) from field and laboratory experiments. *Harmful Algae* 6: 206-217.

LOW SINKING RATES OF *Pseudo-nitzschia*: A COMPETITIVE FEATURE CONTRIBUTING TO THE DEVELOPMENT AND MAINTENANCE OF TOXIC BLOOMS

Benjamin Beall¹, and Charles G. Trick^{1,2}, William P. Cochlan³, Vera Trainer⁴, and Mark L. Wells⁵.

¹Department of Biology, University of Western Ontario, London, ONT, N6A 5B7

²Schulich School of Medicine and Dentistry, University of Western Ontario, London, ONT, N6A 5B7, Canada

³Romberg Tiburon Center for Environmental Studies, San Francisco State University, Tiburon, 94920, USA

⁴NOAA Northwest Fisheries Science Center, Seattle, 98112 USA,

⁵School of Marine Sciences, University of Maine, Orono, ME 04469, USA,

The toxigenic diatom genus *Pseudo-nitzschia*, responsible for amnesic shellfish poisoning, is a significant harmful algal bloom taxon. Efforts to predict the blooms and subsequent fate of *Pseudo-nitzschia* cells require understanding the factors that contribute to the growth and mortality of the diatom. Balancing cell growth and nutrient uptake rates against grazing-determined loss rates is the foundation for many models of bloom maintenance. However, sedimentation is a major loss mechanism for diatoms often not considered in describing bloom development. In addition to their unique cell morphology, *Pseudo-nitzschia* spp. often display behaviours (e.g. optional chain formation) that have the potential to minimize cell loss through sedimentation, and thereby extending the duration and magnitude of toxigenic diatom bloom events. The relative sinking rates of natural communities containing *Pseudo-nitzschia* were studied along the coasts of Washington State, U.S.A., and British Columbia, Canada, as part of the ECOHAB-PNW project. Sinking rates of *Pseudo-nitzschia* were substantially lower than those of the entire phytoplankton community during the early development and midpoint of toxic bloom formation. Measurable sinking rates of *Pseudo-nitzschia* were observed only late in the bloom. Even so, the maximum observed sinking rate of *Pseudo-nitzschia* was less than measured for the phytoplankton community as a whole. This change in *Pseudo-nitzschia* sinking rates corresponded with a sharp reversal of intracellular vs. extracellular domoic acid concentrations, where high concentrations of dissolved DA were released by the cells. The increase in *Pseudo-nitzschia* sinking rates also was accompanied by a higher potential formation of aggregates, measured in a Couette device, that was likely mediated by increased concentrations of exocellular polysaccharides. These results suggest that the low observed sinking rate of *Pseudo-nitzschia* during bloom development could contribute to the competitive success of this taxon, and sedimentation may only be significant after periods of substantial aggregation and subsequent sinking of the diatom flocs.

PHYLOSORT: A PROGRAM FOR DETECTING ENDOSYMBIOTIC AND HORIZONTAL GENE TRANSFER AND ITS APPLICATION TO UNDERSTANDING STX GENE ORIGIN IN *Alexandrium tamarense*

Debashish Bhattacharya¹, and Ahmed Moustafa¹

¹University of Iowa, Iowa City, IA 52242, USA

Genome data offer the potential to clarify long-standing issues in organismal biology. The size and complexity of genome data provides however a significant hurdle in realizing this goal. To aid the identification of genes that have originated through endosymbiotic (i.e., intracellular; e.g., Reyes-Prieto et al. 2006) or horizontal (i.e., foreign source) gene transfer (EGT and HGT, respectively), we have developed a computer program, PhyloSort for automated analysis of trees generated by phylogenomics. Phylogenomic methods allow the user to simultaneously compare dozens of genomes to each other and to generate alignments and phylogenies of each gene homolog. This approach has revolutionized genome annotation and gene family analysis and provided large data sets for multi-gene phylogenetics. Existing phylogenomic pipelines such as PhyloGenie (Frickey and Lupas 2004) and PhyloGena (Hanekamp et al. 2007) generate a large collection, often hundreds or thousands, of phylogenetic trees that the user has to manually inspect to observe general patterns of genome evolution or to address specific hypotheses about gene phylogeny. PhyloSort automates tree topology analysis in a high-throughput fashion. This open-source Java tool allows the user to query the topology of phylogenetic trees to address the most frequently asked question in the field, does a specific gene support the monophyly of specified OTUs? PhyloSort reads Phylip formatted trees as input and can be used via a graphical user interface (GUI) and a text mode command line interface. Initially, the user specifies a source folder containing the input phylogenetic trees and a target folder to where trees that satisfy the search criteria can be copied or moved or alternatively, simply counted. Thereafter sets of taxa (query) that are postulated to be monophyletic are specified and searched in the input trees. The search criteria can be adjusted by setting a minimum bootstrap support value associated with the monophyletic clades and a minimum and/or maximum number of taxa in the trees with matching clades. The trees can also be “clustered” to generate a “uni-tree” set that encompasses gene families. Here we describe PhyloSort using examples of analysis of EGT in different algae. Thereafter, we apply the program to identify homologous genes shared exclusively between the saxitoxin (STX) producing taxa, the dinoflagellate *Alexandrium tamarense* and the cyanobacterium *Anabaena circinalis*. Other prokaryotes and eukaryotes will also be included in this analysis. The goal here is to identify potential HGT events that may have resulted in the origin of STX genes in *Alexandrium* from the cyanobacterium (or some other source). This hypothesis of STX gene origin in *Alexandrium* through HGT is supported by extensive preliminary data that demonstrates that dinoflagellates are masters of harvesting genes from foreign sources.

FRICKEY, T., and A. N. LUPAS. 2004. PhyloGenie: automated phylome generation and analysis. *Nucleic Acids Research* **32**: 5231-5238.

KRISTIAN, H., U. BOHNEBECK, B. BESZTERI, and K. VALENTIN. 2007. PhyloGena -- a user-friendly system for automated phylogenetic annotation of unknown sequences. *Bioinformatics* **23**: 793-801.

REYES-PRieto, A., J. D. HACKETT, M. F. BONALDO, M. B. SOARES, and D. BHATTACHARYA. 2006. Cyanobacterial contribution to algal nuclear genomes is primarily limited to plastid functions. *Current Biology* **16**: 2320-2325.

PUGET SOUND, WASHINGTON: AN EMERGING HOTSPOT FOR *Pseudo-nitzschia* BLOOMS AND DOMOIC ACID TOXIC EVENTS

Brian D. Bill^{1,2}, William P. Cochlan¹, Vera L. Trainer², Mark L. Wells³, Charles G. Trick⁴, and Barbara M. Hickey⁵

¹Romberg Tiburon Center for Environmental Studies, San Francisco State University, Tiburon, CA, 94920-1205

²NOAA, Northwest Fisheries Science Center, Seattle, WA, 98112

³School of Marine Sciences, University of Maine, Orono, ME, 04469-5741

⁴Schulich School of Medicine and Dentistry, University of Western Ontario, London, ONT, N6A 5B7, Canada

⁵School of Oceanography, University of Washington, Seattle, WA, 98195

Pacific Northwest inland embayments with limited direct exchange with oceanic waters contain a number of HAB species, including the Paralytic Shellfish Poisoning (PSP) dinoflagellate genus *Alexandrium*, the fish-killing raphidophyte *Heterosigma akashiwo*, and the Amnesic Shellfish Poisoning (ASP) diatom genus *Pseudo-nitzschia*. While blooms of the toxigenic diatom, *Pseudo-nitzschia* dominate public and scientific attention in the well-studied coastal systems of western North America (e.g., Monterey Bay, Santa Barbara and San Pedro Channels, Juan de Fuca Eddy), the recent *Pseudo-nitzschia* blooms observed within the inland waters of Puget Sound, Washington suggest that toxic diatom events are not isolated to these coastal systems. *Pseudo-nitzschia* have been observed in Puget Sound waters since the 1930's (then identified as belonging to the *Nitzschia* genus), definitively identified as *Pseudo-nitzschia* species since 1990 (Horner and Postel, 1993; Trainer *et al.*, 1998), and although the neurotoxin domoic acid (DA) has been detected since 1991, only recently have levels exceeded regulatory limits and led to shellfish harvest closures in 2003 and 2005 (Trainer *et al.*, 2007). Our monitoring efforts within Puget Sound have revealed the ubiquitous presence of *Pseudo-nitzschia* species, routine detection of DA, as well as adequate macronutrient concentrations and composition to sustain algal blooms, both benign and harmful. Research cruises conducted in June and September 2006 found *Pseudo-nitzschia* cell concentrations as high as 3.4×10^5 cells L⁻¹, particulate DA concentrations as high as 105.2 pg ml⁻¹, and surface water concentrations of nitrate, ammonium, and silicic acid were routinely 18-20, 1.4-2.0, and 50 μM, respectively, well above macronutrient concentrations limiting for growth. Following the first closures of shellfish harvesting due to DA in 2003 and 2005, it appears that Puget Sound is becoming increasingly conducive to toxic DA events. Due at least partially to increasing human population densities and their use of the marine resources within Puget Sound (including aquaculture activities), the increased frequency of toxic DA events raises questions regarding their impacts on ecosystem and human health. Future sustained monitoring efforts, and research designed to characterize local nutrient dynamics (both natural and anthropogenic) and phytoplankton ecology, are necessary to ensure that commercial, recreational, and Tribal subsistence fisheries continue to provide safe products for human consumption.

HORNER R.A. AND POSTEL J.R. 1993. Toxic diatoms in western Washington waters (U.S. west coast). *Hydrobiologia* 269/270: 197-205.

TRAINER V.L., ADAMS N.G., BILL B.D., ANULACION B.F., AND WEKELL J.C. 1998. Concentration and dispersal of a *Pseudo-nitzschia* bloom in Penn Cove, Washington, USA. *Nat. Toxins* 6: 113-126.

TRAINER V.L., COCHLAN W.P., ERICKSON A., BILL B.D., COX F.H., BORCHERT J.A., AND LEFEBVRE K.A. 2007. Recent domoic acid closures of shellfish harvest areas in Washington State inland waterways. *Harmful Algae* 6: 449-459.

LONG-TERM (1992-2006) PATTERNS IN *Phaeocystis* BLOOM MAGNITUDE AND BLOOM DURATION IN MASSACHUSETTS BAY, USADavid G. Borkman¹, P. Scott Libby² and Jefferson T. Turner³¹ Graduate School of Oceanography, University of Rhode Island, Narragansett, RI 02882, USA. dborkman@gso.uri.edu² Battelle Ocean Sciences, Brunswick, ME, USA. libby@battelle.org³ University of Massachusetts Dartmouth, North Dartmouth, MA 02747, USA. jturner@umassd.edu

Blooms of the colonial prymnesiophyte *Phaeocystis pouchetii* are a regular component of the winter-spring phytoplankton of north-temperate coastal seas, including the Gulf of Maine. These high biomass *Phaeocystis* blooms may sequester large portions of ecosystem resources and can alter trophic pathways; making *Phaeocystis* a 'keystone' phytoplankton taxa that may significantly alter ecosystem function. *Phaeocystis pouchetii* abundance was quantified at two-week to monthly intervals (12 to 17 times per year) in Massachusetts Bay (southern Gulf of Maine, NW Atlantic) for fifteen years (1992-2006) as part of a long-term phytoplankton monitoring program. Related hydrographic, nutrient, and zooplankton data were also collected. Time series and statistical analyses were used to identify patterns and infer mechanisms of *Phaeocystis* bloom initiation, maintenance and termination during 1992-2006.

Time series analyses indicated an increasing *Phaeocystis* abundance trend during 1992-2006, driven by the large blooms in 1997, 2000, and during 2004. Coincident with this *Phaeocystis* pattern, diatom abundance had a long-term decline during 1992-2006, with 2005 - 2006 mean levels (ca. 150,000 cells per liter) that were ca. 27% of the peak level (ca. 550,000 per liter) observed during 1994. Within this long-term decline there were relative peaks in diatom abundance in 1994, 1998 and 2002. The relative diatom abundance peaks approximately corresponded with relative nadirs in *Phaeocystis* abundance and correlation analysis of these two trends ($r = -0.54$, $p < 0.0001$) indicated a long-term negative interaction between *Phaeocystis* and winter-spring diatom abundance.

Phaeocystis blooms of greater than one million cells l^{-1} were observed in seven of the 15 years, while greater than 10 million cells l^{-1} were observed during 1997, 2000, and 2004. *Phaeocystis* was present from February through early June, with the annual bloom maximum occurring in April. The maximum *Phaeocystis* abundance of 15.5 million cells l^{-1} was observed during April 2004. The time of *Phaeocystis* bloom initiation was relatively consistent, however, within the limits of detection imposed by the monitoring schedule, bloom duration appears to have increased significantly (linear regression, $r^2 = 0.41$, $p = 0.0237$) at a linear rate of ca. six days per year during 1992-2006. This increased bloom duration was due to an extension of bloom termination which extended *Phaeocystis* presence into May or early June during 2002-2006. Consistent with its physiological threshold, *P. pouchetii* was not present at water temperature greater than 14°C, and the date of attainment of the 14°C water temperature threshold varied by 39 days, occurring as early as early May (2001) to as late as mid June (1993). Annual variation in winter-spring water temperature explained much (ca. 50%) of the annual variation in bloom duration, with blooms continuing later and subsequently having longer duration in years having cool spring water temperature. Impacts of *Phaeocystis* blooms on zooplankton abundance were also evaluated. A pattern of elevated *Calanus* abundance early in the season (Feb-Mar) and reduced *Oithona* and total zooplankton abundance later in the season (April-May), followed by increased *Oithona* abundance during summer was associated with *Phaeocystis* bloom years.

EVALUATION OF SHORT AND LONG LASTING NEUROLOGICAL RESPONSE TO SINGLE AND REPEATED CIGUATOXIN EXPOSURE IN MICE

Marie-Yasmine Bottein¹, Amir H. Rezvani², Christopher J. Gordon³, Edward D. Levin², and John S. Ramsdell¹

¹Marine Biotoxins Program, NOAA-National Ocean Service, Charleston, SC 29412, ² Dept. of Psychiatry, Duke University Medical Center, Durham, NC 27710, ³ US-EPA Research triangle park, NC 27711

Ciguatera fish poisoning is a common illness in tropical and subtropical regions that manifests complex and long-lived neurological symptoms, often more severe in recurrent cases. To better evaluate the hallmark chronic and repeat ciguatera symptoms we measured based upon batteries of tests the neurological signs and blood toxin levels in mice following one or two administrations of ciguatoxin (P-CTX-1, 0.264 ng/g). Telemetric recording of body temperature showed an early hypothermic response (nadir value at 1hr of 32.2°C) followed by a long lasting (at least 5 days) thermoregulatory dysfunction displayed as a stabilized body temperature (at 36°C) with no observable circadian rhythm. The thermoregulatory response was greater following the second exposure, with a significantly sustained (at least 3 days) and stronger (nadir value of 30.13 at 4.5hr) response. Peripheral effect measurement through response latency to radial heat stimulus in the tail flick assay revealed a higher pain threshold in exposed vs. control mice. This long lasting dysfunction of pain perception continued after the recovery from hypothermia. These findings support an acute phase of ciguatoxin toxicity, of short duration, associated as central thermoregulation to lower body temperature and reduced motor activity, and a sensory neuropathy of long duration, featured by persistent reduction in pain threshold and alteration of thermoregulatory process. Biomonitoring of blood toxin levels confirmed ciguatoxin (1.4 pg/ml, limit of detection) for 3 days after exposure, and the concentration measured was significantly higher at 1 hour but not at 4, 24 and 72hr following second exposure. This indicates that repeat exposures lead to larger acute blood levels of ciguatoxin, and these acute internal levels are sufficient for the chronic progression of ciguatera symptoms.

USING REAL-TIME PCR TO DEMONSTRATE CO-OCCURRENCE OF *Karlodinium veneficum* AND THE PARASITIC DINOFLAGELLATE *Amoebophrya* SP. EX. *Karlodinium veneficum*

Holly A. Bowers¹, D. Wayne Coats², Tsvetan R. Bachvaroff², Jason E. Adolf¹, and Allen R. Place¹

¹Center of Marine Biotechnology, Baltimore, MD 21202, USA

²Smithsonian Environmental Research Center, Edgewater, MD 21037, USA

Karlodinium veneficum is an ichthyotoxic bloom forming dinoflagellate common to the Chesapeake Bay and other estuarine systems throughout the world. Blooms of *K. veneficum* are thought to be controlled, in part, by the parasitic dinoflagellate *Amoebophrya* sp. ex. *K. veneficum*. This parasite is also common in estuarine environments where it uses its host for carrying out a simple life cycle, including a free-swimming dinospore that infects the host, a vegetative trophont that grows within the host, and a vermiform stage which breaks down into dinospores that get released upon host death. This parasitic infection disrupts reproduction, resulting in a decline of the host population.

The infection of *Karlodinium veneficum* by *Amoebophrya* sp. ex. *K. veneficum* can easily be observed in controlled laboratory experiments using light microscopy. Making these observations in environmental samples becomes difficult when other organisms of similar size are present. Therefore, we designed two real-time PCR assays, based on Taqman methodology, to target the internal transcribed region (*K. veneficum*) and the 18S locus (*Amoebophrya* ex. *K. veneficum*). Annealing temperature, sensitivity and reaction efficiencies were similar for each assay, allowing them to be used effectively for quantifying the co-occurrence of these two organisms throughout controlled infection experiments and over the course of several bloom events. In controlled infection experiments, there is a clear decrease in *K. veneficum* ITS signal six hours post-infection, while in natural samples real-time PCR data demonstrate a definite co-occurrence of these species.

EVIDENCE OF A SELF-RECOGNITION SYSTEM IN THE SEXUAL LIFE-CYCLE OF *Alexandrium tamarense*

Michael L. Brosnahan¹, Robert J. Olson¹, Deana L. Erdner², and Donald M. Anderson¹

¹Woods Hole Oceanographic Institution, Woods Hole, MA 02540, USA

²University of Texas at Austin Marine Science Institute, Port Aransas, TX 78373, USA

Sexual crosses of Group 1 clones (formerly termed North American ribotype) and Group III clones (formerly termed Western European ribotype) are used to investigate patterns in gamete selection in the dinoflagellate *Alexandrium tamarense*. Cysts resulting from the crosses are analyzed using a nested PCR assay for detection of Group I and Group III ribosomal sequences and also by imaging flow cytometry to assess DNA content. Results are compared to those from natural cysts collected from the Gulf of Maine. These data promise to provide new insights into the cellular mechanism of sexual induction and also patterns of genomic control during the sexual phases of the cells' life cycle. We seek to test several hypotheses derived from a rudimentary model of the dinoflagellate sexual cycle and models derived from other protists. Results thus far suggest a self-recognition system operates as a gate to complete gametic differentiation of these cells in culture. If verified, this finding has significant implications for the interpretation of mating compatibilities and to laboratory study of dinoflagellate sexual processes.

THE NUTRITIONAL ECOLOGY OF THE HARMFUL DINOFLAGELLATE BLOOMS CAUSED BY *Cochlodinium polykrikoides* IN LONG ISLAND BAYS (NY, USA)

Amanda Burson, Ying Zhong Tang, Christopher J. Gobler

Marine Sciences Research Center, Stony Brook University, 239 Montauk HWY, Southampton, NY 11968, USA

Harmful algal blooms caused by *Cochlodinium* species have been reported worldwide, including both the east and west coast of North America. *Cochlodinium polykrikoides* formed dense blooms in the Peconic Estuary and Shinnecock Bay in Long Island, NY, USA during the late summer and fall of 2004-2006. Bloom waters with cell densities higher than 5×10^4 cells ml^{-1} have been documented as being acutely toxic to multiple fin fish and shellfish species. The mechanisms for the initiation and development of these blooms are not clear. We explored the nutrient preferences and requirements and effect of salinity on the growth rate and yield for a strain of *C. polykrikoides*, CP1, which was isolated from the 2006 fall bloom in Shinnecock Bay. *C. polykrikoides* displayed a higher growth rate and cell yield in GSe medium (Doblin et al. 1999) prepared with artificial seawater compared to those obtained in GSe prepared with filtered seawater at the same salinity. *C. polykrikoides* showed a preference for different nitrogen sources (urea = ammonium > nitrate) with regard to growth rate, while the ultimate cell yield was determined by the initial concentration of nitrogen. It was also shown that *C. polykrikoides* favored the lower salinities in the range of 23-37.5 and that selenium was not required for its growth. Further study on the nutritional effects of other nutrient sources on *C. polykrikoides*, CP1, will also be presented.

The importance of nitrogen supply to the occurrence of *C. polykrikoides* blooms was further substantiated during field incubation experiments conducted during the 2006 bloom event on Long Island. Nutrient amendment, bottle incubation experiments were conducted with varying levels of nitrogen, phosphorus, and organic micronutrients (B-vitamins) using bloom water and the net growth rates of *C. polykrikoides* within each treatment were quantified. During all experiments conducted (n=5), only nitrogen enrichment was capable of significantly increasing *C. polykrikoides* growth rates relative to control treatments ($p < 0.05$; Tukey). In contrast to laboratory findings, the response of field populations to varying nitrogen sources did not differ significantly. These results suggest that nitrogen loads will determine the absolute magnitude of *C. polykrikoides* blooms, which in turn will likely influence the toxicity of a bloom event.

Doblin, M. A., Blackburn, S. I., Hallegraeff, G. M. 1999. Growth and biomass stimulation of the toxic dinoflagellate *Gymnodinium catenatum* (Graham) by dissolved organic substances. *J. Exp. Mar. Biol. & Ecol.* 236:33-47.

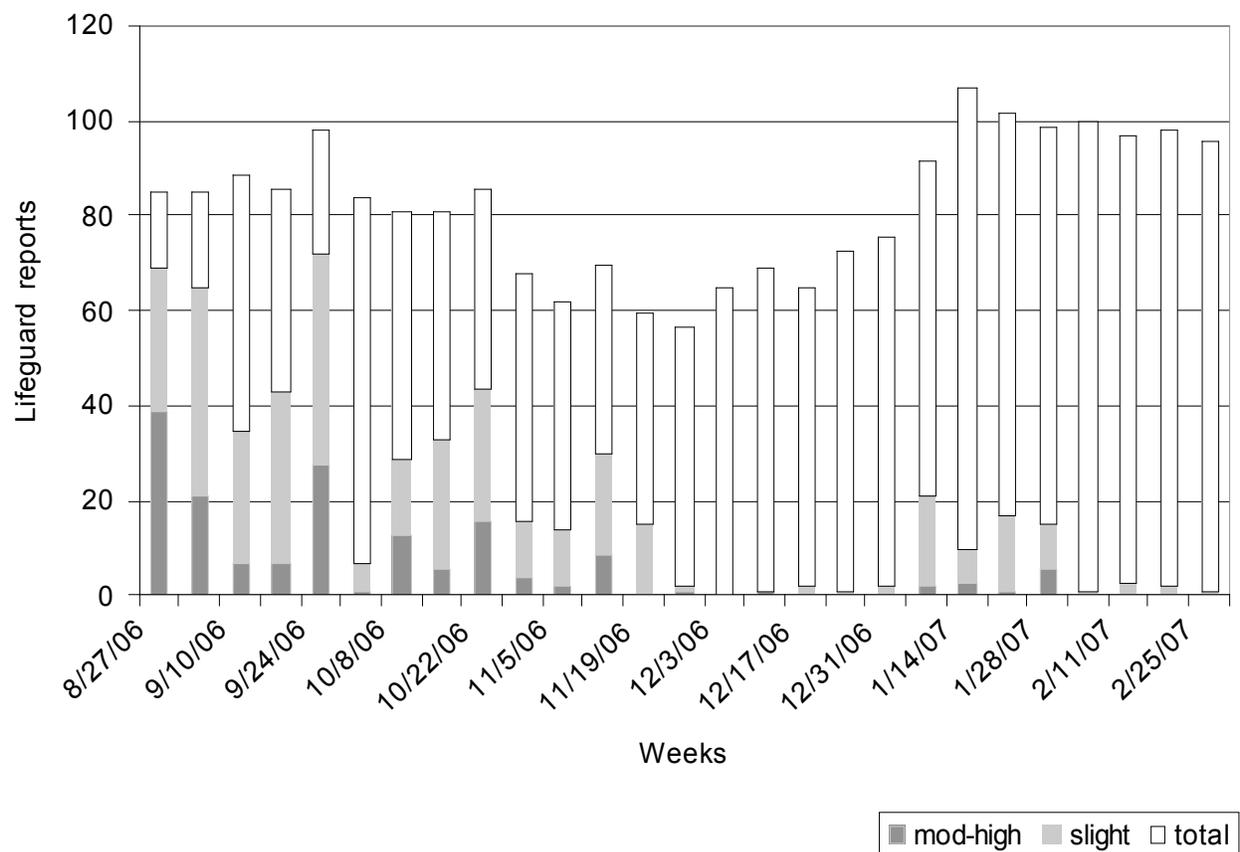
USING A NOVEL LOW-COST MONITORING ARRAY FOR VALIDATION OF BREVETOXIN-INDUCED RESPIRATORY IMPACT IN SOUTHWEST FLORIDA

Julie A. Calkins¹, Richard P. Stumpf¹, Barb Kirkpatrick², Michelle C. Tomlinson¹, Kate Nierenberg² and Bob Currier²

¹NOAA, National Ocean Service, centre for Coastal Monitoring and Assessment, Silver Spring, MD 20910, USA

²Mote Marine Laboratory, Sarasota, FL 34236, USA

Brevetoxins are a known respiratory irritant, posing a health hazard for beachgoers and local residents, particularly asthmatics. Increased incidence of respiratory irritation related to a HAB event is documented when toxic aerosol concentrations exceed 10 ng/m³ of air. This is a major concern for the state of Florida, as it has annual blooms of *K. brevis* off its gulfside beaches. Starting in August 2006, the professional lifeguard corps in Sarasota County began twice daily reports of the presence of respiratory irritation at six permanent sites. This dataset provides systematic results of impact with high temporal and spatial resolution. The benefits of this cheap, low-tech system are real-time monitoring of the health impacts during the presence of a harmful algal bloom and, in addition, validation of the HAB impact forecasts although with the risk of random error between impact classes due to subjectivity. NOAA’s HAB Bulletin forecasts the degree of local health impact during a HAB event, considering the location and extent of the HAB as well as the wind direction forecast (e.g. onshore/offshore). Previously only anecdotal or informal reports of impact were available. A comparison of the datasets has found that 67% of high-moderate respiratory impact occurrences were correctly predicted by the HAB forecast bulletin. The dataset allows exploration of incorrect predictions for causation and with the goal of increasing forecast reliability.



GENETIC VARIATION AMONG ISOLATES OF *Karenia brevis* AS MEASURED WITH MICROSATELLITE MARKERS

Lisa Campbell^{1,2}, Darren Henrichs², Mark A. Renshaw³ and John R. Gold³

¹Department of Oceanography, Texas A&M University, College Station, TX 77843, USA

²Department of Biology, Texas A&M University, College Station, TX 77843, USA

³Center for Biosystematics and Biodiversity, Texas A&M University, College Station, TX 77843, USA

Karenia brevis is the major harmful algae of concern in the Gulf of Mexico, yet the source of bloom populations, especially in the western Gulf of Mexico, is not well known. Ten microsatellite markers, nine previously described (Renshaw et al. 2006) and one newly characterized, were used to assess genetic diversity among 32 clonal cultures of *K. brevis* isolated over the past 50 years from blooms along the coasts of Florida and Texas. The number of alleles at each microsatellite ranged from three to ten and estimated gene diversity per microsatellite ranged from 0.287 to 0.851 (average 6.67 ± 0.178). Variation among the clonal cultures was extremely diverse; 21 unique haplotypes were found among 22 Florida cultures, whereas 10 unique haplotypes were found among the 10 Texas cultures. In the latter, eight of the 10 isolates were from the same bloom. Allele distributions, however, were not correlated with bloom (year) or location (Florida vs. Texas). At eight of the ten microsatellites, the most frequently occurring allele was the same for clonal cultures from both Florida and Texas. Microsatellite allele distributions among individual cells sampled during a 2005 bloom along the coast offshore of Corpus Christi, Texas, revealed that the number of alleles found among the available culture collections is less than the number of alleles found in the field sample. These results indicate estimates of genetic diversity in *K. brevis* based on culture collections may be underestimates of genetic diversity in bloom populations. Our observations of genetic diversity in *K. brevis* are similar to results found in blooms of other phytoplankton species. In *K. brevis*, blooms appear not to be clonal but rather to be extremely diverse. Results from this study emphasize the importance of examining multiple individuals of a phytoplankton species to assess genetic variation.

RENSHAW, M.A. ET AL. 2006. Microsatellite DNA markers for population genetic studies in the dinoflagellate *Karenia brevis*. *Molecular Ecology Notes* **6(4)**: 1157-1159.

COASTAL BLOOM DYNAMICS IN SOUTHERN CALIFORNIA - HAVE THERE BEEN ANY CHANGES SINCE 1917?

Melissa Carter^{1,2}, John McGowan¹, Lilian Busse^{1,3}, Elizabeth Venrick¹ and Mary Hilbern¹

¹Scripps Institution of Oceanography, La Jolla, CA, 92093, USA

³University of San Diego, San Diego CA, 92110, USA

²San Diego Regional Water Quality Control Board, San Diego, CA, 92123, USA

Algal blooms may be increasing in frequency and intensity along the coastal waters of the United States. The objective of this research is to determine if these findings are consistent with monitoring programs at Scripps Pier, La Jolla, California, and investigate questions related to bloom dynamics. We have a unique opportunity to compare over 40 years of time series data collected by E. W. Allen (1917-1939), J. McGowan et al. (1983-2000), and L. Busse et al. (2003-2005) to our current monitoring program (2005-2007).

In order to determine if the frequency of algal blooms has increased over time, we have defined a bloom as 1.5 standard deviations above the long-term mean chlorophyll concentration (14.03 mg/m³, 1983-2000). At least 50 blooms have occurred from 1983-2007, and our results indicate there is a long-term rise in the overall trend of the chlorophyll concentration. The timing of these blooms at Scripps Pier has been found to occur during the late spring and early summer periods.

We will also evaluate the species responsible for the blooms identified and determine if the species assemblage has changed over time or season. Also using the historical Allen data, we will determine if the phytoplankton community structure has changed over time.

LONG-TERM EVALUATION OF A SATELLITE OCEAN COLOR DATASET TO DETECT BLOOMS OF THE TOXIC DINOFLAGELLATE *Karenia brevis* OFF THE WEST FLORIDA SHELF

Gustavo A. Carvalho, Peter J. Minnett, Warner Baringer and Viva Banzon

University of Miami, Rosenstiel School of Marine and Atmospheric Science, Miami-FL 33149, USA

Despite the superb splendor of phytoplankton when observed microscopically, and their key importance on the marine biosphere, a minority of them possess a capability to produce toxins that are noxious to other forms of life. In the case of the toxin producing micro-algae, whenever growth rates exceed cell dispersions it causes what is known as harmful algal blooms (or red tides as they are often called). A nearly annual incidence of blooms of the toxic dinoflagellate *Karenia brevis* affects the West Florida Shelf, posing a threat to public well-being and causing massive fish and marine mammal deaths (Van Dolah, 2000). The present work aims to test the long-term stability of three approaches to remotely detect *K. brevis* blooms from satellite ocean color measurements (SeaWiFS and MODIS). Originally, these approaches were each successfully applied to single images, and are based upon the fact that there is less grazing over *K. brevis* due to the toxin it produces (i.e. brevetoxin; PbTx) such that the inherent optical properties of the water impart from other local bloom forming phytoplankton species. The first technique (Canizzaro et al., 2002) makes use of remote sensing reflectance ratios (at 443, 490 and 555nm) to detect *K. brevis*, while the second one (Canizzaro, 2004) suggests a classification scheme based on two criteria: high chlorophyll plus particulate backscatter at 555nm being lower than that calculated by Morel (1988). The performance of a newer methodology (Carvalho et al., 2007), which takes into account only a single water leaving radiance band at 555nm to flag *K. brevis* blooms, will also be tested on a longer period. The in situ dataset, containing abundance of *K. brevis* (cells per liter), was obtained from the Florida Fish and Wildlife Research Institute and will be used to corroborate the retrievals of each of the three algorithms.

CANNIZZARO, J.P. 2004. Detection and Quantification of *Karenia brevis* Blooms on the West Florida Shelf from Remotely Sensed Ocean Color Imagery. MS Thesis. University of South Florida, St. Petersburg, FL.

CANNIZZARO, J.P., K.L. CARDER, F.R. CHEN, and C.A. HEIL. 2002. Remote detection of red tide blooms on the West Florida Shelf: a novel classification technique. Proceedings of the Ocean Optics XVI Meeting, November/18-22.

CARVALHO, G.A., P.J., MINNETT, W., BARINGER, V., BANZON. 2007. Detection of Florida "red tides" from SeaWiFS and MODIS imagery. Proceedings of the XIII Simpósio Brasileiro de Sensoriamento Remoto (SBSR), Florianópolis, Brazil, INPE, April/21-26.

MOREL, A.. 1988. Optical modeling of the upper ocean in relation to its biogenous matter content (case I waters). Journal of Geophysical Research, 93: 10,749-10,768.

VAN DOLAH, F.M. 2000. Marine algal toxins: origins, health effects and their increased occurrence. Environmental Health Perspectives Supplements, 108(1): 133-141.

MEASUREMENT OF APOPTOSIS IN PHYTOPLANKTON USING FLOW CYTOMETRY AND ANNEXIN V BINDING

Francis J. Chrest and Daniel E. Terlizzi

University of Maryland, Center of Marine Biotechnology, 701 E. Pratt St., Baltimore, Md 21202

The study of bloom development and the effects of programmed cell death (PCD) on the growth and death of various phytoplankton species is the subject of current investigation. We have examined the role of apoptotic PCD through the use of the annexin V affinity assay and flow cytometry. The mechanism for annexin V binding to phosphatidylserine present on the surface of cells undergoing apoptosis has been widely reported in mammalian cells but application in phytoplankton to detect PCD has been limited. Initial measurements of apoptosis using flow cytometry and annexin V binding were evaluated in the unicellular chlorophyte, *Dunaliella tertiolecta*. Previous studies have shown that *D. tertiolecta* undergoes apoptotic PCD following 12 days of light deprivation indicated by caspase induction DNA fragmentation and morphological indicators (Segovia et. al. 2003). Consistent with these findings we found significant levels of annexin V binding and light scatter changes following light deprivation. Compared to normal light conditions, there was a marked increase (30%) in annexin fluorescence and a corresponding decrease in light scatter properties of cells from light deprived cultures. With the addition of the DNA dye propidium iodide, we could further document membrane damage in cells undergoing apoptosis. Using the described methodology we further evaluated the levels of annexin V binding in light deprived cultures of two dinoflagellates including *Amphidinium carterae* and *Karlodinium venificum*. Our initial findings indicate detectable levels of annexin fluorescence associated with light deprivation. Understanding the role of PCD in control of phytoplankton populations will provide insight into both bloom development and management.

Segovia, M., Haramaty, L., Berges, J.A., & Falkowski, P.G. 2003. Cell death in the unicellular chlorophyte *Dunaliella tertiolecta*. A hypothesis on the evolution of apoptosis in higher plants and metazoans. *Pl. Physiol.* 132, 1-7.

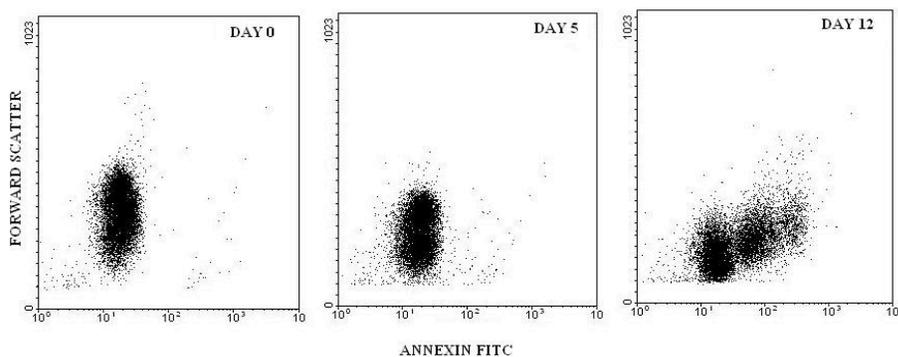


Fig. 1. Increases in Annexin-FITC binding and forward scatter in *D. tertiolecta* at 0, 5 and 12 days darkness.

SINGLE LABORATORY VALIDATION OF THE BREVETOXINS ELISA FOR SHELLFISH CONTAMINATION ASSESSMENT

Francie Coblentz¹ and Jerome Naar¹

¹Center for Marine Science, University of North Carolina Wilmington, Wilmington, NC, USA

The brevetoxins (PbTx_s) are a family of lipid-soluble polyether marine neurotoxins produced by the bloom-forming dinoflagellate *Karenia brevis*. These toxins are responsible for annual massive mortalities of fish and are also accountable for reoccurring mortalities of birds, sea turtles, and marine mammals. Lacking a protective theca, *K. brevis* cells lyse easily and the toxins, under the combined actions of wind and waves, can be aerosolized causing a profound concentration-dependent bronchoconstriction upon inhalation. Aside from respiratory impairments, these toxins also present a human health threat as they accumulate to dangerous levels in filter-feeding bivalve shellfish such as clams, oysters and mussels leading to Neurotoxic Shellfish Poisoning (NSP) if consumed.

In recent years, we have developed a high-sensitivity competitive ELISA for the detection of brevetoxins in numerous environmental and biological matrices. This assay is now routinely used as a diagnostic (and sometime forensic) tool to determine brevetoxin exposure, as a research tool to identify pathways of exposure in humans and animals, and to examine the fate of these toxins in the food web and the environment.

Because of the risk for NSP, shellfish contamination is intensively monitored during and after red tide events. As of today, the only approved method for monitoring is the regulatory mouse bioassay, which is slow to provide results and unethical to perform. Here we present results of a single laboratory validation study of a competitive ELISA to monitor brevetoxin contamination in three shellfish species of commercial interest. This study is the first step toward the official validation of the brevetoxin ELISA as an ethical, precise and rapid alternative method for the regulatory monitoring of these toxins in shellfish.

POPULATION STRUCTURE OF PARALYTIC SHELLFISH POISONING (PSP) RESISTANT SOFTSHELL CLAMS, *Mya arenaria*, IN EASTERN MAINE, USA

Laurie Connell, Scott Hamilton, and Amber Bratcher
School of Marine Sciences, University of Maine, Orono, ME

Atlantic Canadian populations of the softshell clam, *Mya arenaria*, with repeated exposure have been shown to be resistant to paralytic shellfish poisoning blooms (PSP) caused by the toxigenic algae *Alexandrium* spp. (MacQuarrie and Bricelj 2000; MacQuarrie 2002; Bricelj et al. 2005; Connell et al. 2006). This resistance is correlated with a mutation in the sodium channel gene pore region, the binding site for saxitoxin, a principle component of paralytic shellfish toxins (PSTs) (Bricelj et al. 2005). This mutation occurs at one base in Domain II resulting in a single amino acid change (Bricelj et al. 2005). A survey of geographically distant *M. arenaria* populations along the eastern seaboard and northwest coast of North America revealed that the resistant genotype occurs in many populations and that the mutation may have occurred multiple times in separate populations. Selective pressures exerted on clam populations by PSTs have not yet been specifically measured, but a survey of *Alexandrium* spp. bloom history coupled with the extent of PST exposure to various *Mya arenaria* populations may provide clues to the rate of expansion of a STX resistant phenotype.

This study surveyed *Mya arenaria* populations from eight locations in Eastern Maine, half of which have a history of repeated PSP blooms. The Domain II sodium channel pore region was sequenced and the populations were compared for percent of homozygous (resistant or sensitive) and heterozygous individuals. These data were then compared with PSP history and dominant ocean currents that can strongly effect larval recruitment. Results from this survey effect management decisions for local shellfish committees on the source of spat to be set in each bay as well as The Maine Department of Marine Resources biotoxin monitoring program.

- Bricelj, V. M., L. B. Connell, K. Konoki, S. P. MacQuarrie, T. Scheuer, W. A. Catterall and V. L. Trainer (2005). Na⁺ channel mutation leading to saxitoxin resistance in clams increases risk of PSP. *Nature* **434**: 763-767.
- Connell, L. B., S. P. MacQuarrie, B. M. Twarog, M. Iszard and V. M. Bricelj (2006). Population differences in nerve resistance to paralytic shellfish toxins in softshell clam, *Mya arenaria*, associated with sodium channel mutations. *Marine Biology* **150**: 1227-1236.
- MacQuarrie, S. P. and V. M. Bricelj (2000). Does the history of toxin exposure influence bivalve population responses in *Mya arenaria*? II) feeding, survival and toxin accumulation. *Journal of Shellfish Research* **19**(1): 636 (abstract).
- MacQuarrie, S. P. (2002). Inter- and intra-population variability in behavioral and physiological responses of the softshell clam, *Mya arenaria*, to the PSP toxin-producing dinoflagellate, *Alexandrium tamarense*. Halifax, NS, Dalhousie University: 141pp.

DATA MANAGEMENT SUPPORTING REGIONAL VOLUNTEER PHYTOPLANKTON MONITORING EFFORTS

Scott L. Cross¹, Steve L. Morton², Rost Parsons¹

¹NOAA/NESDIS, National Coastal Data Development Center, Bldg. 1100, Stennis Space Center, MS 39522

²NOAA/NOS, Marine Biotoxins Program, 291 Fort Johnson Road, Charleston, SC 29412

The Southeastern Phytoplankton Monitoring Network (SEPMN) is an education and outreach program developed by NOAA's National Ocean Service (NOS) to engage school and community volunteer groups in phytoplankton sampling and identification and to raise awareness of harmful algal blooms. Over the past year the program included approximately 2000 volunteers, mostly secondary students and teachers, sampling 80 sites from North Carolina to Texas. Recently, NOAA's Coastal Data Development Center (NCDDC) has partnered with SEPMN to create an end-to-end data-management system that enhances the volunteer experience, facilitates quality control, and ensures the integrity of the data set over the long term.

Volunteers are able to take advantage of an online data-entry tool to submit data, and are then able to visualize and analyze their own validated data as well as data from their Network peers in an online Geographic Information System (GIS) environment. The data-entry tool is a password-protected website offering digital datasheets that perform simple error checking (units, expected data ranges) to ensure accuracy. The resulting data sets enter a workflow that subjects each record to authorization from SEPM administrators before it is automatically entered into a relational database. Data are then exposed to a web GIS application (<http://www.ncddc.noaa.gov/website/SEPMN/viewer.htm>) that allows students to map phytoplankton abundance measurements across the entire spatial and temporal extents of the SEPMN data set, along with their own submissions. These records include ancillary environmental data (water temperature, salinity), so that by integrating the available data, students can begin to form hypotheses on phytoplankton responses to environmental forcing. In addition, real-time ocean-temperature data from NOAA data buoys can be mapped, allowing students to begin to make testable predictions about phytoplankton occurrences prior to sampling. Finally, the database and associated metadata is periodically uploaded to NOAA's National Oceanographic Data Center for archival, ensuring the long-term integrity of this unique data set.

QUANTITATIVE MAPPING OF CYANOBACTERIAL BLOOMS FROM OCEANSAT-1 OCM SATELLITE DATA: AN EMPIRICAL APPROACH

Padmanava Dash¹, Nan D. Walker¹, Ana Cristina Garcia¹, Sibel Bargu-Ates¹ and James L. Pinckney²

¹Louisiana State University, Department of Oceanography and Coastal Sciences, Baton Rouge, LA 70803

²University of South Carolina, Department of Biological Sciences, Columbia, SC 29208

We present the preliminary results of the first stage of a NASA-funded project which comprises the development of algorithms for mapping cyanobacterial blooms from Oceansat-1 OCM (Ocean Color Monitor) satellite data. This sensor was chosen over SeaWiFS (Sea-Viewing Wide Field-of-view Sensor) and MODIS (Moderate-resolution Imaging Spectroradiometer) due to its superior spatial resolution (250m compared with 1000m pixels). The OCM and MODIS data were obtained in real-time at the LSU Earth Scan Lab direct-broadcast station for Earth Orbiting satellites. In the years 2006-2007, extensive bi-weekly and weekly field campaigns were undertaken from November 2006 through the present to a freshwater lake (salinity < 1), Lac des Allemands, located in the north-western portion of the Barataria estuary system, Louisiana. Weekly sampling trips will be maintained through summer and then bi-weekly sampling through November 2007, to complete an entire year of field data collection. We have obtained a database of photosynthetic pigment concentrations and phytoplankton composition which are being used in tandem with measurements from the OCM sensor to quantify blooms from space. The field data collection includes water samples for HPLC (High Performance Liquid Chromatography), CDOM (Colored Dissolved Organic Matter), TSS (Total Suspended Solids), microscopic analyses, the cyanobacteria unique pigment phycocyanin, and toxin analyses (Table 1). The results have exceeded our expectations, in particular the number of clear-sky OCM and MODIS images which have been fortuitously obtained on seven days, thus far. Preliminary analyses of the data obtained demonstrate clear seasonal variation in chlorophyll *a* concentrations in Lac des Allemands and in the pigments associated with cyanobacteria. Overall, concentrations of pigments were highest in fall and spring and lowest in winter, corresponding most closely with variation in temperature and light conditions. The microscopic analysis revealed the presence of numerous toxic species of cyanobacteria including *Anabaena circinalis*, *Cylindrospermopsis raciborskii* and *Microcystis* sp. The OCM true color and chlorophyll *a* images and MODIS true color images corroborate with the field measurements and our preliminary results towards the development of regional algorithms for chlorophyll *a* and cyanobacteria concentrations will be presented.

Table 1 Field data

Date	Analyses						YSI	Radiometer	Water Depth
	HPLC	CDOM	TSS	Microscopic	Phycocyanin	Toxin			
Sep 11, 2006	x								
Nov 17, 2006	x		x				x		
Dec 19, 2006	x	x	x	x			x		
Jan 10, 2007	x	x	x	x			x	x	
Feb 19, 2007	x	x	x	x			x		
Mar 23, 2007	x	x	x	x			x	x	
Mar 29, 2007	x	x	x	x	x		x		x
Apr 12, 2007	x	x	x	x	x	x	x		x
Apr 20, 2007	x	x	x	x	x	x	x	x	x
Apr 26, 2007	x	x	x	x	x	x	x		x
May 14, 2007	x	x	x	x	x	x	x		x
May 18, 2007	x	x	x	x	x	x	x	x	x
May 24, 2007	x	x	x	x	x	x	x	x	x
June 01, 2007	x	x	x	x	x	x	x	x	x
June 15, 2007	x	x	x	x	x	x	x	x	x

**THE PACIFIC NORTHWEST HAB BULLETIN PILOT PROJECT:
TECHNICAL DEVELOPMENT OF AN OCEAN OBSERVING INFORMATION SYSTEM FOR
THE PROTECTION OF HUMAN HEALTH**

Sheryl A. Day¹, Barbara M. Hickey², Rita A. Horner², Dan L. Ayres³, Frank H. Cox⁴, Jerry A. Borchert⁴,
Miranda S. Wecker⁵, Anthony Odell⁵, Joe Schumacker⁶, and Vera L. Trainer¹

¹ Northwest Fisheries Science Center, 2725 Montlake Boulevard East, Seattle, WA 98112, USA

² University of Washington, School of Oceanography, Seattle, WA 98195, USA

³ Washington State Department of Fish and Wildlife, 48 Devonshire Road, Montesano, WA 98563, USA

⁴ Washington Department of Health, 111 Israel Road Southeast, Tumwater, WA 98501, USA

⁵ Olympic Natural Resources Center, 1455 South Forks Avenue, Forks, WA 98331, USA

⁶ Quinault Indian Nation, PO Box 189, Taholah, WA 98587, USA

The Pacific Northwest HAB Bulletin Pilot Project, with support from the Centers for Disease Control, is a web-based information dashboard developed to provide a comprehensive early warning information system for Washington coast HAB events. Traditionally, HAB-related data are maintained by several organizations in separate locations and in a variety of formats. In addition, no single location or system exists to capture the tacit knowledge of local experts who could identify trends and make calculated predictions based on collected data. The separate locations of these resources result in a lag time between acquiring, correlating, interpreting, and distributing HAB event information to coastal managers. The PNW HAB Bulletin Pilot Project builds upon the successful ORHAB (Olympic Region Harmful Algal Blooms) monitoring program by automating the aggregation of HAB information – biological and physical data as well as summarized conclusions by local experts – into a single location. This information dashboard is a dynamic, database-driven web page template comprised of the following components: Real-time drifter tracks, domoic acid in shellfish, *Pseudo-nitzschia* cell counts, sizes, and critical assessment level indicators, winds (Cape Elizabeth buoy data from NDBC), model currents from an existing Navy operational model, Columbia River model, and corresponding analyses by local experts describing the data and the likelihood of occurrence of HAB events. Several technologies and applications were utilized to generate the dashboard from various data sources; these included custom scripts written in UNIX, PERL, Visual Basic, and JavaScript, customized GIS and plot applications, a MySQL database, PHP, XML, and HTML. This combination of technologies allowed data to be automatically retrieved and manipulated from existing resources, including ARGOS satellite servers, websites, spreadsheets, and databases, with minimal or no extra effort required from the data providers. Automating the retrieval of HAB-related data into a single location allowed the PNW HAB Bulletin to significantly reduce the timeline for delivering HAB information to coastal managers, and multiple technologies allowed this data to be aggregated without requiring HAB information providers to modify their existing information storage methods.

NEW PERFORMANCE DATA FOR *IN SITU* AND SIMULATED EXPERIMENTS USING THE PHYTOFLASH SUBMERSIBLE ACTIVE FLUOROMETER

Chelsea Donovan¹, Lawrence Younan¹, Raphe Kudela² and Tawyna Peterson²

¹Turner Designs, Inc., Sunnyvale, CA 94085, USA

²University of California at Santa Cruz, Inc., Santa Cruz, CA, USA

An *in situ* variable fluorescence system has been developed that will allow real-time measurement of the primary variable fluorescence variables: F_v , F_o and F_m . Advances in solid-state light detectors and the development of advanced signal processing circuitry have led to the development of a new generation of fluorescence instrumentation that can be used to measure photosynthetic parameters in a wider range of platforms and locations. Practical applications include the detection of the onset of algal blooms, indication of nutrient status of planktonic algae, ballast water monitoring, as well as the measurement of algal community change. Performance data presented represents red tide distribution, simulated system impacts in relation to the yield and quantum efficiency of *Crocospaera* (a nitrogen-fixing cyanobacterium) along a transect.

FAST AND ACCURATE DETECTION OF *Alexandrium* SPECIES USING PEPTIDE NUCLEIC ACID PROBES AND SURFACE PLASMON RESONANCE

Janice Duy and Laurie Connell
University of Maine, Orono, ME

Rapid detection of some members of the toxigenic alga *Alexandrium* is of paramount importance in cool temperate waters since these organisms produce a suite of toxins responsible for paralytic shellfish poisoning (PSP). Since *Alexandrium* blooms may not produce cell populations large enough to be visible and these may be below the water surface, early detection can lead to beach closures and fishing restrictions which can prevent human ingestion of contaminated shellfish.

Current identification for field groups still is based on microscopy, which is time consuming and requires skilled personnel. Our proposed detection mechanism involves functionalizing gold surfaces with peptide nucleic acid (PNA) probes and then sensing binding (hybridization) of target sequences through surface plasmon resonance (SPR).

PNAs, which are DNA analogs with a backbone composed of repeating N-(2-aminoethyl)-glycine units linked by peptide bonds, were selected for the probe layer because of their suitability for field use: PNAs are resistant to cleaving by nucleases and hybridize under a broad variety of conditions. SPR is a good choice for the detection of target hybridization because this technique can accurately measure thickness changes on adsorbed species on surfaces such as gold.

Preliminary experiments using atomic force microscopy (AFM) imaging and ellipsometry show that cysteine-terminated PNA sequences attach to flat gold surfaces through gold-thiol bonds. The PNAs used are biotin-terminated and surface attachment can be verified through biotin quantitation, while hybridization efficacy can be confirmed through binding of complementary labeled DNA.

PIGMENT COMPOSITION OF THE TEXAS STRAIN OF *Prymnesium parvum* DURING LOG, STATIONARY, AND SENESCENT GROWTH PHASESRM Errera¹, JL Pinckney², DL Roelke³¹National Oceanic and Atmospheric Administration, Office of Oceanic and Atmospheric Research, Laboratories and Cooperative Institutes, Silver Spring, MD²Marine Science Program and Department of Biological Sciences, University of South Carolina, Columbia, SC ³Department of Wildlife and Fisheries Sciences, Department of Oceanography, Texas A&M University, College Station, TX

The harmful algal species, *Prymnesium parvum*, has been identified in fresh, brackish and coastal environments worldwide. In Texas, *P. parvum* blooms have increased in frequency and intensity over the last 7 years. Based on fish kills often associated with *P. parvum* blooms, rapid identification of *P. parvum*'s presence in the water column is necessary for mediation. We employed Chemical Taxonomy (ChemTax) as a rapid, alternative diagnostic tool. Using measured photopigment concentrations, ChemTax can estimate biomasses of bulk or specific phytoplankton taxa; however, ChemTax is sensitive to the initially specified pigment ratios. This sensitivity may prove problematic when different strains of phytoplankton species have different pigment ratios. We performed incubation experiments to determine if the Texas strain of *P. parvum* correlated with pigment ratios identified in a strain of *P. parvum* from Norway (Rodriguez et al. 2006). The Norway strain of *P. parvum* was chosen because of its similar pigment signature to the Texas strain. We determined that Texas *P. parvum* pigment ratios were dynamic as a function of cell density and physiological state. We then used our ratios and the Norwegian ratios for *P. parvum* to estimate *P. parvum* concentrations for in-lake experiments performed in Lake Whitney, Texas. We discovered that the ChemTax model initialized with the Norwegian ratios underestimated *P. parvum*'s contribution to the total chlorophyll *a* signature. In addition, we noted the presences of zeaxanthin within the Texas strain of *P. parvum*. Our results indicate that Texas *P. parvum* may be showing signs of adaptation to the higher light intensity conditions in this semi-tropical habitat.

Rodriguez F, Chauton M, Johnsen G, Andresen K, Olsen LM, Zapata M. 2006. Photoacclimation in phytoplankton: implications for biomass estimates, pigment functionality and chemotaxonomy. Mar Bio 148:963-971

FACTORS AFFECTING MICROCYSTIN CONCENTRATIONS AND CELL QUOTAS IN THE GREAT LAKES

G Fahnenstiel¹, J Dyble¹, D Millie², P Tester³, and W Litaker³

gary.fahnenstiel@noaa.gov; 231-759-7824

¹ LMFS/GLERL/NOAA, Muskegon, MI; ² FIO/USF, St. Petersburg, FL;

³ CCFHR/NOAA, Beaufort, NC.

As part of GLERL's new Ocean and Human Health Center, an interdisciplinary research program was initiated to determine the factors controlling microcystin production (cell quota) in the Great Lakes. In Saginaw Bay microcystin concentrations were strongly correlated with correlated with *Microcystis aeruginosa* abundance and total phosphorus concentrations. Microcystin cell quotas averaged 140 fg cell⁻¹, and were not correlated with any environmental factor or growth rates. A series of experiments were conducted with Saginaw Bay and western Lake Erie water to determine the influence of nutrients and light in controlling microcystin cell quota. Significant decreases in cell quota were noted with phosphorus additions and irradiance decreases; however, the decreases were limited (most 2-4X). Environmental factors such as nutrients and light appear to be more important in controlling abundance and growth rate of *Microcystis aeruginosa* than cellular pathways of microcystin synthesis.

**UNUSUALLY HIGH LEVELS OF DOMOIC ACID IDENTIFIED IN MINKE WHALE
(*Balaenoptera acutorostrata*) STRANDING DURING CALIFORNIA *Pseudo-nitzschia* BLOOM**

Spencer E. Fire¹, Steve L. Morton¹, Zhihong Wang¹, Michelle Berman²

¹NOAA-National Ocean Service, 219 Fort Johnson Rd, Charleston, SC 29412

²Santa Barbara Museum of Natural History, 2559 Puesta del Sol Rd, Santa Barbara, CA 93105

Blooms of the diatom genus *Pseudo-nitzschia* have been implicated in several large-scale marine animal mortality events along the California coast in recent years, but little toxicological data are available for cetaceans affected by these blooms. The algal toxin domoic acid was detected in several species of cetaceans from an April 2007 marine mammal die-off associated with a Southern California bloom of the diatom *Pseudo-nitzschia*. Domoic acid in minke whale (*Balaenoptera acutorostrata*) feces was detected by liquid chromatography-mass spectrometry (LC-MS) at concentrations of 258 $\mu\text{g/g}$, exceeding the highest reported values for any marine mammals in this region. Scanning electron microscope analysis of the fecal sample detected a high abundance of frustules of the diatom *Pseudo-nitzschia australis*. The minke whale diet consists mainly of schooling planktivorous fish and krill, organisms that previous research has demonstrated as important vectors of domoic acid to higher trophic levels. These data provide direct evidence for the accumulation of domoic acid in this whale species, at potentially lethal concentrations.

ASSESSMENT OF AN OPERATIONAL HARMFUL ALGAL BLOOM FORECAST SYSTEM FOR THE EASTERN GULF OF MEXICO: A COMPARATIVE ANALYSIS OF SUCCESS AND UTILIZATION THROUGH TWO BLOOM SEASONS

Kathleen M. Fisher¹, Zachary E. Bronder¹, Lori E. Fenstermacher¹, Heidi M. Keller¹, Allison L. Allen¹, Mark S. Vincent¹, Richard P. Stumpf², Michelle C. Tomlinson²

¹NOAA, National Ocean Service, Center for Operational Oceanographic Products and Service (CO-OPS), Silver Spring, MD 20910, USA

²NOAA, National Ocean Service National Centers for Coastal Ocean Science (NCCOS), Silver Spring, MD 20910, USA

Blooms of a toxic dinoflagellate, *Karenia brevis*, occur nearly every year on the Gulf coast of Florida with damaging consequences linked to fish kills, marine bird and mammal deaths, human respiratory illness, shellfish bed closures and loss of local tourism. To assist in mitigating local damages resulting from harmful algal blooms (HABs), the Gulf of Mexico HAB Operational Forecast System (GOM HAB-OFS) for HAB detection was transitioned from research to operational status in October 2004 through a multi-office effort of NOAA. Presently in its fourth year of operation, the ecological forecast system issues bulletins twice weekly to coastal resource managers, federal and state agencies, and academic institutions with detailed information concerning the possible presence or confirmation of new harmful blooms, forecasts of spatial bloom extents, movement, intensification conditions, and the daily potential for coastal impacts in the eastern Gulf of Mexico. Since the GOM HAB-OFS' implementation, 7 substantial harmful blooms have been identified and tracked by ecological forecasters in over 250 bulletins utilizing SeaWiFS satellite imagery, past and forecasted winds, a wind transport model and sampling data. The operational system is refining methods for evaluating utilization and skill of the bulletin forecasts based on an assessment of the first two operational years from October 2004 through September 2006. Utilization is determined by the frequency of user response to bulletin information; while skill is evaluated on the accuracy of bloom identification and forecast components. A comparative analysis of bulletin utilization and forecast accuracy during the Forecast System's first two bloom seasons, occurring November 2005 to April 2007, is presented.

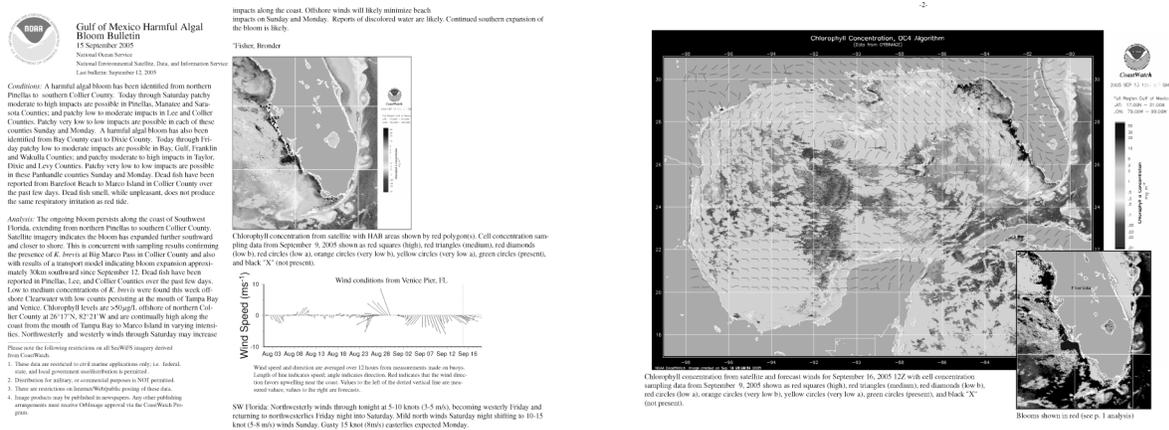


Figure 1. Example Operational Bulletin Distributed by the NOAA Gulf of Mexico HAB Operational Forecast System

DELETERIOUS EFFECTS OF A NON-PSP BIOACTIVE PRODUCED BY *Alexandrium tamarens* ON BIVALVE HEMOCYTES

Susan E. Ford¹, V. Monica Bricelj², Christophe Lambert³ and Christine Paillard³

¹Haskin Shellfish Research Laboratory, Rutgers University, Port Norris, NJ 08349 USA; ²National Research Council, Institute for Marine Biosciences, Halifax, N S B3H 3Z1, Canada; ³Institut Universitaire Européen de la Mer, LEMAR, 29280 Plouzané, France

The known negative effects of paralytic shellfish toxin (PST)-producing dinoflagellates on feeding, burrowing and survival of some bivalve mollusks has prompted questions concerning whether they might also impair the immune system and thus make the affected bivalves more susceptible to disease agents. The primary components of the cellular defense system are hemocytes, circulating cells similar to mammalian white blood cells. Recent *in vivo* and *in vitro* studies have failed to detect substantial or consistent impacts on hemocyte properties or functions as a consequence of exposure to intact PST-producing dinoflagellates. However, hemocytes would most likely be exposed to intracellular toxins only after the algae are consumed and digested, and the water-soluble toxins released. Therefore, we conducted a series of experiments in which hemocytes from two suspension-feeding bivalves – the Manila clam, *Ruditapes philippinarum*, and the softshell clam, *Mya arenaria* – were exposed *in vitro* to filtered extracts of one highly toxic PST-producing and one non-PST-producing strain of *Alexandrium tamarens* (isolates PR18b, 76 ± 6 STXeq cell⁻¹ and CCMP115, containing undetectable PST, respectively). We measured adherence and phagocytosis, two hemocyte attributes previously shown to be compromised by bacterial pathogens and other stressors. The response of hemocytes from individual clams was determined. We found no measurable effect of a cell-free extract from a highly concentrated suspension of the PST-producing strain on hemocytes of either bivalve species. Instead, extract from the non-PST producing strain had a consistent negative effect on hemocytes of the two species, resulting in significantly lower adherence and phagocytosis compared to strain PR18b and filtered seawater controls. The bioactive produced by strain CCMP115, which has yet to be characterized, may be similar to the PST-independent allelopathic compounds described for *Alexandrium* spp., which act on other plankters (Tillmann and John, 2002). These compounds and those produced by other harmful algae are known to cause immobilization, cellular deformation and lysis of co-occurring target organisms, and their cytotoxic effects may be similar to those of ichthyotoxins (Tillmann et al., in press). Thus, non-PST producing *Alexandrium* spp., which do not cause paralysis and burrowing incapacitation of clams (Bricelj et al., 2005), may still produce a bioactive that has negative effects not only on hemocytes, but on other molluscan cell types and their functions, as well.

BRICELJ, V.M., L. CONNELL, K. KONOKI, S.P. MACQUARRIE, T. SCHEUER, W.A. CATERALL, and V.L. TRAINER. 2005. Sodium channel mutation leading to saxitoxin resistance in clams increases risk of PSP. *Nature* **434**: 763-767.

TILLMANN, U. and U. JOHN, 2002. Toxic effects of *Alexandrium* spp. on heterotrophic dinoflagellates: an allelochemical defence mechanism independent of PSP-toxin content. *Marine Ecology Progress Series* **230**: 47-58.

TILLMANN, U., JOHN, U., KROCK, B. and A. CEMBELLA. Allelopathic effects of bioactive compounds produced by harmful algae. *Proceedings of the 2007 International Harmful Algae Conference, Copenhagen, in press.*

COUPLING PHYSIOLOGICAL RESPONSES OF THE TOXIC HAPTOPHYTE *Prymnesium parvum* TO PATTERNS IN GENE EXPRESSION

Michael F. Freitag¹, Uwe John¹ & AD Cembella¹

¹Alfred Wegener Institute for Polar and Marine Research, 27576 Bremerhaven, Germany

A normalized cDNA library² from the toxic haptophyte *Prymnesium parvum* was used to design an oligonucleotide-based microarray platform. Allelopathic interactions were investigated between *P. parvum* strain RL10 and the heterotrophic dinoflagellate *Oxyhris marina*. Cultures of *Prymnesium* were incubated together with both whole cells and cell-free filtrate from *Oxyhris* cultures. During incubation with *Prymnesium*, the presence of *Oxyhris* cells, compared to cell-free culture medium, showed no differential effect on the gene expression profile of the haptophyte. A bioassay measuring the toxicity of *Prymnesium* against the sensitive cryptophyte *Rhodomonas baltica* was additionally performed. In this case, *Rhodomonas* viabilities following incubation with *Prymnesium* and *Prymnesium* previously incubated with *O. marina* medium were virtually identical. This suggests no correlation between the presence of *O. marina* medium, and induced toxicity in *P. parvum*. Similar experiments, involving the dinoflagellate *Heterocapsa rotunda* and an unclassified toxic croccal cyanobacteria strain: NIVA CYA 331, are currently underway. All tentative unique genes (TUGs) identified from the microarray hybridization will be further classified through the use of a *Prymnesium* fosmid library prepared from axenic cultures.

²Sára Beszteri, Alfred Wegener Institute, Bremerhaven Germany

BIOACCUMULATION OF CYANOBACTERIAL CYLINDROSPERMOPSIS TOXIN IN LOUISIANA BLUE CRAB, *Callinectes sapidus*

Ana Cristina Garcia¹, Padmanava Dash¹, and Sibel Bargu-Ates¹, ¹Louisiana State University Department of Oceanography and Coastal Sciences, Baton Rouge, LA 70803, USA

Life history characteristics, specifically a preference for feeding on bottom-dwelling filter-feeding organisms, make the blue crab (*Callinectes sapidus*) vulnerable to the accumulation of toxins. The blue crab commercial fishery of Louisiana is one of the largest crab fisheries in the United States in terms of biomass (Guillory and Roberts 1997). Lac des Allemands, located within the Barataria estuary system of Louisiana, serves as a critical nursery ground for blue crab, but is also an area often abundant in species of *Cylindrospermopsis*. Cylindrospermopsin (CYN), produced by *Cylindrospermopsis raciborskii*, *Umezakia natans*, and *Aphanizomenon ovalisporum*, is an alkaloid hepatotoxin that may also cause damage to the kidneys, thymus, and heart. Previous studies have also shown that CYN can break DNA strands, implicating the compound as a possible carcinogen (Saker et al. 2004). Enzyme-linked immunosorbent assay (ELISA), coupled with epifluorescent and scanning electron microscopy, are used to perform cell counts of *Cylindrospermopsis* species, evaluate their toxicity, and quantify CYN present within tissue samples of blue crab taken from Lac des Allemands. By analyzing the concentrations of CYN specifically within edible portions of blue crab, risk to human consumers is determined.

Guillory, V., and K. Roberts. 1997. Status of Louisiana blue crab resource. Summary of Proceedings, Coastal Fishing '97: Use of Louisiana's blue crab resource. Louisiana Sea Grant College Program: Louisiana State University: 1-28.

Saker, M., J. Metcalf, G. Codd, and V. Vasconcelos. 2004. Accumulation and depuration of the cyanobacterial toxin cylindrospermopsin in the freshwater mussel *Anodonta cygnea*. *Toxicon* **43**: 185-194.

DOMOIC ACID IN BENTHIC COMMUNITIES OF THE SANTA CRUZ MUNICIPAL WHARF IN MONTEREY BAY CALIFORNIA

Suzanne M. Garcia¹, Fernanda Mazzillo², Mary W. Silver³, University of California Santa Cruz, Santa Cruz, CA USA

Fish caught from piers are an important and often overlooked part of the recreational fishery. Generally, recreationally caught fish are not subject to the usual state-implemented monitoring programs for phycotoxins. Santa Cruz Municipal Wharf is one such heavily fished pier located in the northern end of Monterey Bay in central California. Some species of fish caught from this pier have been found to possess domoic acid (Fire and Silver 2005), a water soluble phycotoxin locally produced by several species of the diatom *Pseudo-nitzschia*. The diet of fish caught from this nearshore environment varies from species to species and also may differ considerably from that of offshore species. The exact pathway by which pier-caught fish acquire domoic acid (DA) is not well known. This study focuses on the presence of DA in various benthic prey species of fish caught from the wharf. During four separate sampling events, divers collected a variety of benthic invertebrates from the more inshore region of the wharf and another set from the outer, deeper (~6 m) end of the Wharf, including samples taken directly from the pier pilings. The species selected for collection were confirmed prey items, as they had been observed in the stomachs of pier-caught fish (Mazzillo personal communication 2007). Some specimens collected were not confirmed prey items, however, divers collecting the samples had observed fish feeding upon these organisms. Domoic acid levels were measured in the viscera of the invertebrate specimens using both HPLC (High Performance Liquid Chromatography) and ELISA methods, the results of which will be discussed. It is anticipated that the benthic community is an important contributor of DA to the food web, and a few recent studies have shown the presence of DA in several local benthic invertebrates. The extent to which particular benthic organisms acquire domoic acid and also serve as important food sources for commonly caught fish, should be a predictor of contamination of those fish. By understanding the routes by which DA is delivered through the diet of commonly caught pier fish, the regulatory community may better anticipate which fish species will be most contaminated during toxic *Pseudo-nitzschia* events and thereby institute protections for recreational fishermen who consume fish caught from the Wharf.

METABOLISM OF ALGAL STEROLS BY BAY SCALLOPS AND BRINE SHRIMP

José-L. Giner,¹ Hui Zhao,¹ Mark S. Dixon,² and Gary H. Wikfors²

¹Department of Chemistry, SUNY-ESF, Syracuse, NY 13210.

²NOAA, NMFS, 212 Rogers Avenue, Milford, CT 06460

Many HAB species contain sterols with unusual structures. We have hypothesized that these sterols serve as chemical defenses by interfering with the nutrition and growth of marine invertebrates (Giner et al.). There are multiple mechanisms by which such sterols might exert harmful effects. They may be refractory to the normal bioconversion of dietary sterols to cholesterol, leading to a deficiency of the cholesterol needed for cell growth. Alternatively, unusual marine sterols may inhibit enzymes involved in the production of physiologically important steroidal compounds, such as the arthropod molting hormone ecdysterone. To better understand the fate of algal sterols in mollusks and crustaceans, metabolic studies were carried out. Methods for the synthesis of substantial quantities of algal sterols were developed and used to prepare specifically ¹³C-labeled compounds. These were incorporated into the microalgal diets of juvenile bay scallops (*Argopecten irradians*) and brine shrimp (*Artemia salina*). Analysis by ¹³C-NMR spectrometry showed the metabolic fates of the sterols. Addition of a sterol bearing the label in a different position provided a positive internal control in cases where no bioconversion of the test sterol was detected. The brine shrimp were found to be better at metabolizing algal sterols than scallops, and specific sterols were better metabolized than others. The brown tide sterol 24-propylidenecholesterol was metabolized to cholesterol, but dinoflagellate sterols brevesterol and dinosterol were not and accumulated unchanged in the organisms.

Giner, J.-L.; Faraldos, J. A.; Boyer, G. L., "Unique Sterols of the Toxic Dinoflagellate *Karenia brevis* (Dinophyceae): A Defensive Function for Unusual Marine Sterols?" *J. Phycol.* **2003**, 39, 315-319.

POTENTIAL GOOD IN A “SEA” OF HARM: CHARACTERIZATION OF A MAMMALIAN RECEPTOR ASSOCIATED WITH VOLTAGE-SENSITIVE SODIUM CHANNELS USING BREVENAL PRODUCED BY *Karenia brevis*

Elena P. Gold^{1,2}, Henry M. Jacocks², Sophie Michelliza² and Daniel G. Baden^{1,2}

¹University of North Carolina Wilmington; ²Center for Marine Science, Wilmington, NC 29403, US

Blooms of the marine dinoflagellate *Karenia brevis* are known to produce brevetoxins, which cause neurotoxic and upper respiratory problems in humans and mammals and are both toxic and lethal to many marine organisms. In contrast, these dinoflagellates also produce an antitoxin, brevenal, shown to be helpful in the treatment of brevetoxin poisoning in manatees and blocks brevetoxin-induced bronchoconstriction in sensitized sheep. Brevenal also protects against the deleterious effects of brevetoxin in fish bioassays. Although the mechanism of action in respiratory systems is poorly understood, it has been established that in mammalian neuronal receptors, brevetoxins act on site 5 of voltage sensitive sodium channels (VSSC) causing four distinct effects: 1) shifting the action potential to more negative values, 2) prolonging channel open time, 3) inhibiting inactivation and 4) inducing subconductance states for Na⁺ across the channel. In addition to the brevetoxin receptor, there are five other known receptors located on the alpha subunit of VSSC.

Earlier studies suggested that brevenal is a direct competitor for site 5 using rat brain synaptosomal receptor binding assay (Boudelais *et al.*, 2004). Our results indicate that brevenal indirectly competes with brevetoxin as the result of an allosteric interaction between a complex of receptors where both brevetoxin and brevenal have unique receptors of their own. Investigation into the brevenal receptor was performed by using a novel radiolabeled ligand, tritiated brevenol (³H-Brevenol)—a reduced product of brevenal. Our receptor binding studies indicate that while brevenal reduces labeled brevetoxin-3 (³H-PbTx-3) associated with receptor site 5, brevetoxin does not reduce specific binding of ³H-Brevenol from its specific binding site. The brevetoxin receptor has an affinity in the range of 1-7 nM while the affinity of brevenal for its receptor is in the 95-100 nM range. Nevertheless, no other VSSC-specific ligand reduces ³H-Brevenol binding. These results suggest the existence of a novel receptor/pharmacore for brevenal/ol. Furthermore, we believe that ³H-brevenol will be an important probe in elucidating the effects of brevenal in several physiological systems. Thus, brevenal is the good in the “sea” of harm caused by the actions of brevetoxins produced *K. brevis*.

Bourdelsais, A.J., Campbell, S., Jacocks, H., Naar, J., Wright, J.L.C., Carsi, J., and Baden D.G. (2004). Brevenal is a Natural Inhibitor of Brevetoxin Action in Sodium Channel Receptor Binding Assays. *Cell. Mol. Biol.* 24(4):553-563.

**HARMFUL CYANOBACTERIAL BLOOM PROLIFERATION IN FLORIDA BAY, FL, USA:
ZOOPLANKTON GRAZING AND THE ROLE OF SALINITY**

Jennifer A. Goleski¹ and Christopher J. Gobler

School of Marine and Atmospheric Sciences, Stony Brook University, Stony Brook, NY 11794-5000

Florida Bay has been plagued with a series of ecological disturbances since the late 1980s including the proliferation of harmful cyanobacterial blooms caused by *Synechococcus*. Eutrophication is often suspected to promote algal blooms and nutrients have been a focus of Florida Bay water management and restoration efforts. However, an absence of adequate grazing pressure may also be an important factor in the proliferation of these blooms. Hypersaline conditions are known to occur in Florida Bay and some zooplankton are sensitive to such conditions. Zooplankton grazing can also be disrupted by the biochemical characteristics of cyanobacteria, further promoting blooms. For this study, *in situ* grazing rates of meso- and micro-zooplankton were determined within nine major basins of Florida Bay during all four seasons. We observed that growth rates of the phytoplankton community, up to $1.2 \pm 0.1 \text{ d}^{-1}$, typically exceeded microzooplankton grazing rates during algal blooms in the North-Central and Southern basins where blooms are prevalent. At these sites, $< 20\%$ of the standing stock was consumed by microzooplankton per day, a percentage below average for estuarine ecosystems. During a July 2006 bloom event, the grazing rate by microzooplankton on the total phytoplankton standing stock was $1.0 \pm 0.2 \text{ d}^{-1}$ while microzooplankton grazed at a rate of $0.33 \pm 0.16 \text{ d}^{-1}$ on the *Synechococcus* standing stock. Moreover, increased concentrations of mesozooplankton periodically elicited little or no change in phytoplankton growth and occasionally yielded increases in growth. As such, an absence of adequate grazing pressure may allow *Synechococcus* blooms to occur in Florida Bay. Cell-specific grazing rates by micro- and mesozooplankton and the role of trophic cascades in bloom occurrence will be also presented.

GLYCOLIPID FAMILIES IN PERIDININ-CONTAINING DINOFLAGELLATES

Cynthia G. Gray, Andrew D. Lasiter, Jeffrey D. Leblond

Department of Biology, Middle Tennessee State University, Murfreesboro, TN 37132, USA

As the engines of photosynthetic productivity in bloom-forming dinoflagellates, chloroplasts play a crucial role in energy harvesting and conversion. The photosynthetic machinery of dinoflagellates, as well as all other algae and plants, is housed within a matrix thought to be composed primarily of two glycolipids, monogalactosyldiacylglycerol (MGDG) and digalactosyldiacylglycerol (DGDG). It is only recently, with the advent of analytical techniques such as electrospray ionization-mass spectrometry (ESI-MS) and electrospray ionization-mass spectrometry/mass spectrometry (ESI-MS/MS), that two key questions regarding these lipids can be addressed:

- 1.) What are the intact forms of MGDG and DGDG present in dinoflagellates, and
- 2.) How are these forms biosynthesized?

This study addresses the lack of information regarding dinoflagellate glycolipid composition by presenting the results of a full-scan (mass range of 600-1,200 daltons) positive ion ESI-MS survey of the glycolipids of 35 peridinin-containing dinoflagellates, including several harmful species. Further analysis using positive ion ESI-MS/MS was performed to determine the positional distribution of fatty acids associated with MGDG and DGDG.

The dinoflagellates examined were divided into two groups based on the forms of MGDG and DGDG present. The first had lipid masses representing 18:5/18:4 MGDG (with the 18:5 fatty acid attached to the *sn*-1 position of the glycerol and the 18:4 fatty acid attached to the *sn*-2 position), 18:5/18:5 MGDG, 18:5/18:4 DGDG, and 18:5/18:5 DGDG. The second group of dinoflagellates had 20:5/18:4 MGDG, 20:5/18:5 MGDG, 20:5/18:4 DGDG, and 20:5/18:5 DGDG. Of the genera represented by more than one species, 4 genera, including *Amphidinium* and *Coolia*, were present only in the C₂₀/C₁₈ glycolipid group, while *Symbiodinium* species were confined to the C₁₈/C₁₈ group. *Alexandrium*, *Gymnodinium*, and *Prorocentrum* species were present in both groups. The differentiation between C₁₈/C₁₈ and C₂₀/C₁₈ glycolipids indicates differing biosynthetic pathways, as well as a possible evolutionary divergence between the two dinoflagellate groups.

DEVELOPMENT AND VALIDATION OF A NOVEL *IN VIVO* NITRATE REDUCTASE ACTIVITY ASSAY

Jennifer J. Griffith¹, Alicia Mangum², Gulnihal Ozbay², Mark E. Warner¹ and Kathryn J. Coyne¹

¹University of Delaware College of Marine and Earth Studies, Lewes, DE 19958, USA

²Delaware State University Department of Agriculture and Natural Resources, Dover, DE 19901, USA

Differences in nitrogen utilization can significantly impact competitive outcome in phytoplankton communities, leading to blooms of harmful algae. A better understanding of the growth dynamics of HAB species in relation to nutrient input and coastal eutrophication will require a more detailed analysis of their physiological responses to nitrogen source. The enzyme, nitrate reductase, catalyzes the rate-limiting step in nitrate assimilation. Currently there are no methods available to evaluate species-specific nitrate reductase enzyme activities in mixed communities. Bulk assays that determine changes in nitrate reductase activity for the entire community are of little value due to the diverse responses among different algal groups. Here we describe a novel method to evaluate species-specific changes in nitrate reductase activity. This *in vivo* method employs 6-chloro-9-nitro-5-oxo-5*H*-benzo[*a*]phenoxazine (CNOB) as a fluorescent substrate for nitrate reductase and flow cytometry to evaluate cell specific fluorescence. When excited at 488 nm, the intensity of the emission peak at 583 nm is inversely proportional to the amount of substrate reduced by the enzyme. To validate the proposed method, comparisons to a traditional spectrophotometric nitrate reductase assay were made using a unialgal culture of *Heterosigma akashiwo*. Enzyme activity over time and inhibition studies show similar trends for both methods. With the traditional spectrophotometric activity assay, it was also shown that increasing concentrations of CNOB inhibit the ability of purified nitrate reductase to reduce nitrate to nitrite due to competition for the active site of nitrate reductase. We also confirmed that CNOB is a substrate for purified nitrate reductase using a spectrofluorometer to observe the fluorescence decrease as CNOB is reduced by the enzyme. A similar reduction of CNOB by nitrate reductase in unialgal cultures was also demonstrated. Using a flow cytometer, species-specific enzyme activity within a mixed community was evaluated. The sorting feature of the flow cytometer was used to isolate gated populations of interest in a mixed community based on light-scattering and fluorescence characteristics. DNA extracted from as few as 50 cells collected by the sorting feature was used to identify species within the gated populations by denaturing gradient gel electrophoresis of PCR products. After identification, species-specific reduction of CNOB by nitrate reductase can be calculated from shifts in fluorescence observed in the FL2 channel. In summary, our results suggest that the CNOB method has many advantages over the traditional *in vitro* method for measuring nitrate reductase activity. First, the CNOB method is highly sensitive and requires a minimum amount of culture per analysis. This method also allows *in vivo* investigation of species-specific nitrate reductase activity in mixed populations for the first time compared to the traditional method, which is limited to analyzing only bulk activity. Further molecular identification of populations sorted by flow cytometry enables analysis of mixed communities and environmental samples of unknown composition.

VOLUNTEER-DRIVEN SCIENCE: MAPPING HARMFUL ALGAL BLOOMS IN PUGET SOUND, WASHINGTON

Sheean T. Haley¹, Deana L. Erdner², Jerry A. Borchert³, Bob Lona⁴, and Sonya T. Dyhrman¹

¹Woods Hole Oceanographic Institution, Woods Hole, MA 02543, USA

²University of Texas at Austin, Marine Science Institute, Port Aransas, TX 78373, USA

³Shellfish Program, Washington State Department of Health, Tumwater, WA 98501, USA

⁴Marine Biotoxin Unit, Washington State Department of Health, Tumwater, WA 98501, USA

Puget Sound suffers from annual, costly blooms of *Alexandrium catenella*, which cause paralytic shellfish poisoning (PSP). A primary goal for management in this area is to define PSP dynamics and patterns through increased synoptic sampling and early warning capabilities. The Washington State Department of Health and the Puget Sound Ambient Monitoring Program (PSAMP) uses volunteer samplers to assess the environmental quality of the Sound. Since 1990, volunteer stakeholders and local public health organizations have sampled mussels for PSP analyses every two weeks from 40 sites distributed throughout the Puget Sound. Although PSP monitoring is routine, monitoring for *A. catenella* is not.

As part of an overall goal to interface a proven, high-sensitivity detection method for *A. catenella* into existing PSP monitoring efforts, this network of volunteers have been mobilized for monitoring cell abundances in the Puget Sound. These volunteers were recruited from the Washington State Department of Health, the local tribes and the general community, with the majority working with the State Department of Health in the regular PSP monitoring program. The volunteers were given training on the sampling procedure. In addition to the training, each volunteer was given a sampling pack with all the items necessary to process and archive the samples. Every two weeks, volunteers collected samples for qPCR and for whole cell counts.

The result of this volunteer effort has been tremendous. Almost 500 samples for qPCR and almost 100 samples for whole cell counts were collected by volunteers for the 2006 sampling season. Most of these samples are linked to PSP data also collected by the volunteers. Using a team of volunteers enabled large-scale, repeated, and low-cost qPCR sampling that could not have been achieved otherwise. Moreover, volunteers offer local knowledge, enthusiasm and a sense of ownership and contribution to the area that are a powerful combination in developing a successful monitoring program, reporting on the efficacy of management strategies and raising general awareness of HABs in the environment. In summary, volunteer-driven science partnered with existing monitoring programs enables the collection of a more exhaustive data set needed to support successful management decisions, while also immersing the public in a synergistic view of the scientific process.

DISTRIBUTION OF NUTRIENT DATA IN RELATION TO *Karenia brevis* CELL COUNTS ALONG THE WEST CENTRAL FLORIDA COAST

Emily R. Hall¹, L. Kellie Dixon¹, Merrie B. Neely² and Cynthia A. Heil²

¹Mote Marine Laboratory, Sarasota, FL 34236, USA

²Florida Fish and Wildlife Conservation Commission, Fish and Wildlife Research Institute, St. Petersburg, Florida 33701

Red tides (*Karenia brevis*) are frequent in Gulf of Mexico waters near and around the west central coast of Florida. Routine monitoring of red tide occurrences and nutrient patterns have been ongoing for the past few years (1999 - 2007) and endeavors to capture as many nutrient regimes as possible. With over a thousand sampling periods to date and with red tide present to over three million cells per liter, we have identified ranges of ambient nutrients, both particulate and dissolved and including urea, as a function of counts of *K. brevis*. Data have also been segregated both as to distance from shore and latitudinally to reflect differences in estuarine loadings. Observed concentrations are presented as cumulative distribution functions such that observed concentrations and associated *K. brevis* cell counts can be readily identified and assessed for reasonableness. These can provide useful information for either demonstrating preferred nutrient regimes, draw down of nutrients under bloom progression, the likelihood of terrestrial influences, or setting allowable ranges for ecosystem modeling efforts. Funding support was through the State of Florida and the NOAA EcoHAB:Florida project.

NEW TECHNIQUES FOR NON-LETHAL DNA EXTRACTION FROM, AND PASSIVE INTEGRATED TRANSPONDER (PIT) TAGGING OF, THE SOFT-SHELL CLAM *Mya arenaria*

Scott A. Hamilton and Laurie Connell

University of Maine School of Marine Science, Orono, ME, 04469 USA

Attempts to characterize the population dynamics of the soft-shell clam *Mya arenaria* in relation to paralytic shellfish toxins are complicated by a lack of non-lethal genotyping techniques and reliable tagging methods. An easier and non-lethal technique for clam genotyping is presented here. In addition, I propose a new method for clam tagging, which will increase the consistency of identification when retrieving tagged clams. Both of these techniques will be useful in current and future studies on the effects of red tide toxins on the population genetics of soft-shell clams along the coast of Maine.

A non-lethal method of genotyping, using small amounts of hemolymph, was tested for the first time with *Mya arenaria*. A small syringe was used to extract 200 μ l of hemolymph from the clams' anterior abductor muscle, which was then applied directly in a polymerase chain reaction (PCR) to successfully amplify a 172 bp DNA fragment for sequencing. Afterwards, all tested clams survived. Using this method, clams can easily be genotyped before placement back in natural conditions for observation in relation to red tide toxins.

PIT tags are a useful way to reliably track individual animals in the field. By inserting PIT tags into soft-shell clams between the mantle and shell, the loss of clam identification could be consistently avoided. This project was designed to determine a method in which PIT tags can be non-lethally inserted, and remain in the clam without rejection. Three groups of clams were acclimatized in a natural sea water flow-through tank. One group acted as a control without tags; while the experimental group had tags inserted. A third group received the same treatment as the tagged group without tag insertion. The clams were then monitored for a number of weeks for death and tag rejection.

GROWTH AND TOXICITY OF *Alexandrium* sp. ON THE NORTH SHORE OF LONG ISLAND, NY, USA: DYNAMICS AND INTERACTIONS WITH NUTRIENTS

Theresa Hattenrath, Christopher J. Gobler

School of Marine and Atmospheric Sciences, Stony Brook University, Stony Brook, NY 11794

In the spring of 2006 the first case of saxitoxin poisoning of shellfish occurred on Long Island, NY, USA. A bloom in Northport Bay contained *Alexandrium*-like cells and shellfish within this system contained levels of saxitoxin exceeding 80 μ g STX equiv./100g shellfish tissue causing the New York Department of Environmental Conservation to close the bay to shellfish harvest. Given the recurrent nature of *Alexandrium* blooms, an *Alexandrium* monitoring program was implemented to establish the dynamics and toxicity of *Alexandrium* blooms on Long Island's north shore and to ascertain the role nutrients may play in bloom occurrence. Three estuarine systems were monitored weekly-to-biweekly for densities of *Alexandrium fundyense*, saxitoxin, plankton community biomass and composition, nutrient levels, and standard physical parameters (T, S, light, etc). During May of 2007, an *Alexandrium* bloom associated with elevated levels of saxitoxin in the water column occurred, once again, in Northport Bay. During this bloom, total chlorophyll *a* levels were elevated although concentrations in the >20 μ m size class were only 3 μ g L⁻¹ (23% of the total chl *a*) suggesting this was a fairly low biomass toxicity event. *Alexandrium* cell densities during the bloom exceeded 1,000 cells ml⁻¹. Elevated saxitoxin levels > 1 ng L⁻¹; determined by an ELISA assay) in the water column were present through the bloom, with levels peaking at 31 ng L⁻¹ in late May. During the week following the peak water column saxitoxin concentrations, elevated levels of saxitoxin were also found in shellfish (37 μ g STX equiv./100g shellfish tissue).

To better understand the role of nutrients in bloom occurrence, nutrient amendment experiments were conducted parallel to the field survey. Replicate bottles filled with water from Northport Bay were amended with environmentally realistic levels of phosphate, nitrate, urea and ammonium. Prior to the occurrence of the *Alexandrium* bloom, the addition of phosphate yielded significantly enhanced growth rates of the total phytoplankton community ($p < 0.07$; Tukey) whereas during the late spring, including the period of the *Alexandrium* bloom, the addition of nitrogen, particularly ammonium, resulted in significantly higher growth rates for the total phytoplankton community ($p < 0.05$; Tukey). This suggests that the overall phytoplankton population was phosphorus limited in the early spring and nitrogen limited in the late spring. During the *Alexandrium* bloom, the addition of ammonium resulted in significantly higher particulate saxitoxin concentrations ($p < 0.05$; Tukey). These results suggest that ammonium may promote the formation of toxic *Alexandrium* blooms. By contrast, phosphorus loading caused a significant decrease in saxitoxin concentrations during experiments ($p < 0.1$; Tukey), suggesting phosphorus may influence bloom toxicity in this system. Data on the densities and growth rates of *Alexandrium fundyense* during bloom events as determined by oligonucleotide probes will also be presented.

LONG-TERM, TEMPORAL VARIABILITY IN *Pseudo-nitzschia* POPULATION DYNAMICS AND DOMOIC ACID TOXICITY IN MONTEREY BAY, CA

Kendra Hayashi¹, G. Jason Smith¹, and Nicholas A. Welschmeyer¹
¹Moss Landing Marine Laboratories, Moss Landing, CA 95039, USA

A weekly monitoring program began in May 2003 to assess temporal variability in *Pseudo-nitzschia* population dynamics and domoic acid (DA) toxicity at the Monterey Wharf (MWII; 36.50.993N, 121.49.970W) in Monterey, CA. Water collected from a 5 m vertical net tow (mesh size = 20 μ m) was analyzed for phytoplankton community composition, particulate DA, *in vivo* fluorescence, and photosynthetic pigments. Multiple *Pseudo-nitzschia* blooms occurred in 2003, with *P. australis* representing the dominant species and DA concentrations ranged from 0.002 – 2 nM. Beginning in 2004, blooms were less frequent, generally exhibited reduced *Pseudo-nitzschia* abundance and lower DA toxicity (< 0.1 nM). For the next 3 years, the phytoplankton community was mainly dominated by dinoflagellates as evidenced by microscope counts and pigment analysis. Surprisingly, persistent low levels of particulate DA (~0.005 nM) were detected during this 3 year period even when *Pseudo-nitzschia* were not detected by microscope analysis of the samples. In 2007, diatoms reappeared as the dominate phytoplankton group and the largest and most toxic *Pseudo-nitzschia* blooms seen at the MWII occurred in March. To determine if toxic blooms are characterized by distinct species associations within the *Pseudo-nitzschia* community, bulk nucleic acids archived from this time-series were analyzed using a novel species-specific quantitative PCR (QPCR) assay targeting the ITS domain of rDNA. Results from this analysis will be discussed.

EVALUATING GENETIC DIVERSITY OF *Karenia brevis* BLOOMS IN THE WESTERN GULF OF MEXICO

Darren W. Henrichs¹, Mark A. Renshaw², John R. Gold², Lisa Campbell^{1,3}

¹Department of Biology, Texas A&M University, College Station, TX 77843, USA

²Center for Biosystematics and Biodiversity, Texas A&M University, College Station, TX 77843, USA

³Department of Oceanography, Texas A&M University, College Station, TX 77843, USA

Karenia brevis is a toxic dinoflagellate in the Gulf of Mexico that blooms almost annually in the eastern Gulf but sporadically in the western Gulf. At present, there is little information about the genetic structure of blooms of *K. brevis* and how different blooms are related, if at all. A single cell, DNA-extraction and PCR-amplification protocol has been developed that permits acquisition of genetic data from historical, Lugol's Iodine-preserved field samples. This protocol allows for testing of various spatial and temporal hypotheses and has been applied to field samples taken over a two-month period from a bloom of *K. brevis* near Corpus Christi, Texas during the fall of 2005. Genotypes at six nuclear-encoded microsatellites obtained from >1000 individual cells revealed extensive genetic diversity. One sample, for example, contained 65 unique genotypes (haplotypes) among 68 cells genotyped. This information will provide a better understanding of the genetic structure and dynamics of a bloom over time while also helping to establish connections, if any, between blooms, potentially allowing the source of an individual bloom to be identified.

***Alexandrium catenella* CYSTS AND ENVIRONMENTAL CONDITIONS IN PUGET SOUND, WA: RESULTS OF A CYST SURVEY**

Rita A. Horner¹, Cheryl L. Greengrove², Sian Davies-Vollum², James E. Gawel², James R. Postel¹, Annie M. Cox³, Jeff Hubert², Alexander Abrahamson², Julie Masura², and Bruce W. Frost¹

¹School of Oceanography, University of Washington, Seattle, WA 98195-7940, ²Interdisciplinary Arts and Sciences, University of Washington, Tacoma, WA 98402-3100, ³Graduate School of Oceanography, University of Rhode Island, Kingston, RI 02881

Paralytic shellfish poisoning (PSP), caused by *Alexandrium* spp., has been a problem in western Washington marine waters for hundreds, perhaps thousands, of years based on local Indian culture, early explorers, and adaptive responses by native marine mammals, birds, and fish. There have been no recorded human fatalities in Washington since 1942, but even with a comprehensive state monitoring program, human illnesses continue to occur and shellfish harvest closures are now common in all areas except central and southern Hood Canal. To determine the distribution of *Alexandrium* in Puget Sound, we surveyed sediments for benthic cysts in spring 2005 with a more focused study in Quartermaster Harbor and Dyes Inlet in spring 2006. The purpose of the surveys was to determine where cysts are present in surface sediments now and, for some areas, how long the cysts have been present based on cysts in sediment cores dated using ²¹⁰Pb.

Highest cyst numbers in surface sediments were in Quartermaster Harbor in southern Puget Sound and were present down core in sediment dated to 1955. The second highest cyst numbers were in Sequim Bay in northern Puget Sound and were present to the bottom of a relatively short core dated to 1977, but the first closure there was in 1957. In Penn Cove in northern Puget Sound, cysts were found in core sediment dated to 1957 or at least two decades before a major PSP event in 1978. In Case Inlet in southern Puget Sound, cysts were found in core sediment dated to 1986 or two years before the first harvest closure in that area.

Environmental factors studied when cysts were collected include water properties (temperature, salinity, oxygen, chlorophyll, nutrients), phytoplankton, sediment grain size and organic carbon content (TOC), and metal concentrations. In addition, a 24 hour time series of near bottom current velocity, temperature, salinity, oxygen, chlorophyll and transmissivity and four discrete water samples of suspended sediment and cysts were obtained at key sediment transport locations in Dyes Inlet and Quartermaster Harbor in 2006 to determine if cysts were being transported at maximum ebb and flood tides.

Cysts are transported and deposited as part of the silt-sized fraction of sediment. Thus cyst location may be influenced by sediment transportation and deposition and may also be related to sediment properties such as grain size and TOC content. Grain size of surface and core sediments varied from sand to clay-rich, with coarser sediment generally more common in exposed areas. TOC content averaged 6% by weight with variation between 1-11%. No good correlations have yet been found between either grain size or TOC content with cyst abundance. (See poster by Hubert et al. 2007.)

Sediments in many areas of Puget Sound are contaminated by industrial wastes and runoff leaving non-uniformly distributed metal concentrations that are extremely high in some places. Our results from 2005 show no correlation between total metal concentrations (Cu, Cd, As, and Pb) and cyst abundance. This may be expected from the extremely heterogeneous sampling sites visited that year. However, 2006 results also show poor correlation between metal concentrations and cyst abundance. On the inlet-scale our 2006 results show that metals distributions may be predicted using sediment grain size, suggesting that cyst distribution is dependent at least partially on other factors than sediment transport processes.

Hubert, J., A. Abrahamson, K.S. Davies-Vollum, C.L. Greengrove, and A. Cox. 2007. Do sediment conditions affect the incidence of *Alexandrium catenella* and paralytic shellfish poisoning? A study of sites from Puget Sound. Fourth Symposium on Harmful Algae in the U.S. WHOI, MA.

RELATIONSHIP BETWEEN MICROCYSTIN POTENTIAL AND ENVIRONMENTAL VARIABLES IN LAKES ACROSS NEW YORK STATE

Amber M. Hotto¹, Mike F. Satchwell¹, Gregory L. Boyer¹

¹Department of Chemistry, State University of New York, College of Environmental Science and Forestry, Syracuse, NY 13210, USA

The concentration of microcystin and presence of microcystin synthetase genes were determined for 62 lake, reservoir and river sites across New York State in 2000. Microcystin occurrence was widespread. Cyanobacterial (CYA, 77%) and *Microcystis* sp. (MIC, 40%) DNA was identified statewide using molecular probes. Three genes targeted within the microcystin synthetase (*mcyA*, *mcyB* and *mcyD*) cluster varied in abundance (*mcyA* and *mcyD*, 42% each; *mcyB*, 26%). *Microcystis* sp. and microcystin potential determined by PCR were highly correlated (>80% similarity). The *mcy* genes were generally found at lake sites with a higher trophic state (oligotrophic – 14%; eutrophic and hypereutrophic – 80%). Principal component analysis indicated *mcy* genes were common in smaller lakes (<20 m depth), at a basic pH (8-9.5), at lower latitude and longitude, and in conjunction with increasing microcystin concentration, algal biomass and total phosphorus (TP). Logistic and linear regression revealed that chlorophyll-a was correlated with the appearance of genes encoding cyanobacteria, *Microcystis* and microcystin, whereas TP, lake depth and latitude were associated with chlorophyll-a production. These results confirm that shallow lakes with increased TP are directly linked to lake productivity, and with higher algal biomass there is a greater potential for microcystin gene presence.

DO SEDIMENT CONDITIONS AFFECT THE INCIDENCE OF *Alexandrium catenella* AND PARALYTIC SHELLFISH POISONING? A STUDY OF SITES FROM PUGET SOUND

Jeff Hubert¹, Alex Abrahamson¹, K. Sian Davies-Vollum¹, Cheryl Greengrove¹, and Annie Cox² ¹IAS Program in Environmental Sciences, University of Washington-Tacoma, Tacoma, WA 98406, ²Graduate School of Oceanography, University of Rhode Island, Narragansett RI 02882-1197 USA

Paralytic shellfish poisoning (PSP) is common in the Puget Sound, resulting in restrictions on shellfish harvesting and health concerns for humans. PSP is caused by *Alexandrium catenella*, a toxic dinoflagellate with cyst and motile phases. The cysts reside in bottom sediment, the motile phase is released after excystment and blooms of the motile phase cause outbreaks of PSP. Although such outbreaks are monitored in Puget Sound by the Washington Department of Health, the factors that control their distribution and intensity are poorly understood. Sedimentological, biological and hydrological factors that might affect the incidence of PSP in Puget Sound were studied. The sedimentological component of the work is considered here. Sediment may affect the incidence of PSP in a number of ways. *Alexandrium* cysts may be transported and deposited with sediment and the location of cysts may be related to sediment properties such as grain size and organic content. Considering cyst abundance and distribution in relation to sediment may help predict areas likely to experience future PSP outbreaks. Surface sediment was collected and box cores 10-40 cm in length were extracted from thirty-two sites throughout Puget Sound. Cysts of *Alexandrium catenella* were found at more than half of the sites. Grain size of bottom sediments at sites varied from sand to clay-rich silt, although most sites were predominantly silty. Total organic carbon (TOC) content averaged 6% by weight varying from 1-11%. No good correlations were observed between either grain size or TOC content with cyst abundance. Sequim Bay, in the north of Puget Sound and Quartermaster Harbor (QMH) in the south had the highest cyst abundances and cysts were present to the bottom of the cores from both of these sites. Quartermaster Harbor and Sequim Bay are in areas protected from strong bottom currents and their sediments are predominantly silty sand. Sequim Bay sediment has above average levels of TOC but QMH sediment has average carbon content. Neither site has unusual sediment characteristics that might account for the high levels of *Alexandrium* cysts observed in their sediments. The lack of correlation between sediment properties and *Alexandrium* cyst abundance indicates that sediment probably does not play an important role in determining the intensity and distribution of *Alexandrium* blooms.

THE VALUE OF HAB PREDICTIONS TO THE COMMERCIAL SHELLFISH INDUSTRIES IN THE GULF OF MAINE

Di Jin¹ and Porter Hoagland¹

¹Marine Policy Center, Woods Hole Oceanographic Institution, Woods Hole, MA 02540, USA

During the last several decades, harmful algal bloom (HAB) events have been observed in more locations than ever before throughout the United States. Virtually all coastal regions of the United States are now regarded as potentially subject to a wide variety and increased frequency of HABs, and economic impacts can range into the tens of millions for just one event. From a management perspective, it is crucial to begin developing an understanding of the scale of the economic costs to society of HAB events and to determine the extent to which predictive models might be useful in reducing these impacts. Prediction can be based on process-based or empirical models linking the occurrence of HAB events to observable environmental factors. The scale of economic losses and the value of HAB prediction can tell us something about the appropriate scale of public investments in preventing or mitigating the losses. We develop a well-established general formalism for the problem of HAB prediction, and we apply it to commercial shellfishing and growing industries in the Gulf of Maine. We focus on blooms of algae (*Alexandrium spp.*) that produce paralytic shellfish poison (PSP). Blooms of *Alexandrium* frequently result in the closure of productive shellfish beds along the coasts of Maine and Massachusetts, resulting in significant economic impacts. By developing a model for assessing the value of HAB prediction that can be adapted and applied generally, we provide a basis for investment decisions in scientific research and environmental monitoring to support HAB prediction. Using data from commercial shell fishing and growing industries and economic impact estimates for the 2005 HAB event in the Gulf of Maine, we calculate a value of HAB forecasts for the industries in Maine and Massachusetts. Results of our study suggest that the capitalized value of a HAB prediction and tracking system for the Gulf of Maine can range from one to 11 million dollars, depending on the frequency of HAB events, the accuracy of prediction, and the effectiveness of public and private responses.

CHARACTERIZATION OF NITROGEN UPTAKE BY *Heterosigma akashiwo* GROWN IN TURBIDOSTAT CULTURE UNDER TWO LIGHT INTENSITIESDesmond J. Johns¹ and Patricia Glibert¹

¹University of Maryland Center for Environmental Science, Horn Point Laboratory, 2020 Horns Point Rd, P.O. Box 775, Cambridge, MD 21613, USA. E-mail: djohns@hpl.umces.edu

Raphidophytes were recently identified to contribute significantly to algal blooms and have been implicated in fish kills in the coastal bays of Delaware and Maryland. Ecosystem dynamics facilitating raphidophyte blooms in this system are poorly understood. Nutrient loading, particularly nitrogen in the form of NH_4 and urea, has been increasing for the past decade in these poorly-flushed bays, often reaching levels $>5 \mu\text{M-N}$. To assess the contribution of these forms of nitrogen to raphidophyte growth, specific uptake rates of NH_4 , urea, and NO_3 were determined for the raphidophyte *Heterosigma akashiwo* (CCMP 2393) grown in NO_3 -based media in turbidostat culture at 200 and 100 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$. Growth rates were 1.13 d^{-1} . Rates of uptake were determined using ^{15}N -labeled substrates in concentrations bracketing the range observed in Maryland coastal bays. For cells grown at 200 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$, specific uptake rates were highest for NH_4 , followed by NO_3 , then urea at V_{max} . Rates of V_{max} of NH_4 at 100 and 200 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ were not significantly different and exceeded growth rates by a factor of ~ 5 , suggesting enhanced uptake capability. At 200 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$, the half saturation constant, K_s , was $< 1.0 \mu\text{M-N}$ for urea and NH_4 and was $1.21 \mu\text{M-N}$ for NO_3 . *H. akashiwo* often forms blooms in coastal bays during the late summer after ambient nitrogen concentrations have been depleted. *H. akashiwo* thus displays a higher affinity for NH_4 and urea than NO_3 at ambient concentrations typical of late summer in these bays.

IN SITU NUTRIENT MONITORING: AN EXAMPLE OF RESEARCH, DEVELOPMENT, DEMONSTRATION AND TECHNOLOGY TRANSFER

Vince Kelly¹, Patricia Glibert¹, Louis Codispoti¹, Jeffrey Alexander¹, Bruce Michael², and Cynthia Heil³

¹ University of Maryland Center for Environmental Science, Horn Point Laboratory, PO Box 775, Cambridge, MD 21613, USA

² Maryland Department of Natural Resources, Tidewater Ecosystem Assessment Division, 580 Taylor Ave, D2, Annapolis, MD 21401

³ Florida Fish and Wildlife Conservation Commission, Fish and Wildlife Research Institute, 100 8th Ave. S., St. Petersburg, Florida 33701

Technologies for *in situ* nutrient monitoring have advanced considerably in the past several years. These advancements have included the optimization of chemistries for inorganic nutrients (PO_4^{3-} , NH_4^+ and $\text{NO}_3^- + \text{NO}_2^-$), the development of a method for urea analysis, adaptive sampling for water sampled triggered by pre-set levels of nutrients, and improved data transmission for real time data access. We have been using WS EnviroTech (formerly W.S. Ocean Systems) automated nutrient analyzers in the tributaries and embayments of the Chesapeake and Coastal Bays to obtain time series of PO_4^{3-} , NH_4^+ and $\text{NO}_3^- + \text{NO}_2^-$ and urea during periods when HAB outbreaks routinely occur. Common features in the data included highly varying concentrations on the scale of days and significant increases in PO_4^{3-} , NH_4^+ and $\text{NO}_3^- + \text{NO}_2^-$ following rain events. However, PO_4^{3-} increases were contemporaneous with rain events and ephemeral; NH_4^+ increases were variable and site-specific, but generally contemporaneous with rainfall or delayed for up to a day, while the maximum $\text{NO}_3^- + \text{NO}_2^-$ responses lagged reactive PO_4^{3-} by a period of several days and generally lasted for several days. The increases for all nutrients tended to be many fold, up to an order of magnitude, higher than pre-rain fall concentrations and would generally be missed by traditional sampling. The varying time scale of these fluxes also yielded highly dynamic nitrogen:phosphorus ratios. Algal responses tended to follow the increases in nitrogen underscoring nitrogen limitation in these systems even when ambient concentrations are not depleted. These innovative monitoring technologies are now being adopted for application by the states of Maryland and Florida.

OCEAN OBSERVING SYSTEMS AND PUBLIC HEALTH: THE FLORIDA BEACH CONDITIONS REPORTING SYSTEM TO MINIMIZE EXPOSURE TO *Karenia brevis* AEROSOLS

Barbara Kirkpatrick¹, Robert Currier¹, Kate Nierenberg¹, Andy Reich², Lorraine C Backer³, Lora E Fleming^{4,5}, Richard Stumpf⁶, Gary Kirkpatrick¹

¹ Mote Marine Laboratory, Sarasota, Florida, 34236; ² Florida Department of Health, Tallahassee, Florida, 32399; ³ National Center for Environmental Health, Centers for Disease Control and Prevention, Atlanta, Georgia, 30329; ⁴ NSF NIEHS Oceans and Human Health Center, University of Miami Rosenstiel School of Marine and Atmospheric Sciences, Miami, Florida, 33149; ⁵ University of Miami School of Medicine, Miami, Florida, 33136; ⁶ NOAA National Ocean Service, Silver Spring, MD 20920

With over 50% of the US population living in a coastal county, the impacts the ocean and coastal environment have on people is substantial. These impacts are both positive, such as tourism and recreation opportunities, as well as negative impacts such as exposure to harmful algal blooms (HABs) and pathogens.

Florida's west coast is an example of a coastal community needing public health information from a beach observing system. The west coast experiences annual blooms of the toxic dinoflagellate, *Karenia brevis*. *K. brevis* produces a potent suite of neurotoxins, brevetoxins. Wind and wave action causes the toxins to become part of the marine aerosol. This toxic aerosol causes respiratory irritation as people inhale. Asthmatics who inhale the toxins report increase upper and lower airway symptoms and have measurable changes on pulmonary function. Real time beach reporting for these toxic aerosols may improve asthmatics and local coastal residents' quality of life

An Integrated Ocean Observing System (IOOS) Public Health Pilot Study in HABs has been designed and implemented in Sarasota and Manatee Counties, Florida. This system is based on condition reports from lifeguards at the 8 public beaches. The lifeguards staff these beaches year round, 7 days a week making them ideal sentinels. The lifeguards provide subjective reports of the amount of dead fish on the beach, the level of respiratory irritation, water color, wind direction, surf condition, and the beach warning flag they are currently flying.

A key component in the design of the observing system was to create an easy reporting pathway for the lifeguards to minimize the amount of time away from their primary duties. The system provides a PDA for each of the 8 beaches. The portable unit allows the lifeguards to report from their guard tower. If conditions at the beach change, the lifeguard can input the data easily on site.

The data is transferred via wireless Internet to a website hosted on the Mote Marine Laboratory Sarasota Operations of the Coastal Ocean Observation Laboratories (SO COOL) server. The user can select the beach of interest and a pop up window provides the most recent report for that beach with a date and time stamp.

The system has proven to be robust and well received by the public. The system has reported variability from beach to beach and has provided vital information to users to minimize their exposure to toxic marine aerosols.

A MESOCOSM STUDY EXAMINING THE INFLUENCE OF NUTRIENTS ON *Alexandrium tamarensis/fundyense* TOXIN CONCENTRATION AND COMPOSITION

David M. Kulis¹, Kerry Norton¹, Henry Lind², Linda McCauley¹, Judith Kleindinst¹, Alena Strojsova³ and Donald M. Anderson¹

¹Woods Hole Oceanographic Institution, Woods Hole, MA

²Town of Eastham Department of Natural Resources, Eastham, MA

³University of South Bohemia, Czech Republic

A mesocosm study along the shoreline of Salt Pond (Eastham, MA) was conducted in the spring of 2005 to examine the influence of nutrient composition on toxin composition and content in *Alexandrium tamarensis*. In this study, as well as in one conducted in 2003, natural pond water assemblages containing *A. tamarensis* along with a clonal culture of the same organism were independently incubated under nutrient replete and nutrient limited conditions in 230-liter tanks for a period of about 30 days. Approximately, on a bi-daily basis, samples for chlorophyll content, cell density, cell volume and life cycle status, cellular nutrient status, saxitoxin content and composition, dissolved and particulate nutrients were collected. In the 2003 study, there was a distinct difference in the cellular toxin composition patterns witnessed in the natural versus the cultured populations in the tanks: the cultured cells had significant shifts in their toxin composition during the course of the experiment while the natural populations did not. Both populations exhibited an increase in toxin content under phosphate stress, and a decrease in toxin content under nitrogen stress as compared to the nutrient-replete controls. These increases and decreases in toxin content due to phosphorus and nitrogen limitation respectively appear to be a typical stress-related response within *Alexandrium* saxitoxin-producing species. However, it is not clear why cellular toxin composition changes did, and did not, occur within the different tank populations of *Alexandrium*. Possible hypotheses for this contrast in toxin compositional change include the effect of life cycle transformations such as those found when sexually compatible cells types co-occur (natural assemblages) or perhaps there is a strain or clone-specific response within *Alexandrium*. Data collected during the 2005 study will be compared and contrasted to those collected in 2003 as well as to those from laboratory studies.

SEDIMENTS AND CYSTS ON THE FLOOR OF HARPSWELL SOUND: IS THERE A LOCAL CYST BED FOR THE HIGH TOXICITIES IN LUMBOS HOLE?

Edward Laine¹, Millan AbiNader¹, Alison Chase¹, Roger Flood², Gregory Teegarden³, Gregory Wycka¹, and Collin Roesler⁴

¹Geology Department, Bowdoin College, Brunswick, ME 04011-8468 USA

²SUNY at Stony Brook Marine Science Research Center, Stony Brook, NY 11794

³Marine Sciences Program, Saint Joseph's College of Maine, Standish, ME 04084 USA

⁴Bigelow Laboratory for Ocean Sciences, West Boothbay, Maine 04575 USA

The Maine Department of Marine Resources PSP monitoring program relies on the Lombos Hole location in Harpswell Sound as an indicator site that signals the imminent onset of HABs in the western Gulf of Maine. One model for this behavior is that Harpswell Sound maintains a local cyst bed of *Alexandrium fundyense* which responds to local conditions with intense blooms. A multibeam backscatter map of the deeper portions of Harpswell Sound indicates that the sound is divided into a northern province of high backscatter and a southern province of low backscatter. Grab samples clearly show that the high backscatter province is floored by sandy gravels and gravelly sands and that the southern province is underlain by muds. Counts of viable cysts in surface sediments show that the gravelly sediments have counts 35% of those in the muddy sediments. While sediment cyst densities are generally higher nearer the mouth of Harpswell Sound in the muddy sediments, chlorophyll levels as measured by an oceanographic monitoring buoy are routinely higher nearer the head of the Sound. Reverse estuarine circulation in Harpswell Sound may promote transport of excysted cells towards the head of the Sound (see Teegarden *et al.* this conference). The presence of cysts in sediments does not necessarily indicate deposition from the immediately overlying waters, as cysts formed near the head of the Sound are in all likelihood transported by near bottom currents to the muddy regions with lower near-bottom current velocities. Our data indicate that comprehensive sediment sampling on fine scales can reveal a complex pattern of cyst distribution, which in conjunction with sonar backscatter and grain analysis data can suggest a process of cyst formation, deposition and transport. Comprehensive sampling can also reveal previously unrecognized “hot spots” of potential *Alexandrium* cyst germination, which in combination with characterization of environmental potential for growth and physical circulation data may provide insight into the spatially and temporally variable nature of *Alexandrium* bloom initiation in the western Gulf of Maine.

THE EMERGING RISKS OF CYANOBACTERIA FOR FISH AND WILDLIFE IN FLORIDA

Jan H. Landsberg¹, Jamie Williams¹, Jennifer L. Wolny¹, Leanne J. Flewelling¹, April Granholm¹, Jay P. Abbott¹, Ted Lange², Gina Del Pizzo², Doug Richard², Yasu Kiryu¹, Meghan Shone¹, Loanna Torrance¹, Ryan Pigg¹, Dan Wolf³, Mark Cunningham³, Meredith Grinnell¹ and Shawn Clemons³

¹Florida Fish and Wildlife Conservation Commission (FWC), Fish and Wildlife Research Institute, St. Petersburg, FL 33701, USA

²Florida Fish and Wildlife Conservation Commission (FWC), Fish and Wildlife Research Institute, Eustis, FL 32726, USA

³Florida Fish and Wildlife Conservation Commission (FWC), Fish and Wildlife Research Institute, Gainesville, FL 32653, USA

Florida's Harmful Algal Bloom Task Force (FHABTF) was established in 1997 and mandated by state legislation in 1999. The FHABTF identified cyanobacteria blooms in fresh, brackish and marine waters as emerging harmful algal bloom (HAB) problems in Florida. For cyanobacteria, six priorities were identified: 1) determine the distribution of toxic and nontoxic strains, 2) develop epidemiological studies to determine public health risks, 3) develop economic impact studies to evaluate losses by location or industry, 4) determine the roles of nutrient enrichment and managed freshwater flow in blooms, 5) determine fate and effects of toxins in the food web, and 6) investigate control and mitigation methods. As part of the FWC's mandate to protect fish and wildlife, we have begun to address some of these priority areas. Cyanobacteria (or blue-green algae) are ubiquitous in Florida's freshwater, brackish, and marine habitats. Major bloom forming species include *Lyngbya majuscula*, *L. wollei*, *Cylindrospermopsis raciborskii*, *Microcystis aeruginosa*, *Anabaena circinalis*, *Aphanizomenon flos-aquae*, *Trichodesmium erythraeum*, and *Synechococcus* sp. (Steidinger et al. 1999; Williams et al. 2001). In 1999, a major alligator (*Alligator mississippiensis*) die-off in Lake Griffin was suspected, but unproven, to be associated with *Cylindrospermopsis* blooms (Richey et al. 2001). Saxitoxins have been detected at low concentrations in blue crabs (*Callinectes sapidus*) surveyed from freshwater and low salinity areas with chronic cyanobacterial blooms. A recent *Cylindrospermopsis* bloom and co-occurring mallard duck (*Anas platyrhynchos* and mallard hybrids) die-off has led to an intense investigation for a potential multi-factorial association of etiological factors including cyanotoxins and botulism. An ongoing study funded by CDC/DOH to assess levels of microcystins in four species of freshwater fish from four lakes has confirmed microcystins in the livers of gizzard shad (*Dorosoma cepedianum*) and bluegill (*Lepomis macrochirus*). Surveys for the epiphytic cyanobacterium (family Stigonematales) (Williams et al. 2007) primarily responsible for Avian Vacuolar Myelinopathy have confirmed by PCR several positive substrates for the first time in Florida (J. Williams et al. in prep.). Response efforts and management plans need continuous reappraisal to address the changing scope and impacts associated with cyanoHABs in Florida's waters.

RICHEY, L.J., CARBONNEAU, D. A., SCHOEB, T. R., TAYLOR, S.K., WOODWARD, A.R., AND CLEMONS, R. 2001. Potential toxicity of cyanobacteria to American alligators (*Alligator mississippiensis*). Final report, Florida Fish and Wildlife Conservation Commission, May 2001, 21pp.

STEIDINGER, K.A, LANDSBERG, J.H., TOMAS, C.R., AND BURNS, J.W. 1999. Harmful algal blooms in Florida. Florida Marine Research Institute, Florida Fish and Wildlife Conservation Commission St. Petersburg. 63 pp.

WILLIAMS, C.D., BURNS, J., CHAPMAN, A., FLEWELLING, L., PAWLOWICZ, M., AND CARMICHAEL, W. 2001. Assessment of cyanotoxins in Florida's lakes, reservoirs, and rivers. Final report. St. John's River Water Management District, Palatka, Florida, 97 pp.

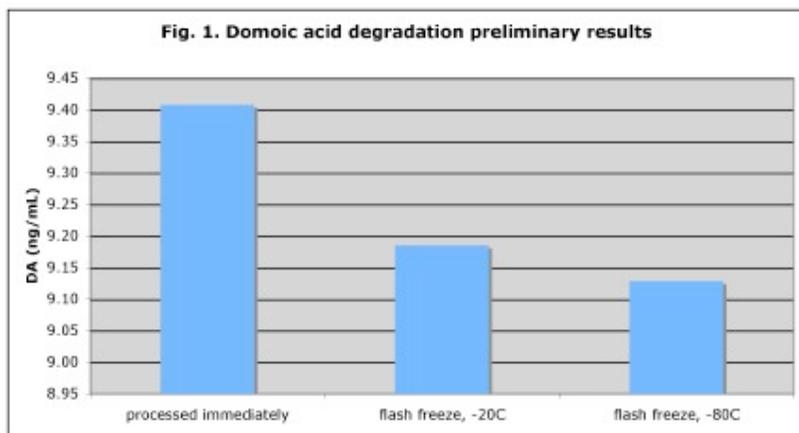
WILLIAMS, S.K., KEMPTON, J., WILDE, S.B., AND LEWITUS, A. 2007. A novel epiphytic cyanobacterium associated with reservoirs affected by avian vacuolar myelinopathy. *Harmful Algae*, 6:343-353.

DEGRADATION OF DOMOIC ACID UNDER COMMON STORAGE CONDITIONS

Jenny Q. Lane¹, Raphael M. Kudela¹

¹University of California Santa Cruz, 1156 High Street, Santa Cruz, CA 95064

Domoic acid (DA), a potent neurotoxin, is produced by various species of the diatom *Pseudo-nitzschia*. The study of *Pseudo-nitzschia* physiology and toxin production in both the field and in the laboratory commonly involves the collection and (preferably) short-term storage of filtered phytoplankton samples. Because it is generally not practical to process individual phytoplankton samples for DA immediately, the effects of storage on the DA content of the samples becomes an important question. The majority of literature addressing domoic acid degradation has focused on shellfish samples^{1,2}. Degradation of DA in filtered phytoplankton samples has been addressed to a lesser extent, predominantly in the context of photodegradation^{3,4}. One exception is a study on DA stability in field samples collected during a *Pseudo-nitzschia* bloom⁵. In this study, particulate DA (DAP) samples were sub-sampled, stored under various conditions (room temperature, -20°C, and 4°C), and measured over the course of 6 months. While this study offered recommendations to improve current DAP sample storage methods, it incited additional questions and concern regarding DA stability in DAP samples. Preliminary work for the study presented here strongly suggests that: 1) there is immediate DA degradation taking place in DAP samples, and 2) the rate of DA degradation varies among storage treatments (Figure 1). We will address some of the still



outstanding questions surrounding DA degradation through a study of DA degradation in DAP samples recovered from a single toxic uniclonal culture of *P. multiseriis*. Recovered DAP samples were stored under a variety of conditions commonly implemented for DAP sample storage (liquid nitrogen [-198°C], -80°C, -20°C, 4°C). To assess relative degradation rates between treatments, filters from each treatment were measured for DA in triplicate roughly every 2 weeks over a 3-month period. More

frequent measurements were taken in the first week of storage, since there is some evidence that the most rapid degradation takes place immediately following initial storage.

References

- 1 E.A. Smith, E.P. Papanagiotou, N.A. Brown, L.A. Stobo, S. Gallacher, and A.M. Shanks. Effect of storage on amnesic shellfish poisoning (ASP) toxins in king scallops (*Pecten maximus*). *Harmful Algae* 5, 9 (2006).
- 2 P. McCarron, Stephen Burrell, and Phillip Hess. Effect of addition of antibiotics and an antioxidant on the stability of tissue reference materials for domoic acid, the amnesic shellfish poison. *Analytical and Bioanalytical Chemistry* 387, 2495 (2007).
- 3 S.S. Bates, C. Léger, M.L. Wells, and K. Hardy, in *Proceedings of the Eighth Canadian Workshop on Harmful Marine Algae*, edited by S.S. Bates (2003), Vol. 2498.
- 4 R.-C. Bouillon, T.L. Knierim, R.J. Kieber, S.A. Skrabal, and J.L.C. Wright. Photodegradation of the algal toxin domoic acid in natural water matrices. *Limnology & Oceanography* 51, 321 (2006).
- 5 K.A. Baugh, Kathi Lefebvre, John C. Wekell, and Vera L. Trainer, presented at the 3rd Symposium on Harmful Algae in the U.S., Asilomar, CA, 2005 (unpublished).

ASSESSMENT OF MICROCYSTIN PRODUCTION IN CYANOBACTERIA USING A NOVEL WHOLE CELL FLUORESCENT IMMUNOLocalIZATION METHOD, FLOW CYTOMETRY AND THE ENZYME-LINKED IMMUNOSORBANT ASSAY (ELISA)

Heidi Langer Atkinson¹, Ellen Braun-Howland^{1,2} and Stephen J. Shost³

¹University at Albany, School of Public Health, Albany, NY 12222

²Wadsworth Center, New York State Department of Health, Albany, NY 12237

³Center for Environmental Health, New York State Department of Health, Troy, NY 12180

Microcystins are hepatotoxic, non-ribosomal peptides produced by several genera of freshwater cyanobacteria. The detection of microcystin-producing strains of cyanobacteria poses a unique challenge to water management officials because of deficiencies in standardized methods aimed at detecting both the toxin and toxin-producing strains. Routine analyses of phytoplankton from surface waters readily discriminate among cyanobacterial genera; however, morphometric analysis is not sufficient to detect toxic strains. The goal of our research is to develop a rapid and cost-effective microscopic assay that isolates toxic strains using whole cell, fluorescence-based, immunolocalization. Fixation conditions, antigen retrieval and antibody sensitivity were tested by comparing the mean fluorescent intensity (MFI) of labeled cells using flow cytometry to relative toxin concentrations estimated using a commercially available ELISA kit for microcystin. The effects of growth conditions and cell cycle on the efficacy of the immunolocalization method were also examined using *Microcystis aeruginosa*. Results indicated that MFI values measured using flow cytometric analysis regularly correlate with toxin concentrations quantified using ELISA. These results point to a direct relationship between the fluorescent intensity of labeling and toxin content within the cell. An optimized immunolabeling protocol will allow for assessment of relative toxin content within individual cyanobacterial cells and in cell populations, thereby offering new insight into the dynamics of toxin production on a per cell basis. Preliminary investigations with environmental samples seeded with toxic cultures suggest the prospective use of whole cell immunolabeling in field samples collected from at-risk recreational waters and potable water supplies.

ADVANCED TREATMENT PROCESSES FOR THE REMOVAL OF CYANOTOXINS FROM LAKE ERIE DRINKING WATERJungju Lee¹ and Harold W. Walker¹¹Department of Civil and Environmental Engineering and Geodetic Science, The Ohio State University, Columbus, OH 43210, USA

The presence of cyanobacteria (blue-green algae) and associated cyanotoxins in surface water is of increasing concern in the United States as well as other parts of the world. In the Great Lakes region, blooms of toxic *Microcystis* have occurred in recent years on Lake Erie, which serves as a source of drinking water for over 11 million people (Ouellette, et al., 2006). Cyanobacteria naturally produce deleterious compounds due to cell lysis, which may cause health problems for animals and humans. Microcystins are the most frequently occurring class of cyanotoxins, of which microcystin-LR is the most toxic and frequently detected congener (Antoniou, et al., 2005). Due to adverse health effects, the World Health Organization (WHO) established a provisional concentration limit of 1 µg/L for microcystin-LR in drinking water and the United States Environmental Protection Agency (USEPA) has placed microcystins on the Drinking Water Contaminant Candidate List. Many approaches such as coagulation, chlorination, activated carbon adsorption, and ozonation have been investigated for the removal of microcystins from drinking water, but typically are not effective to meet the WHO guideline (Lawton and Robertson, 1999). Low-pressure membrane processes are of increasing interest to remove organic contaminants from drinking water. In particular, coupling powdered activated carbon to ultrafiltration (PAC-UF) is an emerging technology for the treatment of organic micropollutants in drinking water. In this study, we investigate the removal of microcystin-LR from drinking water using a membrane filtration only and in combination with powdered activated carbon. Process variables examined included PAC type, PAC dosage, membrane characteristics, microcystin concentrations, operating conditions, and the presence of natural organic matter (NOM). Of five different UF membranes, polysulfone (PS) membranes with highest hydrophobicity most significantly adsorbed microcystin-LR (~91%) presumably through hydrophobic interactions whereas hydrophilic cellulose acetate (CA) membranes did not adsorb the toxin. Membranes with a molecular weight cutoff (MWCO) of less than 5,000 Da rejected microcystin-LR through a size exclusion mechanism. Thin-film (TF) membranes with a MWCO of 1,000 Da, which is close to the molecular size of microcystin-LR, adsorbed 70% and also rejected 70% of the toxin. Initial concentrations of microcystin-LR had a significant effect on the degree of the toxin adsorption. Adsorbed amount of microcystins on the membrane surface linearly increased with increasing initial concentration. Operating conditions such as pressure, permeate flux, and water recovery did not affect the adsorption and rejection performance. Of two different PAC materials, wood-based activated carbon was up to 4-times more effective at removing microcystin-LR than coconut-based carbon due to greater mesopore volume. When PAC was coupled to UF using polyethersulfone (PES) membranes, greater removal of microcystin-LR occurred compared to when CA membranes were used due to sorption of the toxin to the PES membrane surface. The presence of Suwannee River Fulvic Acid (SRFA) reduced microcystin-LR removal by PAC-UF, primarily due to competition between SRFA and microcystin-LR for sites on the PAC surface.

Antoniou, M. G.; de la Cruz, A. A.; Dionysiou, D. D. 2005, Cyanotoxins: New generation of water contaminants. *J. Environ. Eng.* 131 (9), 1239-1243.

Lawton, L. A.; Robertson, P. K. J. 1999, Physico-chemical treatment methods for the removal of microcystins (cyanobacterial hepatotoxins) from potable waters. *Chem. Soc. Rev.* 28, 217-224.

Ouellette, A. J., Handy, S. M., and Wilhelm, S. W. 2006, Toxic *Microcystis* is widespread in Lake Erie: PCR detection of toxin genes and molecular characterization of associated cyanobacterial communities. *Microbial Ecology* 51(2): 154-165.

THE INFLUENCE OF ENVIRONMENTAL CONDITIONS ON THE SEASONAL VARIATION OF *Microcystis aeruginosa* CELL DENSITY AND MICROCYSTINS CONCENTRATION IN SAN FRANCISCO ESTUARY

P. W. Lehman¹, G. Boyer², M. Satchwell² and S. Waller¹

¹Division of Environmental Services, Department of Water Resources, 3251 S Street, Sacramento, CA 95816

²Department of Environment and Forestry, State University of New York, Syracuse, New York

A bloom of the cyanobacteria *Microcystis aeruginosa* was sampled over the summer and fall in order to determine if the spatial and temporal patterns in cell density, chlorophyll *a* concentration, total microcystins concentration and percent microcystins composition varied with environmental conditions in San Francisco Estuary. It was hypothesized that the seasonal variation in *Microcystis* cell density and microcystin concentration was ecologically important because it could influence the transfer of toxic microcystins into the aquatic food web. Sampling of *Microcystis* cell density, chlorophyll *a* concentration, total microcystins concentration and a suite of environmental conditions was conducted biweekly at nine stations throughout the freshwater tidal and brackish water regions of the estuary between July and November 2004. Total microcystins in zooplankton and clam tissue was also sampled in August and October. *Microcystis* cell density, chlorophyll *a* concentration and total microcystins concentration varied by an order of magnitude and peaked during August and/or September among rivers when P^B_m and α^B were high. Canonical correlation analysis identified low streamflow and high water temperature as environmental factors strongly correlated with the seasonal change in *Microcystis* cell density, total microcystins concentration (cell^{-1}) and total microcystins concentration ($\text{chl } a)^{-1}$. Nutrient concentration and nutrient ratios were of secondary importance in the analysis and may be of lesser importance to seasonal variation of the bloom in this nutrient rich estuary. The seasonal variation of *Microcystis* density and biomass and its variation with environmental conditions were potentially important for the structure and function of the estuarine aquatic food web because total microcystins concentration was high at the base of the food web in zooplankton, amphipod, clam and worm tissue during the peak of the bloom.

INFLUENCE OF TEMPERATURE, SALINITY AND NUTRIENT RATIOS ON TOXIN PROFILES OF *Karenia brevis*Danelle K. Lekan¹ and Carmelo R. Tomas¹¹Center for Marine Science, University of North Carolina Wilmington, Wilmington, NC 28409, USA

The toxic dinoflagellate *Karenia brevis* forms extensive, annual blooms in the Gulf of Mexico releasing potent neurotoxins having significant impacts on human health, mortalities of marine mammals, birds and fish. This study focuses on the factors affecting cellular virulence. Effects of environmental factors such as temperature (20-30°C), salinity (20-39) and differing nutrient environments expressed as N:P ratios of 16:1 (balanced), 4:1 (nitrogen limited) and 80:1 (phosphorus limited) were explored with three clones of *Karenia*. The Wilson clone, historically the source of brevetoxin (PbTx) standards for more than 30 years, was used along with the SP3 Super-tox and SP3 Non-tox. Growth as measured by *in vivo* fluorescence was used to assess the effects of temperature and salinity. The SP3 Super-tox and SP3 Non-tox have greater growth rates at a temperature of 20°C and balanced nutrient conditions than at temperatures of 25 or 30°C. In contrast, the Wilson clone has the greatest growth at 25°C and balanced nutrient conditions than at temperatures of 20 or 30°C. Growth rates for the SP3 Super-tox clone at 20°C ranged from 0.14 to 0.33 div/day. While those for the Non-tox clone at 20°C varied from 0.11 to 0.16 div/day. Growth for the Wilson clone at 25°C ranged from 0.26 to 0.36 div/day. Trends for all clones at 30°C were similar to growth at 25°C. At a salinity of 20, none of the clones in any of the temperature/nutrient treatments grew. At a salinity of 25, growth was variable for each clone and temperatures. Good/strong growth occurred at salinities of 30, 35 and 40, with the best growth at salinity of 35. From these results the optimum temperature/salinity environment for growth was determined. Using optimal salinity and temperature conditions, cultures of each clone of *K. brevis* were grown and examined for brevetoxin profiles including PbTx-2, 3, 6, 9 and brevenal, using LC-Mass Spectrometry. The balanced nutrient profile (16:1) contained PbTx-2, 3, 6, 9 and brevenal. The average cellular quota for the *K. brevis* clones was ~15 pg-toxin/cell. A comparison of brevetoxin profiles for SP3 Super-tox and SP3 Non-tox showed both to have significant toxin content, with the same PbTx's present as those for the Wilson clone; however, Non-tox had a notable increase in brevenal. Nutrient stressed experiments (N:P = 4:1 and 80:1) for toxin profiles are presently underway and the effects of the nutrient ratio experiments are yet to be determined. Once completed, these studies will allow a matrix analysis of temperature/salinity and nutrients in determining the cellular production of the various brevetoxins and brevenal. This information will be relevant in evaluating the virulence of strains of *Karenia* and potentially predicting effects of blooms.

DEVELOPMENT AND INITIATION OF HABISS; CDC's MULTI-STATE HAB SURVEILLANCE SYSTEM

Rebecca LePrell, Lorraine Backer, Michael Miller, Sai Gorrepati
National Center for Environmental Health, Centers for Disease Control and Prevention, Atlanta, GA

The extent of human illness caused by environmental exposure to algal toxins in drinking and recreational waters is unknown. There are guidelines from the World Health Organization, Canada, Brazil, and Australia that public health agencies can follow to make decisions about allowing access to drinking water sources and recreational areas with ongoing HABs. However, there are no U.S. federal regulations, and no official guidance from the U.S. Environmental Protection Agency specifying allowable concentrations of HAB-related toxins in the water. In response to the need to support public health decision-making, NCEH has developed a Harmful Algal Bloom-related Illness Surveillance System (HABISS). HABISS is a unique surveillance system that includes the collection of not only human health data, but data from animals made ill by exposure to HABs and environmental data about the HABs themselves. Data collection is organized in modular format that can easily be adapted to state and local needs. State health agencies are particularly interested in using this database to predict future blooms, thus allowing state public health prevention activities to be in place not only in response to reports of human or animal illnesses, but also in advance of anticipated public health problems.

NCEH is currently collecting environmental data from blooms in each state in order to help facilitate future HAB-related illness prevention and control strategies. In 2005, NCEH began beta-testing HABISS in five states supported by the *Pfiesteria* cooperative agreement, including Florida, Maryland, North Carolina, South Carolina, and Virginia. With the help and recommendations by these state partners, NCEH is integrating additional data elements into HABISS, such as mapping, modeling, and real-time notification capabilities. In addition to working with State Health Agencies, we are expanding interaction with other public health partners. For example, we are working with the American Association of Poison Control Centers to develop new case definitions for HAB toxin-related diseases that will be included in both HABISS and the national Poisindex database. These new case definitions should allow us to capture cases in real-time, thereby improving emergency response to human illness.

Future HABISS Activities

HABISS continues to evolve, and several new initiatives are already underway.

- Linking reported weather conditions and meteorological factors to HAB outbreaks
- Collaborating with investigators at the National Oceanic and Atmospheric Administration (NOAA) and National Centers for Environmental Prediction (NCEP) to add a simplified prediction component to HABISS
- Collaborating with the Olympic Harmful Algal Bloom Program (ORHAB) to help build a web-based bulletin for early warning of Washington coast HAB events
- Collecting data on ocean-related diseases in animals and people to ensure that data is collected in a concurrent way that it can be linked to and overlaid with data from other systems
- Providing scientific data from HABISS to aid the discussion of HABs and global climate change (e.g., on the effects of increasing ambient air temperature, changing wind patterns, and variable tidal patterns on HABs and human health)
- Expanding the HABISS network to include international partners and participants

**INTEGRATING NOVEL DATA SOURCES TO IMPROVE THE GULF OF MEXICO
HARMFUL ALGAL BLOOM FORECAST SYSTEM**

Rebecca Love¹, Mary Culver², Gary Kirkpatrick³, Barbara Kirkpatrick³, Bob Currier³, Kate Nierenberg³,
Cory Boyes³, Richard P. Stumpf⁴, Michelle Tomlinson⁴

¹I.M. Systems Group, Charleston, SC 29405, USA

²NOAA Coastal Services Center, Charleston, SC 29405, USA

³Mote Marine Laboratory, Sarasota, FL 34236, USA

⁴NOAA National Centers for Coastal Ocean Science, Silver Spring, MD 20910, USA

Blooms of the dinoflagellate, *Karenia brevis*, are responsible for serious impacts to the ecological, economic, and public health of the Gulf of Mexico. Since 2004, the Harmful Algal Bloom Forecast System has been providing advance warning of *K. brevis* blooms on an operational basis. Traditional techniques such as *in situ* sampling and satellite imagery are used to monitor the spatial extent and movement of the blooms. New methods and technologies are being used to improve the detection and project the impacts of harmful algal blooms (HAB), such as optical data taken from autonomous underwater vehicle (AUV) gliders and moorings and observations of respiratory irritation. AUV gliders are becoming increasingly important observation platforms for investigating HAB initiation sites because of their unique capability for adaptive sampling. The AUVs and several moored sites are equipped with the BreveBuster optical instrument. Developed by Mote Marine Laboratory, the BreveBuster is capable of identifying *K. brevis* blooms by the absorbance signal and provides a similarity index (SI) value that represents the fraction of *K. brevis* biomass in the phytoplankton community. Data from the AUV gliders are sent via satellite to a computer in near real time and are posted to the Internet. Respiratory irritation information collected by volunteers and lifeguards is used to assess health impacts at local beaches. Analysts with the Harmful Algal Bloom Forecast System, a collaborative effort among state and local managers, research scientists, and various offices within the National Oceanic and Atmospheric Administration, use the AUV data to locate subsurface blooms and the respiratory data to communicate potential health impacts to the general public. Open source mapping options such as Google Maps are being considered for displaying these diverse data sets within a system that integrates satellite imagery, field observations, and model output to produce an operational bulletin twice weekly for coastal managers.

PATHOGENICITY, PREY AVAILABILITY AND FUNCTIONAL TYPES IN *Pfiesteria* AND *Pfiesteria*-LIKE SPECIES: THE ROLE OF MICROPREDATORY FEEDING

Vincent J. Lovko, Wolfgang K. Vogelbein

Department of Environmental and Aquatic Animal Health, Virginia Institute of Marine Science, The College of William and Mary, Rt. 1208, Gloucester Point, VA 23062, USA

Heterotrophic dinoflagellates of the genus *Pfiesteria* are purported to produce a potent ichthyotoxin that stuns fish and causes sloughing of epidermal tissues, which the dinoflagellates subsequently consume. A central notion surrounding *Pfiesteria* toxicity is the existence of distinct “functional types” (e.g., Tox-A, Tox-B, Non-Ind) demonstrating varying degrees of toxicity based primarily on culture age and the type of available prey. It is purported that toxic cultures must be maintained on live fish to stimulate and maintain toxicity, which is lost with extended time in the absence of live fish. Our previous work has demonstrated that, in laboratory assays, these dinoflagellates feed directly on fish epidermis by myzocytosis and, given sufficient time and cell density, this results in fish mortality at rates equivalent to those reported for purportedly “toxic” *Pfiesteria* strains. This direct feeding on fish by myzocytosis suggested an alternative mechanism of pathogenicity in *Pseudopfiesteria shumwayae* that accounted for fish mortalities in laboratory assays without involvement of a toxin. The objectives of this study were to determine if maintenance on fish prey increases pathogenicity in a micropredatory *Pfiesteria* culture (relative to maintenance on algal prey) and to determine the mechanisms that account for any perceived increase in pathogenicity.

We conducted a comparative larval fish bioassay using sub-cultures of a pathogenic *P. shumwayae* isolate (CCMP 2089), previously maintained on either juvenile tilapia (*Oreochromis niloticus*) or a cryptophyte algae (*Rhodomonas salinas* CCMP 1319). Fish mortality was significantly more rapid in the fish-exposed culture (100% mortality in fish-exposed vs 50% mortality in algal-exposed at 72 hrs). However, it was also observed that flagellated-cell density increased more rapidly in the fish-exposed culture [1235 cells/ml (S.E. = 83 cells/ml) in fish-exposed vs 665 cells/ml (S.E. = 36 cells/ml) in algal-exposed at 72 hr], possibly accounting for the increased mortality rate. Subsequent studies examined the response of several pathogenic and non-pathogenic strains of *Pfiesteria* and *Pfiesteria*-like dinoflagellates to fish to determine if the observed increase in cell density in the first study was due to prior exposure to fish prey. Algal-fed strains of *P. shumwayae* and *Cryptoperidiniopsis brodyi* (demonstrated to be pathogenic to fish in previous larval fish bioassays) exhibited a significant increase in mean cell volume when exposed to live fish vs the same strains exposed to cryptophytes only. Strains of *P. piscicida* and *Luciella masanensis* (demonstrated non-pathogenic to fish), did not show an increase in mean cell volume when exposed to fish relative to algal-fed cultures of the same strains. Additionally, morphometric studies were conducted on a pathogenic *P. shumwayae* (CCMP 2089) and a non-pathogenic *P. piscicida* (CCMP 2091) given access to fish tissue for a limited period (8 hr). Strain CCMP 2089 showed a large (3x) increase in flagellated cell volume at 12 hr followed by a decrease in cell density and an increase in cyst density (including doublets and tetrads) at 24 hrs, with a decrease in cyst density and a rapid increase in flagellated cell density from 24- 48 hr. In contrast, in strain CCMP 2091, flagellated cell density and volume remained relatively unchanged over the duration of the experiment (96 hrs) and cyst formation (all single cysts) was negligible. Together, these data suggest that pathogenic (micropredatory) strains of *Pfiesteria*-like dinoflagellates feed rapidly and vigorously on fish tissue, becoming greatly enlarged. These cells subsequently encyst and divide (<24 hr), each producing up to 8 daughter cells. Thus, when a pathogenic fish-fed strain is introduced into a comparative bioassay with algal-fed strains, their densities are able to increase more rapidly and fish mortality rates (by micropredation) are greater as a cell density-dependent effect. In contrast, non-pathogenic strains (e.g., CCMP 2091) do not appear to feed vigorously on fish tissue and do not demonstrate the increase in cell volume and reproductive output. These studies were supported by funding from the CDC, ECOHAB and the Virginia Institute of Marine Science.

THE EFFECT OF CARBON DIOXIDE (CO₂) ON GRAZING ACTIVITIES FOR *Karlodinium veneficum*

Alicia Mangum¹, Gulnihal Ozbay¹, Jennifer J. Griffith², Gary H. Wikfors³, and Kathryn J. Coyne²

¹Delaware State University Department of Agriculture and Natural Resources, Dover, DE 19901, USA

²University of Delaware College of Marine and Earth Studies, Lewes, DE 19958, USA

³NOAA Fisheries Service, Milford, CT 06460 USA

In phytoplankton, growth responses are determined by a number of factors (i.e. dissolved gasses, temperature, light intensity, and nutrients), and the effect of individual variables on growth rate can be strongly influenced by interactions with the other factors. As a response to probable environmental changes, this study focuses on the effect of carbon dioxide (CO₂) on growth rate and grazing activity of *Karlodinium veneficum*. *K. veneficum* is a photosynthetic, mixotrophic dinoflagellate that is capable of ingesting prey to fulfill limitations in energy and nutrients. Previous studies showed that for *K. veneficum*, heterotrophic activity is the main mode for obtaining energy and carbon; whereas, phototrophy may be a strategy for survival when environmental conditions for heterotrophic activity are poor. Recent studies of other mixotrophic algal taxa, however, have demonstrated that effects of heterotrophy on growth are diverse. Even though the importance of energy (sunlight) and inorganic nutrients as factors that regulate feeding in mixotrophic flagellates has been recognized, detailed examinations of how other factors influence the physiological state of cells and, therefore, feeding capability are scarce. In particular, the effects of global climate change; including increased atmospheric CO₂ on heterotrophic activities in mixotrophic dinoflagellates has never been examined. Preliminary data indicate that when the prey (*Rhodomonas*) is present, there is a positive correlation with growth rates in high carbon dioxide systems. For this study, *K. veneficum* and *Rhodomonas* were cultured either alone or together in two different environments: one containing ambient CO₂ (375 ppm) and the other an elevated CO₂ level (750 ppm). Cultures were incubated at 26°C, and at 12:12 light:dark cycle to determine the impact of experimentally increased CO₂ conditions on growth and heterotrophic activity. We also investigated photosynthetic activities of *K. veneficum* cells in laboratory culture using the fluorescent probe, Carboxy SNARF-1 and flow-cytometry to monitor internal pH and physiology of this species as dissolved CO₂ levels was varied. This single-cell method for measuring *in situ* physiology may lead to a better understanding of physiological status of an individual species within a community and how anticipated global atmospheric conditions will contribute to overall species abundance and toxin production in dinoflagellates. This study is a work in progress and the results will be discussed.

IMMUNOLOGICAL RESPONSE OF DISTAL LUNG CELL LINES TO BREVETOXINS

Kelli L. Margot^{1,2} and John E. Baatz^{1,2}

¹Marine Biomedicine and Environmental Sciences Center, Medical University of South Carolina, Charleston, SC 29412, USA

²Department of Pediatrics, Medical University of South Carolina, Charleston, SC 29425, USA

Brevetoxins, produced by *Karenia brevis*, are marine algal toxins associated with Florida red tides. Brevetoxin exposure can occur through consumption of contaminated shellfish or toxin inhalation. Various ailments have been documented following brevetoxin inhalation, including lung irritation, cough, wheezing, and congestion. While little is known about how brevetoxin exerts its effects on the lung, previous studies have suggested that brevetoxins may have an impact on the lung immune system. In this study, mouse alveolar epithelial or mouse alveolar macrophage lung cell lines were exposed to 0.5-2 µg/ml brevetoxin-2 and various immunological responses were measured. Western blotting for surfactant protein-A, a protein involved in lung innate immunity, identified that brevetoxin-2 decreases the amount of secreted SP-A. Cytokine antibody arrays primarily indicate a T_H1 response following brevetoxin-2 exposure. Microscopic imaging of macrophages incubated with fluorescently labeled particles indicated that macrophage phagocytosis increases after brevetoxin-2 exposure. These results suggest that brevetoxin-2 alters the immune response in the lung and enhances inflammation. Future work will aim to identify the pathways leading to these altered responses. *This research was supported by a training grant from the National Ocean Service.*

SPATIAL DISTRIBUTION OF PHYTOPLANKTON GROUPS AND TOXIC SPECIES IN A NEARSHORE FRONTAL ZONE SYSTEM IN MONTEREY BAY, CALIFORNIA

Fernanda Mazzillo¹, John Ryan², Mary Silver¹

¹University of California, Santa Cruz, Santa Cruz, CA 95060, USA

²Monterey Bay Aquarium Research Institute, Moss Landing, CA 95039, USA

Frontal systems are well known as concentration sites for phytoplankton biomass. However, detailed understanding of the interactions between nearshore physical processes and phytoplankton community structure is still missing in Monterey Bay, especially with regards to populations of toxic species. We surveyed the spatial distribution of dominant phytoplankton groups and toxic species across a semi-persistent nearshore frontal zone system in northern Monterey Bay, California, where "red tides" are commonly observed using satellite imagery. In this presentation, we report the changes in the dominant phytoplankton from one side of the front to the other, which includes the observation of a toxic *Pseudo-nitzschia australis* bloom not more than a few kilometers away from a *Ceratium divaricatum* "red tide". We also describe the physical properties of the frontal system, the biomass differences, and the conspicuous taxonomic changes that occur across the frontal system. In addition, we also compared biomass and taxonomic changes in the presence and absence of the frontal system. We conclude that the observed variability of the dominant phytoplankton groups is a response to the presence of such feature. This particular frontal system appears to act as a physical barrier, separating two different physical environments with distinct competitive conditions which favor the growth of different phytoplankton species. Such fine-scale hydrographic feature may reflect population variability in toxic species that has not yet received much attention and we discuss significant implications of such feature in this presentation.

A STUDY OF *Pseudo-nitzschia* IN THE GULF OF MAINE: DIVERSITY AND TOXICITY

Linda A. R. McCauley¹, Katie Libera¹, David M. Kulis¹, Deana L. Erdner², Hannah Blossom¹, Deborah Osborn¹, Elizabeth Yranski¹, Lindsay Green¹, Eric Willard¹, and Donald M. Anderson¹

¹Woods Hole Oceanographic Institution, Woods Hole, MA 02543, USA

²University of Texas Marine Science Institute, Port Aransas, TX 78373, USA

In July 2003, more than twenty whales died on Georges Bank while additional whales and forty-two seals were found dead in the near-shore waters of Maine. Domoic acid (DA), produced by pennate marine diatoms belonging to the genus *Pseudo-nitzschia*, was detected in several of the whales, in some cases at high concentrations. Humans, marine mammals, and birds are all susceptible to DA, which causes Amnesic Shellfish Poisoning (ASP). The first cases of ASP were reported in 1987 on Prince Edward Island (PEI). One year after the PEI outbreak, DA was recorded in the southwest Bay of Fundy (BOF). Low levels of DA have subsequently been detected in the BOF nearly every year since 1988. Low levels of DA have also been found in scallops on the southeast shore of Nova Scotia, and in clams, mussels and scallops collected on the Scotian Shelf and Georges Bank (Addison and Stewart, 1989). It is now clear from the whale deaths and toxicity records in eastern Canada that DA is an emerging or cryptic problem that has not been adequately characterized in the Gulf of Maine (GOM), in terms of both human health and ecosystem impacts. While some species of *Pseudo-nitzschia* are capable of producing DA, not all are; therefore, it is important to know what species are found in the GOM and which of them are toxic.

Beginning in 2006, water samples from throughout the GOM region were collected and *Pseudo-nitzschia* cells were isolated into culture. Molecular techniques including sequencing of the D1-D3 domains of the large subunit ribosomal DNA (LSU rDNA) and the ITS1, 5.8S, and ITS2 regions (ITS) were then used to identify these *Pseudo-nitzschia* cultures. The resulting sequences were compared to sequences in NCBI's GenBank. Out of the thirty-nine sequences that resembled *Pseudo-nitzschia*, each most closely matched one of the following five unique species or strains: *P. delicatissima*, *P. pseudodelicatissima*, *P. cf. subpacifica*, *P. pungens*, and *P. fraudulenta*. Further work will be done using scanning electron microscopy to help corroborate the molecular identifications. These sequences were also used to determine that previously designed probes used to identify west coast *Pseudo-nitzschia* strains (Miller and Scholin, 1998) do not match with 100% identity to these east coast strains.

DA toxicity was measured in the GOM *Pseudo-nitzschia* strains using an Enzyme-Linked Immunosorbent Assay (Biosense, Bergen, Norway). Our results show that 15 of these strains are weakly toxic, with a detectable amount of DA ranging from 92-275 pg/ml in an extracted culture containing cells and media, as compared to our positive control, *P. multiseriata*, from Prince Edward Island, Canada, which had a DA level of about 7000 pg/ml. Further toxin analysis using HPLC is in progress to confirm these readings and work is also ongoing to isolate additional strains of *Pseudo-nitzschia* from the GOM.

MILLER, P. E., and C. SCHOLIN. 1998. Identification and enumeration of cultured and wild *Pseudo-nitzschia* (Bacillariophyceae) using species-specific LSU rRNA-targeted fluorescent probes and filter-based whole cell hybridization. *Journal of Phycology* **34**: 271-382.

ADDISON, R.F., and J.E. STEWART. 1989. Domoic acid and the eastern Canadian molluscan shellfish industry. *Aquaculture* **77**: 263-269.

CALIFORNIA PROGRAM FOR REGIONAL ENHANCED MONITORING OF PHYCOTOXINS (Cal-PReEMPT)

Peter E. Miller¹, Gregg W. Langlois², Raphael M. Kudela³, Mary W. Silver³, Shiyamalie Ruberu², Kusum Perera², Jenny Lane³, Carol Boland⁴, Clive Kittredge²

¹Institute of Marine Sciences, University of California, Santa Cruz, CA 95064

²California Department of Public Health, Richmond, CA 94804

³Ocean Sciences Department, University of California, Santa Cruz, CA 95064

⁴Biology Department, California Polytechnic State University, San Luis Obispo, 93407

California's expansive coastline is threatened by blooms of a variety of harmful algal genera, including *Pseudo-nitzschia* and *Alexandrium*, and these pose a threat to public health, fisheries and aquaculture. New methods for species and toxin detection are available, such as the simple-to-use Jellett tests for domoic acid and saxitoxins in phytoplankton and shellfish samples, the receptor binding assay for saxitoxins, and molecular probes for quantitative species detection. Although these technologies are available, a constraint to adoption of them by the California Department of Public Health (CDPH) and other state monitoring programs is the lack of funds for ground-truthing them, a necessary step to evaluate the efficacy of these tools for improving existing monitoring efforts. To bridge the gulf between availability of new tools and integration of those tools into monitoring efforts, NOAA, through its Monitoring and Event Response Program for Harmful Algal Blooms (MERHAB), is providing funding to perform necessary validation and evaluation of whether to incorporate these methods into the CDPH monitoring program and, if so, how best to utilize them. We have established pilot project sites where new technologies are incorporated into an intensive monitoring program. Our approach is to shift much of the monitoring effort to the field, where field technicians pre-screen samples for toxins and toxin-producing species, thus ensuring early warning of impending blooms while reducing un-necessary and expensive lab-based sample testing. With this presentation we provide a summary of our Cal-PReEMPT project, and include results from field testing Jellett kits for DA and saxitoxins in both plankton and shellfish samples, results from analyses using the receptor binding assay for saxitoxins, and results from a comparison of field observations of potentially toxic species using simple field microscopes versus species-specific molecular probes for quantifying *Pseudo-nitzschia australis*, *P. multiseriata*, and *Alexandrium*. We have also been incorporating remote sensing data into the monitoring and decision-making process, and will provide results highlighting the utility and limitations of these methods, focusing on the 2007 domoic acid event in California.

Based on this rigorous intercomparison, we have found that simple field microscopes and phytoplankton net samples provide an effective first line of defense for detecting the onset of HAB events. While using Jellett test kits for detecting phytoplankton and shellfish PSP in the field is easy, there is a learning curve to interpreting the results. Using the kits does not always provide an obvious "yes / no" result, and, at least for PSP toxins, the detection limit of the kits is slightly lower than the regulatory method, giving positive field results for shellfish samples that are non-detectable for these toxins by the standard mouse bioassay. We have begun using a receptor binding assay for quantifying toxin concentrations and we will determine where the Jellett detection limit is for PSP in our samples. Intralaboratory validation of this method will help determine its suitability as a replacement for the MBA in terms of accuracy and precision, sample throughput, and cost. We have also been conducting an intercalibration of ELISA and FMOC-HPLC methods for particulate domoic acid detection; while overall agreement is excellent, there appear to be differential responses to the presence of DA isomers. Both methods work well, with the ELISA kits providing increased sensitivity and speed, but with higher per-sample costs.

BLOOM-FORMING PHYTOPLANKTON IN SAGINAW BAY (LAKE HURON) AND WESTERN LAKE ERIE: ABUNDANCE, DISTRIBUTION, AND CYANOBACTERIAL TOXICITY DURING LATE SUMMER

David F. Millie¹, G. L. Fahnenstiel², J. Dyble³, R. Pigg¹, R. Rediske⁴, D. M. Klarer⁵, R. W. Litaker⁶, P. A. Tester⁶

¹ Florida Institute of Oceanography & Fish & Wildlife Research Institute, St. Petersburg, FL; ² Great Lakes Environmental Research Laboratory-NOAA, Lake Michigan Field Station, Muskegon, MI; ³ Great Lakes Environmental Research Laboratory-NOAA, Ann Arbor, MI; ⁴ Annis Water Resources Institute, Grand Valley State University, Muskegon, MI; ⁵ Old Woman Creek National Estuarine Research Reserve, Ohio Department of Natural Resources, Huron, OH; ⁶ National Ocean Service-NOAA, Beaufort, NC

Phytoplankton blooms occur throughout the nutrient-enriched waters of Saginaw Bay (Lake Huron) and western Lake Erie during periods of warm temperature and water-column stratification. Although Great Lakes assemblages exhibit spatial/temporal variances reflecting equilibriums between growth and loss and mediated by system-specific forcing phenomena, the synergistic interactions and/or feedbacks among environmental conditions, assemblage composition/biomass and cyanobacterial toxicity events only partially are understood. As a component of the NOAA's *Center of Excellence for Great Lakes & Human Health* research initiative (<http://www.glerl.noaa.gov/res/Centers/HumanHealth/>) and to establish a hierarchy of relative importance among environmental conditions influencing Great Lakes bloom-forming phytoplankton, we characterized the synergistic interactions and/or feedbacks among assemblage composition and biomass, toxic phenomena, and system-specific, water-column properties.

Cyanobacteria and diatoms dominated summer phytoplankton assemblages in Saginaw Bay, whereas assemblages were more diverse in western Lake Erie, with diatoms, chlorophytes, cyanobacteria, and cryptophytes dominating assemblages. Although phytoplankton accumulations were spatially and annually episodic throughout both systems, distinct biological 'hot spots' were evident; the greatest accumulations occurred in near-shore water along the southwestern shoreline of Saginaw Bay (near the confluence of the Saginaw River and the Bay) and along western and southern shorelines of Lake Erie (near the confluence of the Maumee River and the Lake and within the lower reaches of Sandusky Bay). Intracellular microcystin concentrations were within the range of concentrations previously reported for these and other Great Lakes' systems, with concentrations (episodically) exceeding the recommended limit for drinking water and (low risk) recreational use.

A suite of variables, indicative of annually-distinct meteorological and hydrological conditions and nutrient-laden inflows, were identified to (collectively) best 'group' sampling stations in a manner consistent with that of chlorophyll *a* concentrations and cyanobacterial biovolumes. However, a great deal of variability between abiotic and biotic patterns remained unexplained and several abiotic variables singularly corresponded with cyanobacterial abundance. Taken together, it appears that multiple environmental conditions (including annual/episodic meteorological patterns, seasonal/intermittent riverine inflows, annual phosphorus loading, etc.) interact with taxon-specific physiological traits to holistically influence late-summer phytoplankton abundance throughout Saginaw Bay and western Lake Erie.

***Alexandrium fundyense* cDNA MICROARRAY: THE HUNT FOR GROWTH-RELATED GENES**

Lilibeth Miranda and Senjie Lin

Department of Marine Sciences, University of Connecticut, 1080 Shennecossett Rd Groton, CT 06340

Understanding of *in situ* cell division rate (CDR) for the PSP-producing dinoflagellate *Alexandrium fundyense* is an important but challenging task. One potential approach is to use cell cycle molecular markers. A correlation between expression of a marker gene and growth rate can be established and *in situ* CDR can be derived from measured gene expression data. This research will screen the *A. fundyense* genome for genes associated with the cell division cycle using cDNA microarray. A full-length cDNA library was created using the 22-bp spliced leader RNA specific to dinoflagellates. Fifteen hundred random inserts from the *A. fundyense* cDNA library were used to construct the microarray. Abundance of transcripts through the diel cycle will be compared and candidate genes showing differential expression throughout the cell cycle will be validated using real-time quantitative polymerase chain reaction (RT-qPCR). Results obtained to date will be presented.

DISTRIBUTION OF KARLOTOXINS AMONG AUSTRALIAN AND NORTH AMERICAN ICHTHYOTOXIC GYMNODINIOID DINOFLAGELLATES (KARENIACEAE)

Ben D Mooney^{1,2}, Gustaaf M Hallegraeff¹, Miguel F de Salas¹ and Allen R Place²

¹School of Plant Science, University of Tasmania, Private Bag 55, Hobart, Tasmania 7001, Australia

²UMBI Center of Marine Biotechnology, 701 E. Pratt St., Baltimore, MD 21202

Gymnodinioid dinoflagellates from the newly formed family Kareniaceae have long been associated with fish kills, however the precise mechanism by many species remains unsolved. Candidate theories include the production of brevetoxin (in *K. brevis* only), reactive oxygen species, lipid phycotoxins (Arzul *et al.* 2000; Mooney *et al.* 2007) and recently characterized karlotoxins. Karlotoxin has been proposed as the primary cause of ichthyotoxicity in the cosmopolitan *Karlodinium veneficum* and strains known to produce karlotoxins have been isolated from the US Atlantic coast, particularly within Chesapeake Bay, Maryland, USA, and the Swan River, Western Australia (Adolf *et al.* 2005; Deeds *et al.* 2006). Toxin analysis of 13 species of Kareniaceae revealed the presence of karlotoxin, KmTx 2, in only a single species (*Karlodinium veneficum*) but with variable activity in strains isolated from the Swan, Huon and Derwent Rivers in Australia. A newly isolated Southern Ocean species, *Karlodinium conicum*, contained a novel non-hemolytic karlotoxin analogue, confirmed by LC-MS and a characteristic UV-spectrum. Isolated from open water in temperatures as low as 5 °C, the presence of widespread Southern Ocean Kareniaceae (*Karlodinium antarcticum*, *K. conicum*, *K. corrugatum* and *Takayama tuberculata*) (de Salas *et al.* 2008) call for a rethinking of the contribution dinoflagellates may make to the chemical and food chain ecology in the Antarctic environment. Species from the closely related genera *Takayama* (*T. helix*, *T. tasmanica*, *T. tuberculata*), *Karenia* (*K. brevis*, *K. mikimotoi*, *K. papilionacea*) and *Karlodinium* (*K. antarcticum*, *K. ballantinum*, *K. corrugatum*, *K. decipiens*) were all consistently negative for karlotoxins. The potentially toxic *Karenia papilionacea* and *Karenia umbella* have been associated with fish kills in Australia and New Zealand however their fish killing mechanism remains to be determined. Proposed toxic mechanisms of *Karenia mikimotoi* includes toxic PUFA and gymnocins but there is still no conclusive evidence on their qualitative contribution to ichthyotoxicity. No universal mechanism for ichthyotoxicity in gymnodinioid dinoflagellates has yet emerged and synergistic interactions between “toxins” (e.g. brevetoxin, karlotoxin, gymnocin), free fatty acids and reactive oxygen species are likely to vary between species, strains and even environmental conditions.

Adolf, J. E., *et al.* (2005). 3rd Symposium on Harmful Algae in the U.S., California, USA.

Arzul, G., *et al.* (2000). Marine lipids: Proceedings of the Symposium held in Brest, 19-20 November 1998. G. Baudimant *et al.* Plouzané, France, IFREMER: 53-62.

de Salas, M. F., *et al.* (2008). Journal of Phycology **44** (in press).

Deeds, J. R., *et al.* (2006). Journal of Aquatic Animal Health **18**(2): 136-148.

Mooney, B. D., *et al.* (2007). Journal of Phycology **43**: 101-111.

IDENTIFICATION OF *Pseudo-nitzschia* AND DOMOIC ACID FROM A NORTH CAROLINA COASTAL BLOOM: LINKAGE BETWEEN VOLUNTEER OBSERVATIONS AND BIOTOXINS RESEARCH

Steve L. Morton¹, Jeff Paternoster¹, Allison Still¹, Katherine Neller², Danielle James², Marcella Turonis³, and Tod Leighfield¹

¹NOAA/NOS, Marine Biotoxins Program, 291 Fort Johnson Road, Charleston, SC 29412

²First Flight high School, 100 Veterans Drive, Kill Devil Hills, NC 27948

³College of the Albemarle, 132 Russel twiford Rd, Manteo, NC 27954

The Southeastern Phytoplankton Monitoring Network (SEPMN) is a community outreach program developed to increase awareness of harmful algae to constituent groups and directly involve volunteers in coastal stewardship by participation in phytoplankton sampling and identification. Currently the program has 80 sites from North Carolina to Texas. The majority of the volunteer groups include teachers and students, however, universities, aquariums, parks and recreational facilities, and environmental and citizen groups also participate. During 2006-2007, approximately 2000 participants were actively involved in SEPMN programs and monitoring activities. Volunteers are instructed on algae identification and sample on a weekly or biweekly basis, reporting their data via a secure web portal to researchers at the Marine Biotoxins Program. Results from volunteer groups enable researchers to identify problem areas to isolate for further study. Since 2001 over 50 blooms have been observed by volunteer groups.

During 2006, a multi-species bloom of *Pseudo-nitzschia* was observed by volunteer monitors students of Kill Devil Hills High School, North Carolina and College of the Albemarle. Preserved and live samples sent to the Marine Biotoxins Program were positively identified using scanning electron microscopy as *Pseudo-nitzschia pungens*, *P. multiseries*, and *P. pseudodelicatissima*. This *Pseudo-nitzschia* bloom peaked at 7,300 cells/L. Analysis of seawater samples detected the toxin, domoic acid using LC-MS/MS techniques at a concentration of 0.6 ng domoic acid/ml of seawater, making this the first report of domoic acid from the Carolina coast of the United States. Oysters collected at the time of this bloom were also positive for domoic acid at a level of 9.6 ng/g. The identification of this multi-species, toxic bloom in North Carolina's waters is an example where a volunteer monitoring program is useful in developing a species list and record of distribution patterns, as well as alerting scientists to the presence of harmful species.

GULF OF MEXICO PHYTOPLANKTON AFFECT FATE OF RED TIDE TOXIN

Tracey L. Myers¹, Emily K. Prince¹, Jerome Naar², and Julia Kubanek^{1,3}

¹ School of Biology, Georgia Institute of Technology, Atlanta, GA 30332, USA

² Center for Marine Science, University of North Carolina at Wilmington, Wilmington, NC 28409, USA

³ School of Chemistry and Biochemistry, Georgia Institute of Technology, Atlanta, GA 30332, USA

The red tide dinoflagellate *Karenia brevis* kills and incapacitates coastal wildlife via the brevetoxins it produces. However, the relationship between *K. brevis* and other Gulf of Mexico phytoplankton species, such as the abundant and widespread diatom *Skeletonema costatum*, is not well studied. We are exploring how chemical cues mediate interactions between *K. brevis* and its phytoplankton competitors, including *S. costatum*. In lab experiments, the addition of live *S. costatum* led to decreased concentrations of brevetoxin B (PbTx-2) associated with *K. brevis* cells. A similar decrease in PbTx-2 concentration occurred when a mixture of brevetoxins (without live *K. brevis* cells) was exposed to *S. costatum*, indicating that *S. costatum* degrades waterborne PbTx-2. LC-MS and ELISA analyses indicated that PbTx-2 was not transformed into other brevetoxins or known brevetoxin metabolites, and instead is decomposed by a previously-unrecognized mechanism. This degradation of PbTx-2 was accomplished by four different *S. costatum* strains from around the world, suggesting that evolutionary experience with *K. brevis* is not a pre-requisite for the ability to degrade PbTx-2. Additionally, phytoplankton-associated bacteria were found to play no role in PbTx-2 degradation, as both axenic and non-axenic *S. costatum* strains degraded PbTx-2. Our results indicate that the metabolic fate of brevetoxins, and therefore the environmental consequences of harmful algal blooms, may depend on which competitors are present during red tides.

THE MITIGATING PROPERTIES OF CYSTEINE ON THE HARMFUL EFFECTS OF RED TIDE

J. Naar¹, L. J. Flewelling², W. M. Abraham³, H. Jacocks¹, A. Lenzi¹, X. Yang¹, A. Bourdelais¹, S. Michelliza¹, C. Tomas¹ and D.G. Baden¹

¹Center for Marine Science, University of North Carolina at Wilmington, Wilmington NC, USA ²Florida Fish and Wildlife Conservation Commission, Fish and Wildlife Research Institute, St Petersburg, FL, USA ³Mount Sinai Medical Center, Miami Beach, Fl, USA

Karenia brevis blooms, also called Florida red tides, occur almost every year in the Gulf of Mexico resulting in severe economic losses¹, deleterious effects on human health² and devastating consequences to wildlife³. The causative agents for these detrimental impacts are the polyether brevetoxins, a family of potent neurotoxins produced by this organism. Brevetoxin exposure results in massive fish kills⁴, marine mammal deaths^{5,6}, and upper and lower airway impairments in humans exposed to contaminated sea-spray^{2,7}. Because shellfish accumulate brevetoxins, to prevent outbreaks of Neurotoxic Shellfish Poisoning, frequent bans on recreational and commercial bivalve harvesting are required, thus also hurting fisheries. Here we show that the adverse effects of *K. brevis* exposure can be drastically reduced by cysteine. This molecule was found to complex almost instantly with some of the toxins (including the most abundant) produced by *K. brevis*, to form very polar non-toxic derivatives. Cysteine treatment of water toxic enough to kill fish within minutes prevented fish mortality. Likewise, in a sheep asthma model, the same cysteine treatment prevented the bronchoconstrictor response that was otherwise observed following exposure to toxic aerosols. Furthermore, pretreatment of the sheep with cysteine partially blocked the effects of the exposure to toxic aerosols and reversed the bronchoconstriction when administered after exposure. Thus cysteine suppressed, prevented and ameliorated airway constriction, a major pulmonary consequence of brevetoxin inhalation^{8,9}. When applied as a preventive treatment, shellfish treated with cysteine accumulated 50% less toxin than untreated shellfish exposed to *K. brevis* under the same conditions. Most remarkably, cysteine treatment during exposure of shellfish to *K. brevis* reduced the toxin accumulation in tissue by 95%. We conclude that cysteine offers new prospects for mitigating the impact of blooms of brevetoxin-producing algae.

REPORTED RESPIRATORY SYMPTOM INTENSITY IN ASTHMATICS DURING EXPOSURE TO AEROSOLIZED FLORIDA RED TIDE TOXINS

Kate Nierenberg,¹ Alexyz Milian,¹ Lora E. Fleming,² Judy A. Bean,³ Adam Wanner,⁴ Andrew Reich,⁵ Lorraine C. Backer,⁶ David Jayroe,¹ and Barbara Kirkpatrick¹

¹Mote Marine Laboratory, 1600 Ken Thompson Parkway, Sarasota, FL, 34236, ²University of Miami School of Medicine and Rosenstiel School of Marine and Atmospheric Sciences, 1801 NW 9th Ave Suite 200 (R-669), Miami, FL 33136, ³Children's Hospital Medical Center and University of Cincinnati, Cincinnati, Ohio, 04524, ⁴University of Miami School of Medicine, Miami, Florida, 33136, ⁵Florida Department of Health, Tallahassee, Florida, 32399, ⁶National Center for Environmental Health, Centers for Disease Control and Prevention, Atlanta, Georgia, 30329

Florida red tides are naturally occurring blooms of the marine dinoflagellate, *Karenia brevis*. *K. brevis* produces natural toxins called brevetoxins. Brevetoxins become part of the marine aerosol as the fragile, unarmored cells are broken up by wave action. Inhalation of the aerosolized toxin results in upper and lower airway irritation. Symptoms of brevetoxin inhalation include: eye, nose, and throat irritation, coughing, wheezing, chest tightness, and shortness of breath. Asthmatics appear to be more sensitive to the effects of inhaled brevetoxin. This study examined data from 97 asthmatics exposed at the beach for one-hour during *K. brevis* blooms, and on separate occasions when no bloom was present. In conjunction with extensive environmental monitoring, participants were evaluated utilizing questionnaires and pulmonary function testing before and after a one-hour beach walk. A modified Likert scale was incorporated into the questionnaire to create respiratory symptom intensity scores for each individual pre- and post-beach walk. Exposure to Florida red tide significantly increased the reported intensity of respiratory symptoms; no significant changes were seen during an unexposed period. This is the first study to examine the intensity of reported respiratory symptoms in asthmatics after a one hour exposure to Florida red tide.

SEASONAL AND INTER-ANNUAL CHANGES IN DINOFLAGELLATES COMMUNITY COMPOSITION IN NEARSHORE ALABAMA WATERS

Lucie Novoveska¹, William L. Smith², Carol P. Dorsey², Charles A. Stapleton¹ and Hugh L. MacIntyre¹
¹Dauphin Island Sea Lab, Dauphin Island, AL 36528, ²Alabama Department of Public Health, Mobile, AL 36608

Alabama coastal waters have experienced significant blooms of potentially-toxic and toxic diatoms, raphidophytes and dinoflagellates. Our goal is to describe the temporal and spatial variability in community composition and to relate it to environmental changes. Here we evaluate the dinoflagellate community composition over a period of 8 years. Dinoflagellate species abundances from 11 sites in the near-shore Gulf of Mexico have been monitored weekly to bi-weekly since 1999. A further 8 sites in Mobile Bay have been monitored quarterly since 1996. Community compositions in both were grouped and averaged by season and location (Gulf of Mexico or Mobile Bay) and analyzed using PRIMER to determine species interactions, trends and patterns. There was a clear shift in dinoflagellate community composition between seasons and between years (e.g. Fig. 1). For instance, *Alexandrium monilatum* was very abundant in the summer but never recorded in the winter. In contrast, *Dinophysis acuminata* was never present in the summer but did appear in moderate numbers in the winter and spring. Populations of several species of *Ceratium* and *Prorocentrum* were very stable, showing consistent abundances in all season and over the 8-year sample period. Of the common species, *Akashiwo sanguinea* was not recorded prior to 2003 and *Gonyaulax digitale* was observed prior to 2002 and again after 2005. There were major blooms ($10^5 - 10^7$ cells/l) of *Karenia brevis* in the coastal Gulf of Mexico in October 2005 and of *Heterocapsa triquetra* and *Prorocentrum minimum*, predominantly in Mobile Bay in February 2006 and 2007. Some of these were associated with fish-kills. Where blooms were found in both areas, cell densities were higher inside Mobile Bay than in the near-shore Gulf. For instance, a bloom of *Prorocentrum minimum* had a maximum density of 690,000 cells /l in February and March of 2007, but reached a peak of 1,400,000,000 cells/l in Mobile Bay during the same time-period.

ACCUMULATION AND DEPURATION OF BREVETOXINS AND MAJOR METABOLITES IN SHELLFISH EXPOSED TO RECURRING *Karenia brevis* BLOOMS

Richard Pierce¹, Michael Henry¹, Patricia Blum¹, Shannon Osborn¹, Robert Dickey², Steven Plakas², Edward Jester², Ann Abraham², Hudson Granade², Kathleen El Said², Leanne Flewelling³, Paula Scott³

¹Mote Marine Laboratory, Sarasota, FL 34236, USA

²U.S. FDA, Gulf Coast Seafood Laboratory, Dauphin Island, AL 36528, USA

³Florida Fish and Wildlife Conservation Commission, Fish and Wildlife Research Institute, St. Petersburg, FL 33701, USA

Brevetoxins are accumulated in filter-feeding molluscan shellfish during exposure to blooms of *Karenia brevis*. Although parent algal toxins are rapidly metabolized, shellfish remain toxic causing neurotoxic shellfish poisoning (NSP). Trophic transfer of brevetoxins also occurs, as evidenced by human intoxication from consumption of carnivorous lightning whelk (*Busycon contrarium*) taken from Sarasota Bay, Florida. This study monitored the composition and concentration of brevetoxins and major metabolites in clams (*Mercenaria mercenaria*), oysters (*Crassostrea virginica*), and lightning whelk (*Busycon contrarium*) collected from a common site in Sarasota Bay exposed to several *K. brevis* red tide events, from 11/1/2003 through 7/28/2006. Water samples were collected from three locations around the shellfish study site to estimate exposure concentrations. Samples of water and shellfish tissue were analyzed by LC-MS for quantitative and qualitative determination of parent brevetoxins and major metabolites. Toxicity of shellfish was assessed by the standard mouse bioassay. Water samples contained primarily PbTx-2, followed by PbTx-3, PbTx-2-carboxylic acid, and PbTx-1. As blooms diminished, PbTx-3 and PbTx-2-carboxylic acid became the most abundant brevetoxins. In clams and oysters, PbTx-2 was not found, and PbTx-3 was observed at relatively low concentrations. Most abundant were PbTx-2-cysteine and cysteine-sulfoxide conjugates (MH⁺: *m/z* 1018 and 1034, respectively), followed by PbTx-1-cysteine and cysteine-sulfoxide conjugates (*m/z* 990 and 1006, respectively). Clams and oysters retained brevetoxin metabolites long after exposure to *K. brevis* blooms. Oysters were toxic by mouse bioassay longer than clams. In whelks, brevetoxins and metabolites were found at very low levels in muscle tissue, however highly concentrated in viscera.

DETERMINING THE ROLE OF *Karenia brevis* BLOOMS IN EMERGENCY ROOM VISITS DUE TO RESPIRATORY AILMENTS IN SARASOTA, FLORIDA

Lara Y. Polansky, Porter Hoagland, Di Jin, and Barbara Kirkpatrick
Marine Policy Center, Woods Hole Oceanographic Institution, Woods Hole, MA 02543, USA

Harmful algal blooms (HABs) resulting from the marine dinoflagellate *Karenia brevis* are an increasing environmental, economic, and health concern. *K. brevis* blooms, which occur annually in the Gulf of Mexico, produce potent brevetoxins responsible for neurotoxic shellfish poisoning (NSP) in humans as well as mass mortalities of marine organisms. Additionally, anecdotal reports, animal experiments, and pilot studies suggest that aerosolized brevetoxins transported by wind are linked to both upper and lower respiratory symptoms, possibly with long-term health implications. Our research aims to determine whether the number of emergency room visits to Sarasota Memorial Hospital due to respiratory ailments changes as a function of the frequency and intensity of *K. brevis* blooms off the west coast of Florida. Our analysis extends the work of Kirkpatrick et al. (2006) regarding the increased rate of healthcare utilization due to aerosolized brevetoxins. In contrast to previous studies, our research will assess the correlation between red tide blooms and respiratory ailments on a finer time scale, as well as over a longer period and will control for environmental factors that function as significant risk factors/triggers of respiratory ailments. These include: pollen and mold counts, influenza outbreaks, ozone and particulate matter concentrations, and forest fire frequency. We will also account for air temperature, relative humidity, wind speed, and wind direction in the *K. brevis* sampling location, as these measures may influence the production and transport of red tide aerosols (Fleming et al. 2005). This work is the first step in a comprehensive project to determine the economic impacts resulting from various intensities of *K. brevis* blooms, which, in turn, will guide management actions to reduce HAB-induced costs-of-illness.

AMMONIUM SURGE UPTAKE AND INHIBITION OF NITRATE UPTAKE BY SMALL AND LARGE CELL-SIZED *Pseudo-nitzschia* SPECIES FROM THE PACIFIC NORTHWESTRegina L. Radan¹, Maureen E. Auro¹, Julian Herndon¹ and William P. Cochlan¹¹Romberg Tiburon Center for Environmental Studies, San Francisco State University, Tiburon, CA, 94920-1205

Despite the importance of nitrogen in maintaining blooms of toxigenic diatoms and their production of the neurotoxic amino acid, domoic acid, the nitrogenous nutrition of *Pseudo-nitzschia* species is still poorly known, and is primarily limited to the larger cell-sized species. Quantification of the physiological capacity for nitrogen uptake by *Pseudo-nitzschia* species under both N-replete and N-deplete conditions is a necessary prerequisite for an understanding of the success of *Pseudo-nitzschia* relative to other phytoplankton in coastal systems, and may help to identify the proximate causes of toxin production by these diatoms. The nitrogen uptake capabilities of three *Pseudo-nitzschia* species, isolated from the Pacific Northwest and maintained in unialgal cultures, are presented here: the transient elevated 'surge' uptake rates of ammonium by *P. cf. delicatissima* and *P. multiseriis*, the ammonium inhibition of nitrate uptake by *P. cf. cuspidata*, and the ammonium and urea inhibition of nitrate uptake by *P. multiseriis*. Elevated ammonium uptake rates were determined in duplicate, N-starved cultures of the two species (cultures depleted of external nitrate for ~ 2 generations) by following the disappearance of ammonium from the medium at varying intervals (5-30 min) over a six-hour period. Both the large and small cell-sized species demonstrated a capacity for transient 'surge' uptake in the first minutes following ammonium enrichment, but *P. cf. cuspidata* exhibited faster initial rates than *P. multiseriis*. However within 0.5-1.0 h, ammonium specific uptake rates decreased and stabilized to values sufficient to support the pre-conditioned growth rates observed for both species prior to N starvation. During multi-day experiments, duplicate nitrogen-replete cultures of *P. cf. cuspidata* demonstrated complete inhibition (suppression) of nitrate uptake by elevated ammonium concentrations. Similarly, duplicate nitrogen-replete cultures of *P. multiseriis* revealed inhibition of nitrate uptake by elevated ammonium and urea concentrations. Only after the ammonium and urea concentrations were decreased by the cells to below the 'threshold' concentration (< ~4-5 μM) did these diatoms begin their utilization of nitrate. These laboratory results will be discussed with respect to the relative surface area per unit cell volume of the large and small cell-sized species, and the possible ecological consequences of simultaneous uptake of nitrate and ammonium at the ambient N concentrations normally found in the coastal waters of the Pacific Northwest.

DIEL SYNCHRONIZATION OF EMBRYONIC DIAPAUSE AIDS PREDICTION OF FETAL TOXICITY OF CALIFORNIA SEA LIONS TO DOMOIC ACID-PRODUCING HARMFUL ALGAL BLOOMS

John S. Ramsdell

Marine Biotoxins Program, NOAA-National Ocean Service, Charleston, SC 29412, USA

California sea lions (CSL) have been a repeated subject of investigation for early life toxicity, which has been documented to occur in mass clusters associated with DDT toxicity in the 1970's and domoic acid toxicity in the last decade. The mass early life mortality events result from the concentrated breeding grounds and a long gestation period of 11 months. Days after conception, development of a CSL embryo is suspended and not resumed until a decreasing photoperiod of 11.48 h/day is reached¹, which occurs approximately 90 days after conception at the major California breeding grounds. The photoperiod trigger reactivates the development of embryos, synchronizing an entire population to proceed with development for the next 242 days until birth. Embryonic diapause is a selectable trait thought to optimize timing for food utilization and male migratory patterns; yet, based on the toxicological perspective presented here, also serves to synchronize developmental toxicity of pulsed environmental events, such as exposure to domoic acid. Research studies in laboratory rodents have identified age-dependent neurotoxic effects during development advancing from neuronal migration disorder to hippocampal-localized seizures to full limbic seizures. Parallel studies have characterized unusual susceptibility of the fetus to domoic acid with retention of toxin and potential dermal re-exposure throughout pregnancy via the amniotic fluid. This presentation will describe comparative allometric projections of rodent neurodevelopment to the CSL to analyze prenatal toxicity and exposure susceptibility of the CSL to domoic acid. This analysis will be applied to forecast fetal toxicity based upon photoperiod-synchronized fetal development of animals originating from the Channel Island rookeries and should prove useful for prediction of fetal outcome after domoic acid producing blooms occurring at different times of the year.

¹Temte JL, Temte J. 1993. Photoperiod defines the phenology of birth in captive California sea lions. *Marine Mammal Science* 9(3):301-308.

ECOLOGY AND BIODIVERSITY OF TOXIC BENTHIC DINOFLAGELLATES AT JOHNSTON ATOLL, PACIFIC OCEAN

Mindy L. Richlen¹ and Phillip S. Lobel¹

¹Department of Biology, Boston University, Boston, MA 02215 USA

A major impediment to understanding the seemingly random occurrence of ciguatera toxicity is uncertainty regarding the field ecology of benthic dinoflagellates that introduce toxins into the coral reef food web. Although broad generalizations have been made, the results of past studies have often yielded contradictory results, particularly between ecological patterns documented in the Pacific versus the Caribbean. This study used standardized methodology to investigate the distribution and abundance of toxin-producing benthic dinoflagellates from the genera *Gambierdiscus*, *Prorocentrum*, *Ostreopsis*, and *Amphidinium* at Johnston Atoll, Pacific Ocean to determine how water flow, depth and habitat type influence patterns of biodiversity. Dinoflagellate abundance was highest at stations located in lagoon/channel habitats and was lowest in reef crest/back reef areas subject to wave activity. In lagoon/channel habitats, *P. mexicanum*, *P. concavum*, and *G. toxicus*/*P.lima* were consistently the dominant species. However, in back reef/reef crest habitats *O. siamensis*, *O. ovata*, *G. toxicus* and *P. emarginatum* were present in highest proportions, suggesting that these species are better able to persist in turbulent habitats. Grazing activity by herbivorous fishes in reef crest areas is highest relative to the lagoon, suggesting that preferential foraging in reef crest/back reef habitats would result in the uptake and accumulation of toxins predominantly produced by these dinoflagellates. This study shows that remarkably similar patterns of abundance, community composition and species associations exist in the Pacific relative to what has been documented in the Caribbean, which demonstrates that the ecology of this community is consistent among geographic regions and greatly contributes to an accurate and coherent characterization of the population dynamics of ciguatera dinoflagellates.

GRAZING, GROWTH, AND BEHAVIORAL REACTIONS OF A CILIATE FED *Alexandrium* SPP: APPARENT LACK OF RESPONSE TO SAXITOXIN

D Schoener¹, GB McManus¹, D Avery¹, and HG Dam¹

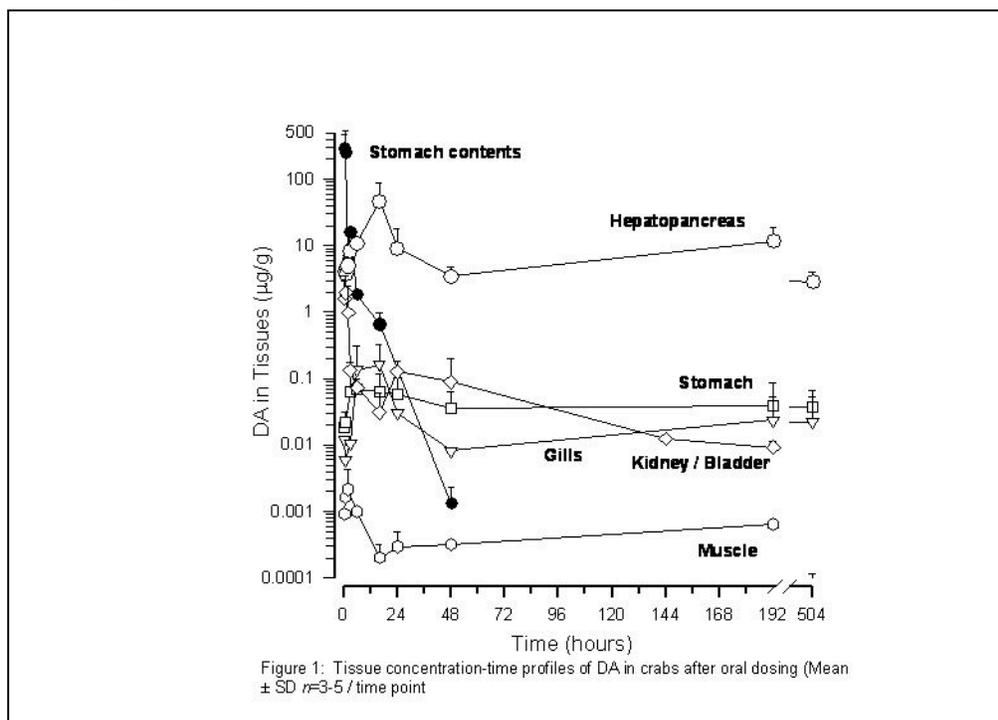
¹University of Connecticut, Marine Sciences, Groton, CT 06340 USA

Harmful algal blooms can cause economic and ecological damage. Ciliates are important grazers of planktonic algae, and may be impacted differently by harmful algal blooms than are other grazers. The bloom-forming dinoflagellate *Alexandrium fundyense* produces saxitoxin (STX) and a number of similar compounds. STX is considered to be a voltage-gated sodium ion channel blocker, but has been shown also to have an effect on calcium channels, which are important in regulation of motility in ciliates. Experiments were carried out with the ciliate *Strombidinopsis* sp. fed two members of the *Alexandrium fundyense* complex, *A. fundyense* and *A. tamarense*. *A. tamarense* is considered to be non-toxic or less toxic than *A. fundyense* because it does not produce STX. While both dinoflagellates make other STX-related compounds, *A. tamarense* produces less of these than *A. fundyense*. The ciliate fed on both species of *Alexandrium*. It survived, but did not grow on *A. fundyense*; however, there was significant mortality in ciliates fed *A. tamarense*. Behavioral assays showed unexpected differences in reactions to *A. fundyense* and *A. tamarense* extracts, with avoidance (backwards swimming) being induced by *A. tamarense* extract but not *A. fundyense*. We are presently developing further the behavior assay and evaluating the effects of calcium channel blockers on motility and grazing in planktonic ciliates. Our goal is to identify the mechanism of toxicity and to assess the role of this interaction in promoting harmful algal blooms.

DOMOIC ACID UPTAKE AND EXCRETION IN FISH, DUNGENESS CRABS, RAZOR CLAMS AND MUSSELS

Irvin R. Schultz¹, Ann Skillman¹ and Dana Woodruff¹. ¹Battelle PND-Marine Science Laboratory, Sequim, WA 98382, USA

Domoic acid (DA) is a neurotoxic amino acid produced by several marine algal species and is the causative agent of amnesic shellfish poisoning in humans, seabirds and marine mammals which have consumed fish that have fed on PN. Profound differences in the toxicokinetics of DA have been identified in a wide variety of shellfish including several species of mussels (*Mytilus* sp.) and razor clams (*Siliqua patula*). We studied the toxicokinetics of DA in razor clams, mussels (*M. galloprovincialis*), Dungeness crabs (*Cancer magister*) and rainbow trout (*Oncorhynchus mykiss*) after intravascular injection and water or oral exposures. Crabs were orally dosed by gavage using homogenized razor clam meat (spiked with DA) and then serially euthanized at selected times and various tissues removed for DA quantification. The uptake of soluble DA in bivalves was studied before and after co-exposure to inhibitors of the multi-xenobiotic resistance (MXR) protein. Oral dosing and initial IV studies used a DA dose of 1 mg/Kg (total weight). Subsequent IV dosing in crabs used a 0.1 mg/Kg DA dose. DA tissue and water concentrations were measured by HPLC-UV / fluorescence or ELISA. After oral dosing in crabs, DA was rapidly and completely absorbed with 2-3 hrs. The majority of the dose was retained in the hepatopancreas, which had DA concentrations 100-200 times that of other tissues such as the kidney and gills. Muscle levels of DA were ~ 10,000 times lower than in the hepatopancreas (Figure 1). After IV dosing in crabs, the initial concentrations of DA in the hemolymph compartment were relatively high and remained largely unchanged for 24 hrs before slowly declining in a log-linear manner. This kinetic behavior was observed for both the 1 and 0.1 mg/Kg doses in crabs. Toxicokinetic analysis using a compartmental model indicated the distributive space of IV injected DA approximated the hemolymph volume of crabs. This is a surprising result given the established ability of the hepatopancreas to extract and retain DA after oral dosing, suggesting a diffusional barrier exists, which prevents DA in the hemolymph from distributing into the hepatopancreas. The kinetic behavior of DA in fish and shellfish will be compared and



contrasted including the potential for the involvement of MXR type proteins in the uptake and tissue distribution of DA.

PHENOTYPIC VARIATIONS IN INGESTION OF TOXIC ALGAE WITHIN POPULATIONS OF A MARINE COPEPOD

Christina Senft^{1,2}, David E. Avery¹, and Hans G. Dam¹

1. Department of Marine Sciences, University of Connecticut, Groton, CT 06340-6098, USA
2. Christina.senft@uconn.edu

Recently, distinct reproductive phenotypes related to PSP resistance have been documented in the copepod *Acartia hudsonica* (Avery and Dam, in press). Furthermore, because a differential population response in ingestion rate of *A. hudsonica* exposed to the toxic dinoflagellate *Alexandrium* spp. has also been documented (Colin and Dam 2002; Colin and Dam in press), we hypothesize that distinct phenotypes for ingestion of toxic algae must exist. Through multiple laboratory studies, this hypothesis was tested. Copepods cultured from regions where toxic *Alexandrium* blooms occur frequently, and others from areas where blooms are infrequent or rare were used in feeding experiments. Ingestion rates of individual copepods were measured when fed either high toxin (*A. fundyense*), or nontoxic (*A. tamarensis*) diets. Frequency distributions of ingestion rates revealed distinctive clustering in animals fed the toxic algae. Normal probability plots and corresponding Shapiro-Wilk tests of normality indicate that the data is non-normal, bolstering the argument that the phenotypic groupings are valid. We propose that these phenotypes represent different levels of resistance to toxic algae. The assays described here could provide a quick test for assessing toxin resistance. In addition, future molecular analysis of preserved experimental copepods should elucidate genotypic variations underlying the phenotypic variations. Coupled with HPLC toxin analysis, these studies will be valuable in assessing toxin dynamics in grazers. The presence of toxin resistant phenotypes in grazer populations has important implications for PSP toxin transfer to higher trophic levels and for bloom control.

References:

- Avery, D.T. and H.G. Dam. In press. Newly discovered reproductive phenotypes of a marine copepod reveal the costs and advantages of resistance to a toxic dinoflagellate. *Limnol. Oceanogr.*
- Colin, S.P. and H.G. Dam. 2002. Latitudinal differentiation in the effects of the toxic dinoflagellate *Alexandrium* spp. on the feeding and reproduction of populations of the copepod *Acartia hudsonica*. *Harmful Algae*. 1: 113-125.
- Colin, S.P. and H.G. Dam. In press. Comparisons of the functional and numerical responses of resistant versus nonresistant populations of the copepod *Acartia hudsonica* fed the toxic dinoflagellate *Alexandrium tamarensis*. *Harmful Algae*.

NORTH CAROLINA HARMFUL ALGAL BLOOM EVENTS 2005 – 2007

Mina Shehee

North Carolina Department of Health and Human Services

Raleigh, North Carolina 27699, USA

The frequency and distribution of harmful (toxin-producing) algal bloom (HAB) events are examined to reveal public health gaps in reducing or preventing human exposure to these toxins. A harmful algal bloom event is defined as a human health complaint or concern originating from a water treatment plant, a state agency, or a citizen. From May 2005 to July 2007, twenty-six events were reported to the North Carolina (NC) Harmful Algal Blooms Program. NC blooms occur in fresh and estuarine waters during late spring to early fall seasons across the western, central, and eastern regions of the state. The majority of these events are related to HABs in drinking water (62%). An algal toxin was present in one-third of the events tested ($n = 21$). Microcystin toxin concentrations ranged from 0.21 to 1.02 $\mu\text{g/L}$ in drinking water and from 0.065 to 0.665 $\mu\text{g/L}$ in recreational (ambient) water. Nearly all of the 2007 events are still pending toxin analyses for anatoxin, cylindropsermopsin, and saxitoxin. The most commonly encountered algae are the cyanobacteria, namely *Anabaena*, *Aphanizomenon*, *Cylindrospermopsis*, *Lyngbya*, and *Microcystis*. A small number of events are due to other algal groups bloom including *Euglena*, *Chattonella*, and *Karlodinium*. Therefore, a NC HAB event profile is characterized as a cyanobacteria species blooming in the warm months, containing a low amount of microcystin, and occurring in any area of the state. The data generated from this investigation are used to establish historical context and drive public health illness surveillance and outreach activities.

MITIGATING THE RISK OF INTRODUCTION OF HARMFUL ALGAE VIA TRANSFER OF BIVALVE MOLLUSCS

Sandra E. Shumway¹, H  l  ne H  garet¹, Gary H. Wikfors²

¹Department of Marine Sciences, University of Connecticut, Groton, CT, USA.

²NOAA-NMFS, Milford, CT, USA.

Bivalve molluscs feed upon microalgae, but may not kill or digest all cells filtered. Shellfish management often involves transplantation, which carries with it the potential for introducing living microalgae in the receiving waters. The risk of introducing some non-native, harmful algal species into new coastal areas within the shells of transplanted molluscan shellfish has been established in the literature, but a wide survey conducted with a consistent protocol has not been done. In the present study, commercially-exploited bivalves and the harmful algal species they can encounter were listed and selected, and laboratory experiments were conducted to assess the risk of introducing the harmful-algal species into new areas when bivalves previously exposed to these algae are transplanted. The selected bivalve species were exposed for two days to an artificial, harmful-algal bloom and then transferred into filtered seawater for several time periods. Fecal samples were collected, observed under the microscope for presence or absence or intact cells, and cultured to assess the potential recovery of viable propagules. Results show that, in most cases, intact, harmful-algal cells were present in the feces. These cells recovered and grew, indicating widespread risk of introducing harmful algae into areas receiving transplanted bivalve molluscs. This risk, however, can be mitigated by 24h depuration in seawater or in air for the species that can survive such emersion (Tables 1A, 1B and 2). When bivalves are shipped, they very often undergo a long time period out of the water; thus, these results indicate that regular practices may mitigate the risk of inadvertent introductions. Simple precautions taken during transplanting and transfer can be effective safeguards.

	<i>Alexandrium fundyense</i>	<i>Alexandrium monilatum</i>	<i>Aureococcus anophagefferens</i>	<i>Heterosigma akashiwo</i>	<i>Karenia mikimotoi</i>	<i>Prorocentrum minimum</i>
<i>Argopecten irradians</i>	+			+		+
<i>Crassostrea virginica</i>	+	+		no feces		+
<i>Mercenaria mercenaria</i>	+	-		+		+
<i>Mya arenaria</i>	-			-		-
<i>Mytilus edulis</i>	+			-	+	+
<i>Perna viridis</i>		+				
<i>Venerupis philippinarum</i>					+	

	<i>Alexandrium fundyense</i>	<i>Alexandrium monilatum</i>	<i>Aureococcus anophagefferens</i>	<i>Heterosigma akashiwo</i>	<i>Karenia mikimotoi</i>	<i>Prorocentrum minimum</i>
<i>Argopecten irradians</i>	-			+		-
<i>Crassostrea virginica</i>	-	+		+		-
<i>Mercenaria mercenaria</i>	-	-		-		-
<i>Mya arenaria</i>	-			-		-
<i>Mytilus edulis</i>	-			-	-	-
<i>Perna viridis</i>		-				
<i>Venerupis philippinarum</i>					-	

Table 1 A and B. Recovery of HAB cells from the biodeposits cultured in the tubes containing FSW after 24 h (1A) and 48 h (1B) depuration in FSW (+ motile cells documented, - no motile or intact cells detected, shaded box, HAB/mollusc pair not tested)

	<i>Heterosigma akashiwo</i>	<i>Prorocentrum minimum</i>
<i>Crassostrea virginica</i>	-	-
<i>Mercenaria mercenaria</i>	-	-
<i>Mytilus edulis</i>	-	-

Table 2. Recovery of HAB cells from the biodeposits cultured in the tubes containing FSW after 24 h out of the water and 24h depuration in FSW (+ motile cells documented, - no motile or intact cells detected, shaded box, HAB/mollusc pair not tested)

DOMOIC ACID IN OCEANIC *Pseudo-nitzschia*: IS IT AN ISSUE?

Mary Silver¹ and Sibel Bargu²

¹Department of Ocean Sciences, University of California at Santa Cruz, 1156 High Street, Santa Cruz, California 95064, USA

²Department of Oceanography and Coastal Sciences, Louisiana State University, 1235 Energy, Coast & Environment Building, Baton Rouge, Louisiana 70803, USA

Pseudo-nitzschia is a nearly ubiquitous genus of pennate diatom that has received greatly increased attention in the last two decades due to the production of the neurotoxin domoic acid (DA) by some of its species. There is now considerable evidence of DA production in many temperate zone coastal environments, sufficient to have led to monitoring programs being instated to protect the public from DA poisoning in commercial and sometimes recreational harvests of shellfish. Interestingly, a considerable amount of the literature on DA coastal “events” is focused on contamination of vertebrate predators such as marine birds and mammals that are often the indicators of toxic blooms in these ecosystems. Thus DA is recognized as a potential toxin capable of being vectored through food chains to result in mortality of animals several trophic levels away from the alga that produced the toxin. The phenomenon of toxicity in the genus has largely been discussed as a nearshore phenomenon.

The goal of this presentation is to present background information to assess the possible production of DA in offshore, oceanic environments and to discuss some concerns about proposed environmental manipulations that may increase *Pseudo-nitzschia* there. We briefly review the state of knowledge about oceanic species of *Pseudo-nitzschia* that may be DA producers *in situ*. We then summarize published results of iron fertilization studies that have found “*Nitzschia*” or “*Pseudo-nitzschia*” to respond to such enrichments. Oceanic iron enrichments are now being widely discussed as a comparatively low cost remedy for carbon dioxide build up in the atmosphere, especially with the possibility of using them to earn “carbon credits” in the global carbon marketplace, as global warming concerns rise. We summarize results from published lab and oceanic experiments that suggest *Pseudo-nitzschia* responses to such iron enrichments and also data that suggest the genus may be particularly suited to respond to such additions. We suggest some possible food web responses that might accompany a response to iron fertilization by blooms of DA-producing *Pseudo-nitzschia*. The second, related talk in this 2-part series will describe some very recent results from a cruise to study *Pseudo-nitzschia* and domoic acid from an oceanic area that contains both naturally iron-enriched regions and ones in iron-limited “high nutrient, low chlorophyll” (HNLC) regions, the type of environment where such iron enrichments are being proposed.

SURVEY OF ALGAL TOXINS IN SOURCE AND FINISHED DRINKING WATERS

James Sinclair¹, Benjamin Southwell² and Judy Westrick²

¹U.S. Environmental Protection Agency, Office of Water, Technical Support Center, Cincinnati, OH 45268

²Lake Superior State University, Sault Ste. Marie, MI 49783

The United States Environmental Protection placed freshwater cyanobacteria, algae and their toxins on its second drinking water Contaminant Candidate List in 2005. Contaminants on this list are considered for possible regulation in drinking water. To make a decision whether or not to regulate them, sufficient information is needed in several areas. These areas include health effects, if they can be controlled by drinking water treatment, and their occurrence in water. More information is needed on the occurrence of algal toxins in water before a regulatory decision can be made. Because there are more than 80 individual toxins or toxin congeners, and not all are equally important in waters of the United States, the USEPA created a short priority list of algal toxins based on the best estimates of their frequency in surface waters of the U.S. and their toxicity. The priority list contains five hepatotoxins, four congeners of microcystin (LR, RR, LA, YR) and cylindrospermopsin, and a neurotoxin, anatoxin-a. This project was a preliminary study of the occurrence of these priority toxins in source and finished drinking waters. Five drinking water utilities, which had a prior history of occurrence of the priority toxins, were sampled. These utilities were in California, Oklahoma, Texas, Vermont, and Florida. Samples were taken from these utilities weekly for 12 weeks between May and August, 2005. Microcystins, cylindrospermopsin, anatoxin-a and cyanobacterial counts for toxin-producers were determined in samples. Microcystin was detected in source water from 4 of the 5 utilities sampled. One of the source water samples had more total microcystins than the 1.0 ppb WHO drinking water guideline value for microcystin LR. Cylindrospermopsin was detected in one source water sample, but anatoxin-a was not detected in any source water sample. None of the toxins were detected in finished water samples. Therefore, at the levels of toxins detected in this study, drinking water treatment appeared to be effective. The detection of toxins in the source waters was not necessarily related to cell density, and highlights the need to analyze water for toxins, rather than rely solely on cell counts as an indication of potential toxin presence.

AXENIC CULTIVATION OF THE HETEROTROPHIC DINOFLAGELLATE *Pfiesteria shumwayae* ON A SEMI-DEFINED MEDIUM

Hayley M. Skelton¹, JoAnn M. Burkholder¹ and Matthew W. Parrow²

¹North Carolina State University, Center for Applied Aquatic Ecology, Raleigh, NC 27606, USA

²University of North Carolina at Charlotte, Department of Biology, Charlotte, NC 28223, USA

The potentially toxic dinoflagellate, *Pfiesteria shumwayae*, is an obligate heterotroph, and its abundance is influenced by the availability of suitable prey. Although *Pfiesteria* spp. feed upon various protists and fish, the nutritional requirements of these dinoflagellates are poorly known. Axenic cultivation, the growth of a species in the absence of other metabolizing cells, allows examination of biochemical requirements without the potentially confounding interactions associated with other living organisms. We developed a semi-defined, biphasic culture medium that has supported the axenic growth of three strains of *P. shumwayae* (up to 1.5×10^5 cells/mL) for 1.5 years, ongoing. The medium contains high concentrations of certain dissolved and particulate organic compounds, including amino acids and lipids. This culture medium will enable further investigations on the nutritional physiology of *P. shumwayae* under controlled conditions. Development of a semi-defined medium represents significant progress toward defining the nutritional requirements of this species, needed to advance understanding about its ecology in eutrophic estuaries.

DEVELOPMENT OF AN INTERNAL STANDARD FOR THE MEASUREMENT OF FREE MICROCYSTINS IN FISH TISSUE AND SEDIMENTS

Juliette L. Smith¹, Kimberly L. Schulz¹, and Gregory L. Boyer²

¹ Environmental and Forest Biology, SUNY-College of Environmental Science and Forestry, Syracuse NY 13210.

² Chemistry, SUNY-College of Environmental Science and Forestry, Syracuse NY 13210.

Microcystins (MCs), a class of potent liver/hepatopancreatic toxins produced by numerous species of freshwater cyanobacteria, are well known for their toxic effects on aquatic organisms. As a result, much effort has been put forth over the last decade towards developing efficient extraction methods, clean-up steps, and quantification methods as a means to evaluate exposure routes and accumulation in aquatic organisms. The recovery efficiency of MCs, however, varies greatly between studies due to the different extraction techniques that are utilized (e.g., methanol vs. EDTA) and the matrix being extracted (e.g., fish tissue vs. sediment). We developed a unique internal standard, with a mass different than any known MCs, which can be spiked into field samples and quantified via LC-MS along with natural MCs already present in the sample. The new compound, a derivative of microcystin-LR, has been modified at the Mdha residue (the site of covalent binding with target molecules), therefore providing an accurate measure of only free MCs, the form thought to be most bioavailable. The internal standard and microcystin-LR chromatograph together and interact with the matrices in a similar manner based on dual spike experiments. Examples of the internal standard being used with both fish tissue and sediment field samples will be presented.

SMALL SCALE BLOOM DYNAMICS OF RAPHIDOPHYTE AND DINOFLAGELLATE POPULATIONS IN KING HARBOR, CALIFORNIA

Beth A. Stauffer¹, Ivona Cetinic¹, Xuemei Bai¹, David A. Caron¹

¹University of Southern California, Los Angeles, CA 90089 USA

King Harbor in the City of Redondo Beach, California was the site of massive fish kills during 2005 following intense and prolonged red tide events. Weekly monitoring since early 2006 revealed the presence of an abundant and diverse community of dinoflagellate and raphidophyte species in the harbor with highly heterogeneous spatial and temporal distributions. Vertical migration and photoacclimation of dinoflagellates and raphidophytes were investigated as mechanisms for dealing with changing light levels in the King Harbor marina over a 24-hour cycle on 19-20 June 2007. PAR, CTD, chlorophyll fluorescence, dissolved oxygen concentrations, active chlorophyll fluorescence, backscattering, and light absorption and attenuation data were measured every four hours using sensor arrays. Discrete water samples were analyzed for pigment concentrations, particulate and dissolved inorganic nutrients, and phytoplankton community composition using both microscopical and molecular techniques. The overall phytoplankton community composition changed significantly during the 24-hour cycle, from a raphidophyte-dominated community (mainly *Heterosigma akashiwo*) to a mixed community of dinoflagellates (*Prorocentrum*, *Ceratium*, and gymnodinoids), raphidophytes and diatoms. A shallow subsurface chlorophyll maximum at 1-2 m was observed during the day, but was located near the bottom (3.5 m) in the evening, with the remainder of the phytoplankton biomass more evenly distributed vertically in the middle of the night. The raphidophyte and dinoflagellate populations tended to constitute the daytime subsurface maxima, whereas the less abundant diatoms were more evenly distributed throughout the water column at all time points. These data suggest a high degree of small-scale heterogeneity in vertical distribution of harmful algal populations and provide important insights into mechanisms that impact community composition within red tide assemblages in King Harbor.

RAPID ENZYME-LINKED IMMUNOSORBENT ASSAY FOR THE DETECTION OF DOMOIC ACID

Thomas N. Stewart¹, R. Wayne Litaker², Bich-Thuy LeEberhart³, John C. Wekell³, Vera L. Trainer³, Raphe Kudela⁴, Rance Hardison² and Pat Tester²

¹Mercury Science Inc., Raleigh, NC 27607, USA

²National Ocean Service, NOAA, Beaufort, North Carolina 28516, USA

³Northwest Fisheries Science Center, NOAA, Seattle, Washington 98112, USA

⁴Ocean Sciences & Institute for Marine Sciences, UCSC, Santa Cruz, CA 95064, USA

Domoic acid (DA) is a potent toxin produced by bloom-forming phytoplankton in the genus *Pseudo-nitzschia* and is responsible for causing amnesic shellfish poisoning (ASP) in humans. ASP symptoms include vomiting, diarrhea, and in more severe cases confusion, memory loss, disorientation and even death. This paper describes the development and validation of a rapid sensitive enzyme linked immunosorbent assay (ELISA) for detecting DA. The assay gives equivalent results to those obtained using standard HPLC and FMOC-HPLC methods. It has a detection range from 0.1 to 3 ppb and can successfully measure DA in razor clams, mussels and phytoplankton. Correlation of ELISA results versus FMOC-HPLC over a broad range of concentrations in razor clam extracts is shown below in Figure 1. Extracts containing high concentrations of DA are quantitated by appropriate dilution. Up to thirty six sample extracts can be analyzed simultaneously in approximately 1.5 hours. The use of a monoclonal antibody and an internal control eliminates the need to run standard curves with each assay. Concentrations of DA are calculated using a predetermined calibration curve specific for this ELISA. The assay uses eight well strips to enable as few as three samples to be analyzed at a time. This eliminates the habit of delaying analysis until a large number of samples are accumulated, thereby saving time and reducing some anomalies that can occur during storage. The ELISA is specific for DA and does not cross-react significantly with glutamate, kainic acid, epi-DA or iso-DA. The relatively low cost, sensitivity, and rapid analysis time provided by this assay make it useful to environmental managers and public health officials for monitoring DA concentrations in environment samples.

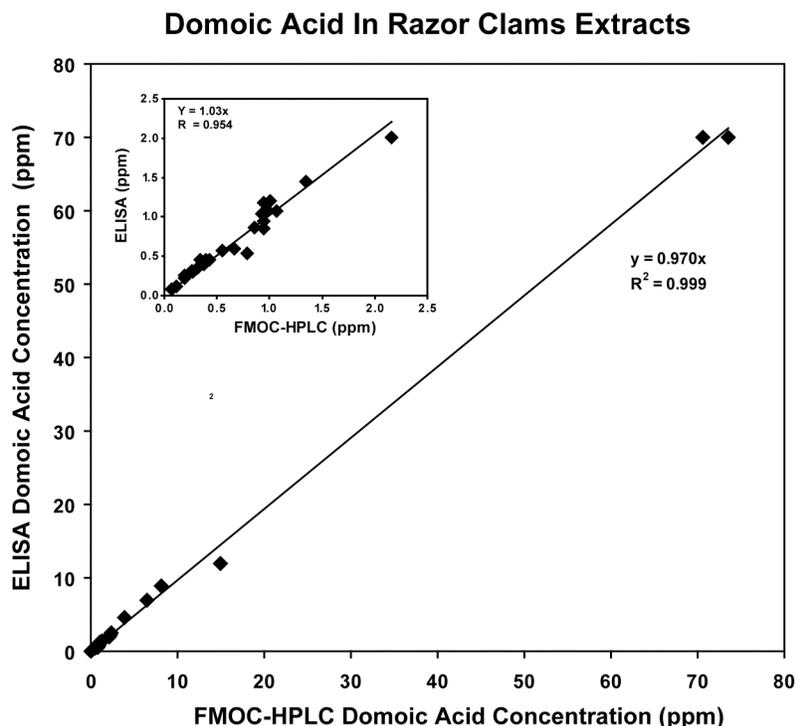


Figure 1. Domoic acid concentrations in razor clam tissues determined from replicate tissue extracts using HPLC and ELISA. The insert shows an expanded version of the regression analysis for sample containing less than 2.5 ppm domoic acid.

NEW SATELLITE DATA PRODUCTS FOR THE DETECTION OF OREGON HABS

Pete Stratton¹, Dave Foley², Michelle Wood³, Jacqui Tweddle¹ and Deb Cannon⁴

¹College of Oceanic and Atmospheric Sciences, Oregon State University

²NOAA CoastWatch, West Coast Regional Node

³Department of Biology, University of Oregon

⁴Oregon Department of Agriculture

With funding from NOAA's Oceans and Human Health Initiative, we have established a collaborative team to understand and better predict the occurrence of HABs in Oregon coastal waters. Our approach is to use satellite and *in situ* data to further our knowledge of the oceanographic conditions that lead to HABs and their interaction with the coast. The principal phytoplankton genera of interest are *Pseudonitzschia* and *Alexandrium* which can cause domoic acid or saxitoxin accumulation (respectively) in coastal shellfish, primarily razor clams. The Oregon Department of Agriculture (ODA) monitors coastal shellfish for phycotoxins, with a view to protecting public health. This database is most comprehensive for both toxins from about 1998, which corresponds to the era of the SeaWiFS ocean color satellite. NOAA CoastWatch has begun routine dissemination of a chlorophyll anomaly product that can be used to document recent increases or decreases in chlorophyll. This can be used to identify the genesis or demise of blooms, but can not yet definitively identify toxic blooms due to the lack of a known optical signal for *Pseudonitzschia* or *Alexandrium*. However, with the inclusion of ancillary data and careful *in situ* monitoring, we believe these new satellite products will enhance coastal management and human health protection. This work evaluates the efficacy of this new product for detecting blooms, and provides examples of combining the anomaly product with physical oceanographic data and coastal wind data to predict the interaction of blooms with coastal shellfish. We present examples of successful and unsuccessful HAB detection and suggest modifications for more effective monitoring and prediction.

***Dinophysis* SPECIES AND DIARRHETIC SHELLFISH TOXINS IN MONTEREY BAY, CA**Cristy Sutherland¹ and Mary Silver¹¹Ocean Science Department, University of California, Santa Cruz, California 95064, USA

In 2000, water samples from Monterey Bay, California that were enriched with species of *Dinophysis* tested positive for toxins responsible for Diarrhetic Shellfish Poisoning (DSP) (unpublished data, R. Weber and T. Yasumoto). No shellfish samples were taken concurrently to show the extent to which the toxins were being accumulated, and shellfish have not been tested for DSP in Monterey Bay or California to date, to our knowledge, other than the testing reported here. *Dinophysis*, including species known to produce DSP elsewhere, however, have been recognized for many decades in this region and, indeed, throughout much of California. Our goal was to determine the annual cycle for the local dominant *Dinophysis* species, *D. acuminata* and *D. fortii*, and to discover whether there is a correlation between their abundance and DSP toxins in CA mussels, *Mytilus californianus*, in the bay. We therefore collected weekly water and mussel samples for 16 months at the Santa Cruz Municipal Wharf in Monterey Bay, and measured *Dinophysis* cell densities in our lab and sent the mussel samples (hepatopancreas) to the Canadian Food Inspection Agency (CFIS) in Dartmouth, Nova Scotia for DSP toxin analysis. For the former we counted cells in 100 ml samples using standard Utermöhl techniques and for the latter, the Canadian Agency used LC-MS to detect the suite of shellfish toxins associated with DSP.

We found a significant association between *Dinophysis* abundance and DSP toxins in mussels. Peak densities of *D. acuminata* and *D. fortii* occurred during the summer months when the majority of DSP toxins were detected in the CA mussel samples. A significant correlation ($p < 0.01$) between *D. fortii* cell numbers and okadaic acid (OA) concentrations in mussels during the 2004-2005 sampling period indicates this species may be the OA source. The correlation coefficient weakened with the addition of *D. acuminata*, suggesting that *D. acuminata* may have little to no role in OA production in Monterey Bay. Since DSP toxins are lipophilic, we correlated toxin levels in mussels not just with the cell densities on the week the mussels were harvested but also with densities averaged over the prior several weeks. Results of these correlations indicate toxins in mussels were mostly strongly related to cell averages obtained over the prior several weeks, rather than on the week of collection. Although none of the CA mussel samples contained toxin levels that exceed the regulatory limit set by Canada, ($1.0 \mu\text{g g}^{-1}$ of digestive gland for any combination of OA/DTX-1), DSP may potentially be a health threat during peak *Dinophysis* events in Monterey Bay, and possibly more broadly in California. In California, recreational harvesting of mussels is prohibited from spring through fall because of the danger of saxitoxin (STX) contamination at this time, which is the anticipated time that DSP toxin levels could be highest in mussels, though commercial harvesting is not similarly prohibited. Instead, for the commercial harvest, mussels are always tested for STX to be sure that the product does not exceed mandated safety limits for STX concentration, a practice that may need to be considered for DSP toxins, given the results presented here.

CHARACTERIZATION, DYNAMICS, AND ECOLOGICAL IMPACTS OF HARMFUL *Cochlodinium polykrikoides* BLOOMS ON EASTERN LONG ISLAND, NY, USA

Ying Zhong Tang¹, Amanda Burson¹, Dianna L. Berry¹, Christopher J. Gobler¹

¹Marine Sciences Research Center, Stony Brook University, Stony Brook, NY 11794-5000, USA

We report on the emergence of *Cochlodinium polykrikoides* blooms in the Peconic Estuary and Shinnecock Estuary, NY, USA, during 2004 - 2006. Blooms occurred during late summer when temperatures and salinities ranged from 20-25°C and 22-30 ppt, respectively. Bloom patches achieved cell densities exceeding 10^5 ml⁻¹ and chlorophyll *a* levels exceeding $100 \mu\text{g L}^{-1}$, while background bloom densities were typically 10^3 - 10^4 cells ml⁻¹. Light, scanning electron and ultrathin-section transmission electron microscopy suggested that cells isolated from blooms displayed characteristics of *C. polykrikoides* and provide the first clear documentation of the fine structure for this species. Sequencing of a hypervariable region of the large subunit rDNA confirmed this finding, displaying 100% similarity to other North American *C. polykrikoides* strains, but a lower similarity to strains from Southeast Asia (88-90%). Bioassay experiments demonstrated that 24 h exposure to bloom waters ($> 5 \times 10^4$ cells ml⁻¹) killed 100% of multiple fish species (1-week old *Cyprinodon variegatus*, adult *Fundulus majalis*, adult *Menidia menidia*) and 80% of adult *Fundulus heteroclitus*. Microscopic evaluation of the gills of moribund fish revealed epithelial proliferation with focal areas of fusion of gill lamellae, suggesting impairment of gill function (e.g. respiration, nitrogen excretion, ion balance). Lower fish mortality was observed at intermediate *C. polykrikoides* densities (10^3 - 10^4 cells ml⁻¹), while all fish survived for 48 hr at cell densities below 1×10^3 cells ml⁻¹. The inability of frozen and thawed-, or filtered ($0.2 \mu\text{m}$)-bloom water to cause fish mortality suggested that the thick polysaccharide layer associated with cell membranes and/or a toxin principle within this layer may be responsible for fish mortality. Juvenile bay scallops (*Argopecten irradians*) and American oysters (*Crassostrea virginica*) experienced elevated mortality compared to control treatments during a nine-day exposure to bloom water ($\sim 5 \times 10^4$ cells ml⁻¹). Surviving scallops exposed to bloom water also experienced significantly reduced growth rates. Moribund shellfish displayed hyperplasia, hemorrhaging, squamation, and apoptosis in gill and digestive tissues with gill inflammation specifically associated with areas containing *C. polykrikoides* cells. In summary, our results indicate *C. polykrikoides* blooms have become annual events on eastern Long Island and that bloom waters are capable of causing rapid mortality in multiple species of finfish and shellfish. Results regarding sequences of large subunit rDNA from other North American *C. polykrikoides* strains and the general characterization of North American *C. polykrikoides* clones compared to Asian clones will also be presented.

THREE YEAR ASSESSMENT OF CYANOHAB FORECASTING ON THE TIDEWATERS OF THE POTOMAC RIVER, CHESAPEAKE BAY, USA

Peter J. Tango¹

¹U.S. Geological Survey, Chesapeake Bay Program Office, 410 Severn Ave, Suite 109, Annapolis, MD 21403, USA.

Cyanobacteria (blue-green algae) blooms have been one of the most frequently occurring toxic plankton blooms that present living resource and human health risks in the tidal Chesapeake Bay. Sufficient resolution of water quality monitoring data was available from 1985-2004 on the Potomac River to explore environmental event patterns and assess predictive power for blooms of toxigenic *Microcystis aeruginosa*. In 2005, the first Harmful Algal Bloom forecast for the Potomac River was generated as one of several ecological forecasting communication tools regarding important Bay phenomena. Using a categorical model, the occurrence of bloom activity was predicted months ahead of the actual events. Levels of success are more variable when predicting important characteristics of the blooms (e.g., time of bloom onset, seasonal bloom duration and amount of area coverage or distance in stream miles for a bloom event) leaving ample opportunity for enhancement of the model and regions of application. Model and monitoring results from 2005-2007 are compared and a lag-time regression model of pre-season flows versus bloom detection rates is evaluated in this presentation.

HEMOLYTIC ACTIVITY OF SELECTED HAB FLAGELLATES: A TOXIN COMPLEX STRATEGY

Avery O. Tatters¹ and Carmelo R. Tomas¹

¹Center for Marine Science, University of North Carolina Wilmington, 5600 Marvin K. Moss Lane, Wilmington, NC 28949.

Selected dinoflagellates consisting of *Amphidinium carterae*, *Prorocentrum minimum*, *P. mexicanum*, *Karenia brevis* and *Gyrodinium instriatum* as well as *Prymnesium parvum* and the raphidophytes, *Heterosigma akashiwo*, *Chattonella subsalsa*, *C. marina*, *C. antiqua* and *Fibrocapsa japonica* cultures were tested with the Erythrocyte Lysis Assay (ELA) for hemolytic activity. Whole cultures, cell pellets and supernatants were tested with osmotically adjusted samples to determine the effect on lysis of human erythrocytes. The highest ELA values consistently were found in the cell pellet followed by the supernatant fractions. *Prymnesium parvum* had the highest activity with log phase cells nearly having 100% lysis of the saponin control and surpassing equivalent samples of other flagellates having similar chlorophyll biomass. Elevated activity of 76-77% of controls was found with *Heterosigma akashiwo*, 36-57% for *Gyrodinium instriatum* and *Prorocentrum mexicanum*, 24-36% for two clones of *Karenia brevis*, 27-32% for *Amphidinium carterae* and 21-27% for *Fibrocapsa japonica*. Modest activity ranging from 9-21% was found for three *Chattonella* species. No activity was detected for *Prorocentrum minimum*. These results suggest that hemolytic activity is more common among HAB species than previously thought and that this aspect of toxicity may play an important role in the concept of “toxin complex or cocktail” acting in synergy with other toxins. Studies involving the evaluation of toxic virulence of HAB species should therefore take this synergy factor into consideration. In addition, single toxin estimations measured on a cellular basis may underestimate the toxic effects of the complex normally found in these cells. The influence of nutrient stress on hemolytic activity is presently being assessed for the different species.

THE IMPORTANCE OF SURFACE CURRENTS IN HIGH PSP TOXICITY IN LUMBOS HOLE, HARPSWELL SOUND, MAINE

Gregory Teegarden¹, Edward Laine,² Scott O'Donnell¹, Millan AbiNader², and Collin Roesler³

¹Marine Sciences Program, Saint Joseph's College of Maine, Standish, ME 04084

²Geology Department, Bowdoin College, Brunswick, ME 04011-8468 USA

³Bigelow Laboratory for Ocean Sciences, West Boothbay, Maine 04575

Over many years, weekly toxicity sampling by the Maine Department of Marine Resources has shown that the Lumbos Hole location in Harpswell Sound is an indicator site that signals through timing and PSP toxicity levels the imminent onset of *Alexandrium fundyense* HABs elsewhere in the western Gulf of Maine. One model for this behavior is that surface currents transport active blooms or viable cysts of *Alexandrium* into Harpswell Sound, or alternatively, that downwelling currents retain developing *Alexandrium* blooms in a favorable environment. In 2007 toxicities in Harpswell Sound began to rise in late April and peaked in late May. Surface currents measured at an oceanographic buoy moored in Harpswell Sound show that the mean residual currents have been almost entirely into Harpswell Sound since mid-February. Preliminary analyses of weekly plankton samples taken in Harpswell Sound indicate that the numbers of *Alexandrium fundyense* began to rise in late April, with significant increases in May. Chlorophyll signals from the oceanographic buoy show seasonal development of the phytoplankton spring bloom, and also higher chlorophyll at the head of the Sound, passing the buoy at ebb tides, and lower levels of chlorophyll from open coastal water moving past in flood tide. During certain tidal phases, surface currents flow towards the head of the Sound continuously, varying in speed but not direction, regardless of flood or ebb cycle. The reverse estuarine flow apparent in Harpswell Sound retains the near-surface populations of *Alexandrium*, which may be derived from local cysts that are spatially variable but of sufficient density to initiate blooms (see Laine *et al.* this conference). Advection of *Alexandrium* cells from outside Harpswell Sound into the Sound, with subsequent retention, is also possible, but not necessary for bloom development in this indicator region. Closures of shellfish beds for PSP contamination indicate toxicity in the Harpswell Sound region, but not in adjacent bays, in late May/early June, suggesting that local conditions govern bloom development and subsequent toxicity. Shellfish toxicity was also apparent to the southwest, near the New Hampshire border, likely derived from offshore *Alexandrium* populations advected southward by the western Maine coastal current (Anderson *et al.* 2005). The distinct spatial character of the two types of closures suggests different mechanisms are probable for bloom development in the Casco Bay region of the western Gulf of Maine. A better understanding of these mechanisms would benefit managers who rely on indicator sites such as Lumbos Hole to inform their monitoring.

References

Anderson, D.M., C.A. Stock, B.A. Keafer, A.B. Neslon, D.J. McGillicuddy, M. Keller, B. Thompson, P.A.

Matrai, J. Martin (2005). *Alexandrium fundyense* cyst dynamics in the Gulf of Maine. Deep Sea Research II 52: 2522-2542.

THE EFFECT OF SALINITY ON DOMOIC ACID PRODUCTION BY THE DIATOM *Pseudo-nitzschia multiseriis*

Anne E. Thessen^{1,2}, K. L. King³, Q. Dortch^{2,4} and G. J. Doucette³

¹Present address: University of Maryland Center for Environmental Science, Horn Point Laboratory, POB 775, Cambridge, MD 21613

²Louisiana Universities Marine Consortium, 8124 Highway 56, Chauvin, LA 70344

³Marine Biotoxins Program, NOAA/National Ocean Service, 219 Fort Johnson Rd. Charleston, SC 29412

⁴Present address: NOAA/COP N/SC12, Rm 8342, 1305 East West Highway, Silver Spring, MD 20910

Domoic acid (DA) is a potent algal neurotoxin produced primarily by members of the diatom genus *Pseudo-nitzschia*, most of which are considered cosmopolites and can produce harmful blooms in estuarine and coastal waters. Many of these habitats are subject to extreme fluctuations in salinity and are utilized extensively as shellfish growing/harvesting areas. Knowledge of how salinity influences DA production is essential to evaluating and ultimately predicting the potential impact of *Pseudo-nitzschia* blooms on shellfish resources as well as various wildlife populations (e.g., marine mammals). Herein, we examine the effect of different salinities (10, 20, 30, 40 psu) on the growth and DA production rates of *P. multiseriis*. Specific growth rates remained maximal ($\sim 0.9 \text{ d}^{-1}$) and essentially unchanged at the three highest salinities tested, but decreased by about half at 10 psu ($\sim 0.4 \text{ d}^{-1}$). By comparison, total (particulate and dissolved) DA quotas ($\sim 30 \text{ fmol DA cell}^{-1}$) and toxin production rates ($\sim 12 \text{ fmol DA cell}^{-1}\text{day}^{-1}$) were similar and maximal at 30 and 40 psu, yet both declined significantly (three- to seven-fold) once adapted to 10 and 20 psu. These results suggest that *P. multiseriis* is able to maintain a high growth rate at 20 psu, but at the expense of continuing to produce DA at elevated levels. Since DA production requires a supply of bioenergetic metabolites generated by photosynthesis, we propose that the additional energy needed to grow rapidly while maintaining an osmotic balance at a likely sub-optimal salinity of 20 psu reduces that available for toxin synthesis, leading to the observed decline in DA levels. Our findings suggest that DA levels should be greatest in higher salinity coastal waters versus estuaries, which is consistent with recent field observations along the Louisiana coast and may help to explain the lack of DA outbreaks in this and other low salinity estuarine-based shellfish harvesting areas.

GERMINATION OF *Alexandrium catenella* CYSTS FROM SURFACE SEDIMENTS IN QUARtermaster HARBOR, WA

Elizabeth D. Tobin, Rita A. Horner, Daniel Grünbaum
School of Oceanography, University of Washington, Seattle, WA. 98195-7940

Paralytic shellfish poisoning (PSP) has a long-term history in the Pacific Northwest of causing regulatory closures of shellfish harvesting. Occurrences of these toxic outbreaks within the Puget Sound have been documented as increasing over the last four decades, however little is known about the biology of the causative organism, *Alexandrium catenella*.

A. catenella has a dual-stage life cycle consisting of motile vegetative cells in the water column and cysts that rest in the sediments. Cyst formation may be an advantageous strategy that promotes survival when conditions in the water no longer support growth. The formation of cysts is thought to be influential in bloom dynamics, but the transitions into and out of the cyst phase are poorly understood. Previous studies have reported a wide range in dormancy period durations from one week to over three months for *A. catenella* from geographically distinct regions (Joyce and Pitcher 2006, Figueroa et al. 2005). This emphasizes the need to determine specific cyst dynamics for geographically distinct *A. catenella* populations.

A survey of *A. catenella* cysts in the Puget Sound, funded by NOAA ECOHAB, has found that of 32 locations sampled, Quartermaster Harbor, in south Puget Sound, has the highest concentrations. Preliminary studies from this project found that *A. catenella* cysts collected from different sites in the Puget Sound (including Quartermaster Harbor) in March and incubated in April and May germinated in approximately 3-6 days. Light was required for germination and the highest excystment occurred around 14°C (Hoffer et al. 2005). A later study from sediment samples collected in January 2006 found no germination to occur in January and February.

In our study, *A. catenella* cysts collected from surface sediments in Quartermaster Harbor in October of 2006 and maintained in the dark at 4°C are being used in a time-series germination experiment to evaluate the duration of the dormancy period of Puget Sound *A. catenella* populations. Well plates containing sediments and f/2 medium have been set up new each month for one year from this sediment sample. The well plates are incubated at 13°C under a 12:12 L:D period and monitored daily for the first week of incubation, and once a week there after for the appearance of motile cells. Preliminary observations have revealed that germination occurs in all months from November through June. Motile cells are not observed on set-up, but germination can occur within 24 hours of being restored to conditions that support growth. These findings differ from the earlier observations of Hoffer et al. (2005). This suggests that cyst germination in the Puget Sound population may occur more rapidly than previously thought. In the next phase of this study, total cyst abundance in sediment sub-samples will be determined using epifluorescence microscopy after sonicating and staining with Primuline (Yamaguchi et al. 1995). Cyst counts will be compared to the number of motile cells observed in fixed well samples to evaluate germination potential.

Figueroa, R.I., I. Bravo, and E. Garcés. 2005. Effects of nutritional factors and different parental crosses on the encystment and excystment of *Alexandrium catenella* (Dinophyceae) in culture. *Phycologia* 44:658-670.

Hoffer, S., R.A. Horner, and C.L. Greengrove. 2005. Germination experiments with *Alexandrium catenella* cysts collected from surface sediments in Puget Sound. Abstract, Third Symposium on Harmful Algae in the U.S. Pacific Grove, CA.

- Joyce, L.B. and G.C. Pitcher. 2006. Cysts of *Alexandrium catenella* on the west coast of South Africa: distribution and characteristics of germination. *African Journal of Marine Science*. 28:295-298.
- Yamaguchi et al. 1995. A rapid and precise technique for the enumeration of resting cysts of *Alexandrium* spp. (Dinophyceae) in natural sediments. *Phycologia* 34:207-214.

THE DEVELOPMENT OF A BEACH IMPACT MODEL FOR FLORIDA *Karenia brevis* BLOOMS

Michelle C. Tomlinson¹, Richard P. Stumpf¹, Julie A. Calkins¹, Robert Currier², Kate Nierenberg², Barbara Kirkpatrick² and Lorraine C. Backer³

¹National Ocean Service, Silver Spring, MD 20910, USA

²Mote Marine Laboratory, Sarasota, FL 34236, USA

³Center for Disease Control and Prevention, Atlanta, GA 30333, USA

Since October 2004, the National Oceanic and Atmospheric Administration has provided operational forecasts of beach impacts to the state of Florida in response to recurrent *Karenia brevis* blooms. *Karenia brevis* produces brevetoxin, an aerosolized toxin which causes respiratory irritation in humans. Forecasts provide information as to the likelihood of beach impacts, on a half-county basis. The rule-based model currently being used contains the following: (1) proximity of the bloom to the coast, (2) transport of the bloom, (3) concentration of cells in the water, and (4) wind speed and direction. The resolution of impact forecasts is coarse, providing information daily on a half county basis. Therefore, our current system is capable of predicting whether an impact, and at what level, will occur anywhere within approximately 30 km of coastline on a particular day. However, the forecast lacks the resolution to provide detailed information for a particular beach. In August 2006, professional lifeguards in Sarasota and Manatee Counties began providing real-time respiratory impact information twice a day. These data combined with meteorological measurements are being used to develop a finer resolution respiratory impact model, with the goal of providing more accurate estimates of beach impact, on a twice daily basis.

ECOHAB PACIFIC NORTHWEST (ECOHAB PNW) OUTREACH: OPENING THE SCIENTIFIC JOURNEY TO THE WORLD

Vera L. Trainer¹, William P. Cochlan², Mark L. Wells³, Charles G. Trick⁴, and the ECOHAB education group (Lauren Kuehne⁵, Christine Muir⁶, Dennis Costello⁷)

¹NOAA Northwest Fisheries Science Center, 2725 Montlake Blvd. E., Seattle, WA 98112

²Romberg Tiburon Center for Environmental Studies, San Francisco State University, Tiburon, CA 94920

³School of Marine Sciences, University of Maine, Orono, ME, 04469-5741

⁴Schulich School of Medicine, University of Western Ontario, London, ONT, N6A 5B7, Canada

⁵Evergreen State College, Olympia, WA, 98505

⁶Woodside Priory School, Portola Valley, CA 94028

⁷North High School, Torrance CA, 90504



Public and education outreach has been an integral part of the ECOHAB Pacific Northwest project from its inception in 2002. Public outreach activities include a project-dedicated web site, public contact, and multiple interactions with journalists over the course of the study. Educational activities have included participation at in-service teacher workshops, and hosting four teachers-at-sea on research cruises through: 1. the NSF Research Experience for Teachers (RET), 2. the West Coast Center for Oceans and Human Health, 3. the NSF ARMADA program, 4. individual NSF research grants, and 5. the NOAA Hollings undergraduate summer scholarship program. Real-time cruise journals targeting high school level students and the general public were created and maintained while at sea. These journals detailed the scientific journey of oceanographic research, and in particular the challenges investigating harmful algal blooms, collaborative research activities, and human-interest aspects of life at sea. During our final cruise in September 2006, as a collaboration of

undergraduate, and graduate students, principal investigators, and Evil Bunny Films, two documentary films (10- and 20-minutes) were produced. These films detail the scientific problem, the collaborative approach to oceanographic research, and the vision for the future of forecasting of harmful algal blooms in the Pacific Northwest. The 20-minute film is targeted for both secondary and undergraduate audiences and details the complexity of science at sea. It is accompanied by the "Harmful Algal Bloom Hunter's Handbook", a series of lesson plans consisting of classroom activities and experiments, cruise journals, a plankton identification chart and a glossary. These outreach materials will be presented, and the films displayed at the poster session during the 4th US HAB Symposium. See also: www.ecohabpnw.org/outreach

***Pseudo-nitzschia* GROWTH AND TOXIN PRODUCTION IN THE JUAN DE FUCA EDDY IN THE PACIFIC NORTHWEST – ENVIRONMENTAL STIMULATORS OF A TOXIC BLOOM**

Charles G. Trick¹, William P. Cochlan², Mark L. Wells³, Vera L. Trainer⁴.

¹Schulich School of Medicine and Dentistry, University of Western Ontario, Ontario N6A5B7, Canada, ²Romberg Tiburon Center for Environmental Studies, San Francisco State University, Tiburon, 94920, USA, ³School of Marine Sciences, University of Maine, Orono, ME 04469, USA, ⁴NOAA Northwest Fisheries Science Center, Seattle, 98112 USA

As part of the ECOHAB-PNW project to study the community formation, domoic acid (DA) toxicity and transport of *Pseudo-nitzschia* cells from the shelf waters of Washington State and British Columbia to the shoreline, we examined spatial distribution of key HAB parameters: total phytoplankton biomass, contribution of *Pseudo-nitzschia* species, and absolute and cell specific DA concentrations. In September 2003 and September 2004, we used deck-board incubation “grow-out” experiments to evaluate if the natural phytoplankton community could be stimulated with modifications of the water chemistry to either increase the biomass further, increase the relative frequency of *Pseudo-nitzschia*, increase the physiological capacity of the community (P vs. E), or increase the cell-specific or community integrated toxicity due to DA. While there were variations in the community responses to macronutrient additions, cells from several specific water masses responded primarily to nanomolar additions of iron, or to the addition of a nanomolar mixture of iron and copper. We assessed the stimulation of the HAB component of the community physiology by the extent of enhanced growth of *Pseudo-nitzschia*, the cellular and dissolved levels of DA, the recovery of the photosynthetic capacity of the phytoplankton, and to a less frequent extent, alterations in the buoyancy/sinking trend of the phytoplankton. Yearly variation in the late summer phytoplankton communities indicates that there is a complex regional regulation of phytoplankton community rich in *Pseudo-nitzschia*, but the commonly implicated macronutrient concentrations are poor predictors of either *Pseudo-nitzschia* dominance or toxicity. Rather, micronutrient additions of iron and copper stimulated the dominance, toxicity and physiological health in large areas of the Juan de Fuca eddy region and implicate spatial variation in trace metals as the prime proponent of toxicity in these waters.

CHARACTERIZATION OF NOVEL COMPOUNDS FOUND IN *Karenia brevis* CULTURES

Laura T. Truxal and Daniel G. Baden

University of North Carolina Wilmington, Center for Marine Science, Wilmington, NC 28403, USA

Florida red tides occur in the Gulf of Mexico almost annually as the result of blooms of the unarmored marine dinoflagellate *Karenia brevis*. *K. brevis* is best known for the production of a family of polyether neurotoxins known as brevetoxins. Brevetoxins are compounds found in the organic phase (chloroform layer) of the liquid/liquid extraction of *K. brevis* cultures. Additional compounds produced by *K. brevis* include brevenal, brevisin, brevisamide, hemibrevetoxin B (Prasad and Shimizu, 1989), as well as fatty acids and phytopigments. Although the chloroform layer has been extensively mined for bioactive components, very little emphasis has been put on exploration of the more polar and non-polar components of the *K. brevis* cultures. This project focuses on the petroleum ether (non-polar) layer and the water (polar) layer of *K. brevis* culture. Methods for this project include liquid/liquid extraction, solid phase extraction, LC/MS, bioassay guided fractionation, chromatography techniques, and NMR. The various bioassays used to determine active compounds include antibacterial assays, antifungal assays, cytotoxicity assays, receptor binding assays, and fish bioassays. Previous work with *K. brevis* cultures, or samples from natural blooms of *Karenia* in Gulf Coast waters, have identified polar brevetoxin metabolites related structurally to known A and B type brevetoxins, in which the lactone ring of the brevetoxin backbone structure is open (Abraham et al., 2006). If these brevetoxin metabolites are found, along with any other compounds, they will provide useful standards for analytical work and tools for biological research. Results from this project will help to better characterize *K. brevis* blooms and their associated physiological effects, as any compound produced by *Karenia* has the potential to effect humans and animals. Any new bioactive components found may add to the growing list of marine natural products used as new drug candidates and will be evaluated for physiological effects and contributions to red tides. Preliminary work has determined a method to separate the non-polar petroleum ether layer of *Karenia* culture and has identified both cytotoxic (masses: (m+1)=413, (m+1)=801, (m+1)=871) and antibacterial (masses: (m+1)=373, (m+1)=618) compounds. A method for separating the polar water layer of *Karenia* culture has been determined and has yielded novel cytotoxic compounds with masses of (m+1)=237, (m+1)=251, and (m+1)=546.

Abraham, A., Plakas, S.M., Wang, Z., Jester, E.L.E., El Said, K.R., Granade, H.R., Henry, M.S., Blum, P.C., Pierce, R.H., Dickey, R.W., 2006. Characterization of polar brevetoxin derivatives isolated from *Karenia brevis* cultures and natural blooms. *Toxicon* 48, 104-115.

Prasad, A.V.K. and Shimizu, Y., 1989. The structure of hemibrevetoxin B: A new type of toxin in the Gulf of Mexico red tide organism. *Journal of the American Chemical Society* 111 (16), 6476-6477.

THE RELATIONSHIP BETWEEN COASTAL OCEAN DYNAMICS AND SHELLFISH CLOSURES: A SATELLITE BASED STUDY OF OREGON HABS

Jacqui Tweddle¹, Pete Strutton¹, Michelle Wood², Deb Cannon³ and Brittany Scott²

¹College of Oceanic and Atmospheric Sciences, Oregon State University

²Department of Biology, University of Oregon

³Oregon Department of Agriculture

Toxic blooms of *Pseudonitzschia* or *Alexandrium* in Oregon coastal waters can lead to accumulation of domoic acid or saxitoxin (respectively) in coastal shellfish, primarily razor clams. Since 1979, the Oregon Department of Agriculture (ODA) has monitored coastal shellfish for phycotoxins, with a view to protecting public health. This database is most comprehensive for both toxins from about 1998, which conveniently corresponds to the era of the SeaWiFS ocean color satellite. This work analyzes seasonal and interannual patterns in the toxin database in conjunction with satellite data. Using satellite chlorophyll data from SeaWiFS, sea surface temperature data from NOAA AVHRR satellites and several wind products (satellite, in situ and an upwelling index) we have quantified the seasonal and interannual variability in coastal ocean physics and biology. There are at least three features which significantly impact the coastal ocean and the frequency of HAB impacts: The Columbia River outflow, Heceta Bank and Cape Blanco. The Columbia and Heceta Bank regions are characterized by a broader near-coastal band of high productivity and more frequent shellfish closures, particularly for domoic acid. Cape Blanco is associated with a 'break' in the patterns of coastal winds and currents. The physics and biology to the north and south of the cape differ in phase and magnitude, with concomitant impacts on HABs and shellfish contamination. At all locations there is significant interannual variability in the timing of HAB events, driven mainly by the major transitions to upwelling and downwelling conditions. The results of these analyses enable us to make generalizations about the likelihood of HABs and shellfish contamination within subregions of the Oregon coast, which in turn increases the efficacy of monitoring programs.

IDENTIFICATION AND ENUMERATION OF *Pseudo-nitzschia* IN PACIFIC NORTHWEST COASTAL WATERS USING THE FLOWCAM® CONTINUOUS IMAGING PARTICLE ANALYZER

Elyse A. Walker¹, Dana Woodruff²

¹University of South Carolina, Columbia, SC 29225, USA; ² Battelle Marine Sciences Laboratory, Pacific Northwest National Laboratory, Sequim, WA 98382, USA

Toxicogenic blooms of *Pseudo-nitzschia* are becoming increasingly apparent recently in the Puget Sound waters of Washington State. Until recently, the primary impact has been on the razor clam and Dungeness crab fisheries on the outer coast of the state causing recreational, commercial and tribal subsistence closures on a routine basis. Early detection and rapid screening methodologies are an important component of understanding and mitigating for these bloom events. In this preliminary investigation we focused on the development of a methodology using the FlowCAM for quickly identifying *Pseudo-nitzschia* in the waters of Sequim Bay Washington, a coastal embayment that has recently been plagued with *Pseudo-nitzschia* blooms. The FlowCAM® couples the capabilities of a flow cytometer with a digital-imaging microscope to automate phytoplankton detection and enumeration. For this study, we collected weekly samples during the summer months at the mouth of Sequim Bay on an incoming and outgoing tide. During this time a bloom of *Pseudo-nitzschia* occurred which provided the opportunity to explore possible methods of identifying *Pseudo-nitzschia* in natural phytoplankton assemblages using the FlowCAM, with a particular emphasis on identifying bloom conditions. A variety of parameters were examined including length, width, aspect ratio, area based diameter, equivalent spherical diameter, transparency, perimeter, etc.... Based on statistical analysis of these particles, only the aspect ratio was consistent and unique for *Pseudo-nitzschia* during this particular bloom event. Further research is needed to determine the consistency of these results under other bloom conditions and locations with varied community composition. However, these results show promising for rapid detection and screening of phytoplankton samples for *Pseudo-nitzschia* in Pacific Northwest coastal waters.

EFFECT OF CADMIUM, COPPER, NICKEL, AND ZINC ON A MINUTE GOLDEN BROWN ALGA *Aureococcus anophagefferens*

Bin Wang¹, Lisa Axe¹, Liping Wei², Sima Bagheri¹, Zoi-Heleni Michalopoulou³

¹Department of Civil and Environmental Engineering,

²Department of Chemistry and Environmental Science,

³Department of Mathematical Science

New Jersey Institute of Technology, Newark, NJ 07102, USA

The project focuses on the harmful algal blooms. *Aureococcus anophagefferens* is a toxic pelagophyte observed in the northeast of US. The present study examines the effect of Cd, Cu, Ni and Zn on *A. anophagefferens*. Two ranges of free metal concentrations are studied for accumulation and toxicity of metals. For the low free metal concentration (10^{-12} M - 10^{-9} M for Cd, Cu and Ni, 10^{-12} M - 10^{-8} M for Zn), *A. anophagefferens* is grown in Aquil and the growth kinetics studies are carried out. The free metal concentrations are controlled by adding 100 or 10 μ M EDTA and are calculated by MINEQL. Thus, the optimal and toxic free metal concentration for *A. anophagefferens*'s growth and its growth rate are obtained. For the high free metal concentration (10^{-10} M - 10^{-5} for Cd, 10^{-10} M - 10^{-8} for Cu, 10^{-10} M - 10^{-5} for Zn, and 10^{-10} M - 10^{-6} for Ni), *A. anophagefferens* cells are subject to short term exposure (30 min to 24 hr) of single metal in synthetic ocean water (SOW, pH = 8.1, ion strength = 0.7, no EDTA addition). Metal accumulation by cells is determined by inductively coupled plasma mass spectrometry (ICP-MS) upon rinse with SOW-EDTA solution. Therefore, the cellular and intracellular metal fractions, metal uptake, and the affinity of each metal for *A. anophagefferens* are being investigated. Based on these results, the effects of co-contaminants as well as mechanistic uptake of metals on *A. anophagefferens* will be studied.

GROWTH RESPONSE AND GLUTATHIONE PRODUCTION OF BROWN TIDE BLOOM ALGA (*Aureococcus anophagefferens*, CCMP 1984) UPON DIFFERENT SALINITY, METALS, NITROGEN SOURCE, SEWAGE AND HERBICIDE METOLACHLOR EXPOSURE: LAB CULTURE STUDIES AND IN SITU INCUBATION STUDIES

Liping Wei¹, Bin Wang²

¹ Chemistry and Environmental Science

² Civil and Environmental Engineering

New Jersey Institute of Technology

Newark, NJ 07102

wei@adm.njit.edu

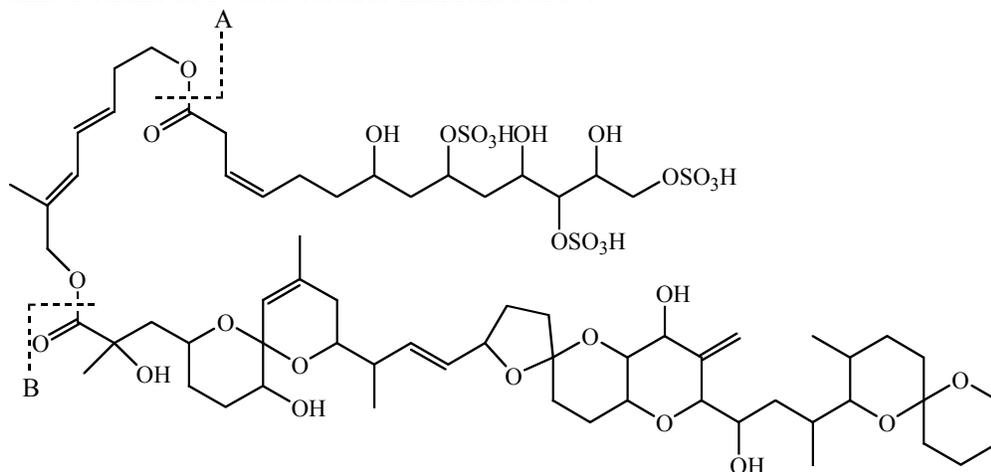
It is well known that N, P and trace metals strongly affect phytoplankton growth. Brown tide bloom is caused by the overgrowth of pelagophyte *Aureococcus anophagefferens*. Organic nitrogen source, metals, salinity seems to affect bloom formation. We study how environmental factors - salinity, sewage overflow, herbicide metolachlor and trace metals affect the growth and cellular redox status of *A. anophagefferens*. We report the growth and cellular glutathione levels of *A. anophagefferens* (CCMP 1984) upon long term exposure to different salinities, herbicide metolachlor and sewage overflow in artificial seawater media Aquil or L1. We also conduct *in situ* incubations of *A. anophagefferens* in natural seawater, where lab cultured *A. anophagefferens* are inoculated to natural seawater collected from Barnegat Bay-Little Egg Harbor, and Newark Bay, NJ, with amended nitrate, phosphate, Fe, Zn, Cu, glycine, urea, sewage, and herbicide metolachlor, and are incubated in Newark Bay NJ during July 2007. *A. anophagefferens* cells in the *in situ* incubations are monitored overtime by monoclonal antibody according to Caron et al., 2003. Together we hope to find out the controlling factors toward brown tide bloom initiation.

STUDIES ON THE ENZYMATIC HYDROLYSIS OF DSP ESTERS TO PRODUCE THE TOXIN, OKADAIC ACID, IN THREE STRAINS OF *Prorocentrum lima*

Kristy V. White¹, Eve M.H. Wright and Jeffrey L.C. Wright

University of North Carolina at Wilmington, Department of Chemistry and Biochemistry, Center for Marine Science, Wilmington, NC 28409, USA

Diarrhetic shellfish poisoning (DSP) is a human gastrointestinal disease that is caused by the consumption of shellfish contaminated by okadaic acid (OA) or one of the OA isomers, dinophysistoxin-1 (DTX-1) and -2 (DTX-2) (Windust et al., 1997). DSP toxins are polyether fatty acids that are all structurally similar and vary only by the number of pendant methyl groups attached to the main chain (Murata, M. et al., 1985). DSP toxins are produced by dinoflagellates belonging to the genera *Dinophysis* and *Prorocentrum* (Souto et al., 2001), and enter the food chain when filter-feeding bivalves ingest them. Symptoms of DSP include diarrhea, nausea, vomiting, abdominal pain and chills (Morton et al., 1996). DSP poses serious impacts on public health and the aquaculture industry worldwide (Souto et al., 2001). The focus of this research was to examine the hypothesis that in the dinoflagellate, *Prorocentrum lima*, DSP toxins are stored in the biologically inactive form within the cell as water-soluble sulfated diesters (e.g. DTX-4 and DTX-5), and that these storage products can be hydrolyzed via an esterase or lipase to OA diol-ester and OA upon rupture or attack of the *Prorocentrum* cell. As exhibited in the figure below, hydrolysis of DTX-4 at "A" yields the pro-toxin OA diol-ester; hydrolysis of DTX-4 at "B" yields OA itself. Cells and cell medium from 3 different strains of *P. lima* cultures (CMS TAC PL 010, CCMP 1743 NS, and CCMP 1746 Bz) were examined for the presence of an esterase or lipase that specifically converts the DSP derivatives (e.g. DTX-4 and OA diol-ester) to OA. Through the employment of a surrogate substrate, hydrolytic activity was followed throughout the duration of growth in both the cells and cell medium of *P. lima* cultures. This hydrolytic activity was then correlated with the concentration of OA measured throughout the growth period in both the cells and cell medium of *P. lima* cultures. Additionally, the purity of the hydrolytic enzyme was increased through several chromatography steps utilizing a surrogate substrate to guide the purification process. The hydrolytic enzyme of increased purity was then monitored for specific activity against the natural DSP ester substrates isolated from the same 3 strains of *P. lima* cultures mentioned above.



Morton, S.L. et al., (1996). "Determination of okadaic acid content of dinoflagellate cells: a comparison of the HPLC-fluorescent method and two monoclonal antibody ELISA test kits." *Toxicon* 34: 947-954.

Murata, M. et al., (1985). "Diarrhetic shellfish toxins." *Tetrahedron* 41(6): 1019-1025.

Souto, M.L. et al., (2001). "Influence of amino acids on okadaic acid production." *Toxicon* 39: 659-664.

Windust, A.J. et al., (1997). "Comparative toxicity of the diarrhetic shellfish poisons, okadaic acid, okadaic acid diol-ester and dinophysistoxin-4 to the diatom *Thalassiosira weissflogii*." *Toxicon* 35: 1591-1603.

VARIABLE EXPRESSION OF TOXICITY IN *Prorocentrum minimum*, AND POSSIBLE RELATIONSHIPS WITH TROPHIC STATUS

Gary H. Wikfors¹, Yaqin Li¹, Shannon L. Meseck¹, H el ene H egaret², Gulnihal Ozbay³, and Allen R. Place⁴

¹NOAA Fisheries Service, Milford, CT 06460 USA; ²Department of Marine Sciences, University of Connecticut, Groton, CT 06340 USA, ³Delaware State University, Dover, DE 19901 USA; ⁴Center of Marine Biotechnology, University of Maryland, Baltimore MD 21202 USA

Variable toxicity in harmful microalgal species has been long recognized but not thoroughly evaluated in evolutionary and ecosystem contexts. Recent findings suggest that expression of toxicity in several Harmful Algal Bloom (HAB) species represents an active survival strategy. For example, toxicity in *Alexandrium minutum* is induced by the presence of copepod grazers (Selander et al. 2006). *Dinophysis* populations regulate toxin content per cell based upon population density (Lindahl et al. 2007). Karlotoxins are produced by *Karlodinium veneficum* as “predatory venom,” slowing the swimming speed of cryptophyte prey (Adolf et al. 2006). Thus, toxins produced by photosynthetic or mixotrophic protists may have defense or nutritional functions, and expression of toxicity, i.e., transcription and translation of the genetic capacity for toxin synthesis, represents a response to a specific environmental stimulus.

The estuarine dinoflagellate, *Prorocentrum minimum*, has been recognized as a “HAB,” based upon observations of mortalities of marine organisms coincident with blooms, as well as laboratory experiments showing inimical effects upon grazers (Heil et al. 2005). Nevertheless, blooms of this dinoflagellate can occur without apparent harmful effect, and laboratory experiments yield variable results (Wikfors 2005). Although the chemical structures of putative *P. minimum* toxins have not been described, biological effects caused by pure cultures include bacteriostatic activity (Trick et al. 1984), allelopathy (Denardou-Queneherve et al. 1999), digestive and immunological disruption in molluscs (Wikfors 2005, H egaret & Wikfors 2005), and acute toxicity to mice when extracts are injected intraperitoneally (Denardou-Queneherve et al. 1999). A repeated theme in these studies is that cultures “in decline” tend to be more toxic than those growing vigorously. We have enhanced expression of toxicity in *P. minimum* cultures by starving them for phosphorus (P), inducing bacteriostatic activity, or restricting air flow in aerated cultures, enhancing molluscicidal activity. An ecological interpretation of these findings suggests that P stress may induce a switch from photosynthetic to bacteriotrophic nutrition, but a nutrient-replete, photosynthetic population limited by inorganic carbon may inhibit grazing to maintain population growth. We have found changes in proteins released into the medium by *P. minimum* when stressed for P or inorganic carbon, as well as changes in fatty-acid composition and pigment content. We hypothesize that expression of toxicity in this dinoflagellate may be associated with a shift from autotrophic to heterotrophic nutrition and are testing this hypothesis experimentally.

References:

- Adolf, J.E., T.R. Bachvaroff, D.N. Krupatkina, H. Nonogaki, P.J.P. Brown, A.J. Lewitus, H.R. Harvey & A.R. Place. 2006. Species specificity and potential roles of *Karlodinium micrum* toxin. *Afr. J. Mar. Sci.* 28, 415-419.
- Denardou-Queneherve, A., D. Grzebyk, Y.F. Pouchus, M.P. Sauviat, E. Alliot, J.F. Biard, B. Berland & J.F. Verbist. Toxicity of French strains of the dinoflagellate *Prorocentrum minimum* experimental and natural contaminations of mussels. *Toxicon* 37, 1711-1719.
- H egaret, H. & Wikfors, G.H. (2005). Time-dependent changes in hemocytes of eastern oysters, *Crassostrea virginica*, and northern bay scallops, *Argopecten irradians irradians*, exposed to a cultured strain of *Prorocentrum minimum*. *Harmful Algae* 4, 187-199.
- Heil, C.A., P.M. Glibert & C. Fan. 2005. *Prorocentrum minimum* (Pavillard) Schiller: A review of a harmful algal bloom species of growing worldwide importance. *Harmful Algae* 4, 449-470.

- Lindahl, O., B. Lundve & M. Johansen. 2007. Toxicity of *Dinophysis* spp. in relation to population density and environmental conditions on the Swedish west coast. *Harmful Algae* 6, 218–231.
- Trick, C.G., R.J. Andersen, & P.J. Harrison. 1984. Environmental factors influencing the production of an antibacterial metabolite from a marine dinoflagellate, *Prorocentrum minimum*. *Can. J. Fish. Aquat. Sci.* 41: 423-432.
- Selander, E., P. Thor, G. Toth & H. Pavia. 2006. Copepods induce paralytic shellfish toxin production in marine dinoflagellates. *Proc. R. Soc. B*, 273, 1673–1680.
- Wikfors, G.H. (2005). A review and new analysis of trophic interactions between *Prorocentrum minimum* and clams, scallops, and oysters. *Harmful Algae* 4, 585-592.

SPECIFICITY OF BACTERIAL ASSEMBLAGES ASSOCIATED WITH THE TOXIN-PRODUCING DIATOM, *Pseudo-nitzschia*Michele L. Wrabel¹ and Gabrielle Rocap¹¹University of Washington, School of Oceanography, Seattle, WA 98195, USA
Pacific Northwest Center for Human Health and Ocean Studies

The marine diatom *Pseudo-nitzschia*, responsible for Amnesic Shellfish Poisoning (ASP) via production of the toxin domoic acid, is widely distributed in Puget Sound, Washington. Over the past four years, three shellfish bed closures have occurred due to domoic acid events in this estuary. Previous laboratory studies have linked enhanced domoic acid production to higher bacterial abundance and morphological diversity, among other potential triggers. The nature of interactions between bacteria and *Pseudo-nitzschia* has not been elucidated, and few data have described bacterial assemblages native to *Pseudo-nitzschia*. We used Automated Ribosomal Intergenic Spacer Analysis (ARISA), a DNA fingerprinting technique based on length differences in the ribosomal RNA intergenic spacer, to characterize bacteria associated with laboratory cultures of 19 *Pseudo-nitzschia* strains (representing 7 species) and 11 other diatom strains. Individual strains supported significantly different bacterial assemblages. Likewise, *Pseudo-nitzschia* species were associated with different bacterial assemblages. However, differences in bacterial ARISA profiles did not exist between diatom genera. These data suggest that specific associations between diatoms and bacteria have evolved at the diatom species and strain levels. For some strains, attached bacteria (> 3 μm fraction) were significantly different from free-living bacteria (0.2 – 3 μm fraction). No significant differences in bacterial assemblages existed during exponential and stationary phases of *Pseudo-nitzschia* growth. One *Pseudo-nitzschia* strain, sampled three times over its initial nine months in culture, did not exhibit any shifts in bacterial assemblages. Furthermore, a substantial number of ARISA operational taxonomic units (OTUs) were shared between such recently-isolated Puget Sound strains and field samples corresponding to the diatoms' origins. Several of these OTUs are responsible for statistically significant differences in bacterial assemblages between *Pseudo-nitzschia* species. We conclude that bacteria coexisting with our Puget Sound *Pseudo-nitzschia* cultures are representative of species-specific associations found in the field. Further work will identify, isolate, and culture *Pseudo-nitzschia*-associated bacteria for physiological experiments testing specificity of associations as well as bacterial influence on toxin production. Analyses of field data will investigate the extent to which species-specific associations between diatoms and bacteria influence the distributions of both groups in Puget Sound.

NUTRIENT-REGULATED TRANSCRIPTOME PROFILING IN THE BROWN-TIDE FORMING ALGA *Aureococcus anophagefferens*

Louie L. Wurch¹, Sheean T. Haley¹, Elizabeth D. Orchard¹, and Sonya T. Dyhrman¹

¹Woods Hole Oceanographic Institution, Woods Hole, MA 02540, USA

HABs are a global problem with widespread effects on public health, the coastal environment, and the economy. Understanding the links between HAB dynamics and nutrient supply is a critical area of ongoing study. Assessing the nutritional physiology of a single HAB species in a mixed community remains a challenge, although our increasing molecular-level understanding of nutritional physiology in some species is providing both the knowledge for hypothesis building and testing in the field. *Aureococcus anophagefferens*, the species responsible for “brown-tide” events, provides an excellent model for studying HABs with the upcoming availability of its genome sequence. In this study, we used Long-SAGE (Serial Analysis of Gene Expression) to identify gene expression patterns in *A. anophagefferens* grown under nitrogen (N) and phosphorus (P) deficient and replete conditions. This approach generates tag libraries without *a priori* knowledge of gene sequences via the detection of 21 bp nucleotide sequence tags. These tags were mapped back to individual genes and used to examine transcriptional responses to nutritional state. The sampling frequency of these tags in different libraries indicates their differential gene expression pattern. To date, we have sampled over 86,000 tags representing 8,601 unique sequences and annotation of these tags is currently in progress. A broad-scale comparison shows a greater number of tags showing elevated transcriptional signal in the low P library than in the low N library. We have identified a number of significantly up-regulated tags (R-value >2) in both the low N and low P libraries. Relative to the nutrient replete library, 14 tags from the low P library and 5 tags from the low N library show a higher than 10-fold change in expression levels. At present, none of these highly regulated tags map to publicly available sequence data. However, the pending genome coupled with tag validation may identify genes that can be good diagnostic indicators of cellular nutritional physiology. Through a species-specific quantitative RT-PCR assay that we are developing, it will be possible to examine the expression patterns of these key genes in field populations. Looking at the N and P physiology of the cells in the field over the course of a bloom event will provide insight into the factors influencing bloom dynamics.

DETECTING CYANOBACTERIA BLOOMS USING MERIS

Timothy T. Wynne, Michelle C. Tomlinson¹, Richard P. Stumpf¹

¹National Ocean Service, Silver Spring, MD 20910, USA

MERIS, a European ocean color satellite, has increased spectral resolution relative to its domestic counterparts, MODIS and SeaWiFS. Using the increased spectral resolution it is possible to improve differentiation of cyanobacteria blooms from blooms of phytoplankton due to differences in their optical properties. Cyanobacteria often produce surface scum when cells are present in large concentrations. Additionally many species of cyanobacteria can regulate their buoyancy with gas vacuoles, making them extremely efficient at scattering light. Cyanobacteria also have phycocyanin, an auxiliary pigment, which most species of phytoplankton lack. Previously published methods used to detect blooms were run on a time-series from western Lake Erie, where *Microcystis* spp. is a potential bloom forming species. The remote sensing products were then entered as attributes into a rule based model, and the model was trained to produce an output of yes or no, meaning that a cyanobacteria bloom is present or a cyanobacteria bloom is not present. Initial results are promising. The results from this model were run on a quasi-realtime basis during the summer of 2007 in an effort to try and detect and monitor cyanobacteria blooms in the Laurentian Great Lakes.

COMPARATIVE REACTIVITY OF DIFFERENT CYSTEINE CONGENERS AS DETOXYFING AGENTS OF BREVETOXINS

Xingye Yang¹, and Jerome Naar¹

¹University of North Carolina at Wilmington, Center for Marine Science, Wilmington, NC 28409, USA

Brevetoxins are potent marine neurotoxins produced by the marine dinoflagellate *Karenia brevis*. During blooms of this organism (red tides) in the gulf of Mexico, brevetoxin exposure results in massive fish kill, marine mammal deaths, and upper and lower airway impairments in humans exposed to contaminated sea-spray. These toxins also accumulate to dangerous levels in shellfish causing Neurotoxic Shellfish Poisoning if consumed. We have recently demonstrated that cysteine possesses striking properties as it can spontaneously react with some of these toxins to form non-toxic derivatives and thus could be used as a mitigating agent of red tides impacts¹. The present research has been conducted to evaluate the potential reactivity of different cysteine congeners with brevetoxins to identify potential other mitigating agents. Several congeners have been examined for their efficiency in reacting with brevetoxins in aqueous environment. Both type 1 and 2 brevetoxins (PbTx1 and PbTx2) have been selected to react with these congeners at different molar ratios. Since red tides of *K. brevis* have also been observed in springs and winters, the reactivity of the congeners with brevetoxins was examined at different temperatures (22°C and 8°C). The disappearance of these toxins and the appearance of the non-toxic derivatives of brevetoxins over time have been monitored by LCMS and compared to identify the best conditions for mitigating the harmful effects of brevetoxins.

NAAR, J., and L. J. FLEWELLING, W. M. ABRAJAM, H. JACOCKS, A. LENZI, X. YANG, A. BOURDELAIS, S. MICHELLIZA, C. TOMAS, and D. G. BADEN. The mitigating properties of cysteine on the harmful effects of red tide. This conference.

FLORIDA'S RED TIDE CONTROL & MITIGATION GRANT PROGRAM: THE BEGINNING

Leigh Zimmermann¹, Meghan Shone², and Leanne Flewelling²

¹Solutions To Avoid Red Tide, Bradenton, FL 34205, USA

²Florida Fish and Wildlife Conservation Commission, Fish and Wildlife Research Institute, St. Petersburg, FL 33701, USA

A newly formed competitive grant program in Florida is dedicated to funding research for control of *Karenia brevis* blooms and strategies to mitigate their effects. The Red Tide Control & Mitigation Grant Program is designed to explore environmentally acceptable techniques or technologies which minimize the size, intensity, or duration of *K. brevis* blooms or reduce environmental, economic, social, or public health impacts of red tide blooms in Florida. The program is implemented by the Florida Fish and Wildlife's Conservation Commission's (FWC) Fish and Wildlife Research Institute (FWRI). A grant to a local non-profit organization, Solutions To Avoid Red Tide (START), provides promotion of the Red Tide Control & Mitigation Grant Program and coordination of the Red Tide Control & Mitigation Panel. A nine-member external panel of representatives from state and local governments, agencies, and advocacy groups has been appointed to three-year, renewable terms to provide peer reviews of grant submissions. The grant panel assisted in the development of the first request for proposals, established the review process and created evaluation criteria. Panelists then reviewed, recommended, and prioritized projects for funding. Based on the panel's recommendations, funded research covers a broad range, including determining socioeconomic costs associated with various elements of Florida red tide, investigating reduction of toxicity through a variety of controls, and the production of a documentary on *K. brevis*. This program addresses an identified need in funding for applied research and responds to a public outcry in the State of Florida. As called for in Harmful Algal Bloom Research and Response: A National Environmental Science Strategy (HARRNESS, 2005), the Red Tide Control & Mitigation Grant Program is directed at identifying practical strategies to mitigate bloom impacts on the people and resources of Florida. The backbone of this program will possibly serve as a model for other regions in the United States hoping to proactively address mitigation and control of harmful algal blooms.

HARRNESS, 2005. Harmful Algal Bloom Research and Response: A National Environmental Science Strategy 2005-2015. Ramsdell, J.S., D.M. Anderson, and P.M. Glibert (Eds.), Ecological Society of America, Washington DC, 96 pp.

AUTHOR INDEX

Abbott, J.P.	133	Bowers, H.A.....	20, 89
AbiNader, M.	132, 176	Boyer, G.L.	47 , 77, 125, 137, 168
Abraham, A.	33, 63, 66, 155	Boyes, C.	51, 140
Abraham, W.M.	152	Brandsrud, S.....	57
Abrahamson, A.	123, 126	Bratcher, A.	98
Adams, C.M.	25	Braun-Howland, E.....	135
Adams, N.G.	19	Bricelj, V.M.	27 , 108
Adolf, J.E.	20 , 55, 56, 89	Bronder, Z.E.	107
Alexander, J.	40, 129	Brosnahan, M.L.	90
Allen, A.L.	107	Brown, C.	29
Altland, K.	22	Burbacher, J.	42
Alvarado, N.	43	Burkholder, J.M.....	167
Ammerman, J.	81	Burson, A.	91 , 173
Anderson, D. M. 21 , 37, 57, 58, 70, 90, 131, 145		Busse, L.....	94
Anderson, J.T.	82	Calkins, J.A.....	92 , 180
Armbrust, E.V.	48	Campbell, L.	93 , 122
Auro, M.E.....	83 , 157	Cannon, D.	171, 184
Avery, D.E.....	22 , 30, 157, 162	Carmichael, W.	23
Axe, L.	186	Caron, D.A.....	69, 169
Ayres, D.L.	101	Carter, M.	94
Baatz, J.E.....	143	Carvalho, G.A.....	95
Bachvaroff, T.R.....	20, 62, 89	Cassedy, A.	39
Backer, L.C.....	23 , 39, 130, 139, 153, 180	Cembella, A.D.	109
Baden, D.G.....	113, 152, 183	Cetinic,I.....	69, 169
Bagheri, S.	186	Chase, A.	132
Bai, X.	169	Chen, L.	30
Ballauer, J.M.	60	Cheng, Y.-S.....	23
Bammler, T.....	52	Chrest, F.J.	96
Banzon, V.	95	Clemons, S.	133
Bargu Ates, S.....	24 , 100, 110, 165	Coats, D.W.....	70, 89
Baringer, W.	95	Coblentz, F.....	97
Bauer, M.	25	Cochlan, W.P. ... 31 , 45, 53, 83, 84, 86, 157, 181, 182	
Beall, B.	84	Codispoti, L.	129
Bean, J.A.	39, 153	Connell, L.....	98 , 103, 119
Berg, M.	41	Costello, D.	181
Berman, M.....	106	Couture, D.....	21
Berry, D.L.....	41, 173	Cox, A.M.	123, 126
Beyer, R.....	52	Cox, F.H.....	101
Bhattacharya, D.....	57, 85	Coyne, K.J.....	116, 142
Bill, B.D.	86	Cross, S.L.	99
Blackmore, C.....	66	Culver, M.	140
Blossom, H.	145	Cunningham, M.	133
Blum, P.	155	Currier, R.	51, 92, 130, 140, 180
Boland, C.....	146	Dam, H.G.	22, 30, 160, 162
Borchert, J.A.	36, 101, 117	Dash, P.	100 , 110
Borkman, D.G.	87	Davies-Vollum, S.	123, 126
Bottein, M.-Y.	65, 88	Davis, T.W.	32
Bourdelais, A.....	152	Day, S.A.	101
Boushey, C.	42		

de Salas, M.F.	149	Grassle, J.F.	81
Del Pizzo, G.	133	Grattan, L.M.	42 , 67
Dickey, R.W.	33 , 63, 66, 76, 155	Gray, C.G.	115
Dixon, K.	118	Green, L.	145
Dixon, M.S.	112	Greenfield, D.L.	34, 43
Donovan, C.	102	Greengrove, C.L.	123, 126
Dorsey, C.P.	154	Griffith, J.J.	116 , 142
Dortch, Q.	177	Grigorev, I.	41
Doucette, G.J.	34 , 43, 75, 177	Grinnell, M.	133
Duy, J.	103	Gross, T.	29
Dyble, J.	35 , 105, 147	Grünbaum, D.	178
Dyhrman, S.T.	36 , 50, 117, 192	Hackett, J.D.	57
El Said, K.R.	33, 63, 155	Hails, A.	51
Elliott, C.T.	34	Haley, S.T.	36, 50, 117 , 192
Erdner, D.L.	36, 37 , 57, 90, 117, 145	Hall, E.R.	118
Errera, R.M.	104	Hall, N.S.	56
Etheridge, S.M.	38	Hallegraeff, G.M.	149
Evans, R.	69	Hamilton, S.A.	98, 119
Everlove, C.	43	Hammond, R.	66
Fahnenstiel, F.L.	35, 105 , 147	Hardison, R.	170
Farin, F.	52	Hattenrath, T.	120
Feldman, J.	34	Hauser, L.	19
Fensin, E.E.	56	Hayashi, K.	121
Fenstermacher, L.E.	107	He, R.	21, 58
Fire, S.E.	106	Hégaret, H.	44 , 164, 189
Fisher, K.M.	107	Heil, C.A.	118, 129
Fitzpatrick, E.	69	Henderson, K.	51
Fleming, L.E.	39 , 130, 153	Henrichs, D.W.	93, 122
Flewelling, L.J.	63, 66, 133, 152, 155, 195	Henry, M.	63, 155
Flood, R.	132	Herndon, J.	157
Foley, D.	171	Herwig, R.P.	19
Ford, S.E.	108	Hickey, B.M.	31, 45 , 53, 86, 101
Freitag, M.F.	109	Hilbern, M.	94
Gaasterland, T.	55	Hill, V.R.	23
Gallagher, E.	52	Hillier, J.	51
Frost, B.W.	123	Hoagland, P.	46 , 127, 156
Garcia, A.C.	100, 110	Hollenbeck, J.	39
Garcia, S.M.	111	Hood, R.	29
Gawel, J.E.	123	Horner, R.A.	101, 123 , 178
Gill, J.	55	Hotto, A.M.	125
Giner, J.-L.	112	Howard, K.	47
Glibert, P.M.	40 , 128, 129	Hubbard, K.A.	48
Gobler, C.J.	32, 41 , 91, 114, 120, 173	Hubert, J.	123, 126
Gold, E.P.	113	Hunter, M.	69
Gold, J.R.	93, 122	Irvin, M.	23
Goleski, J.A.	114	Jacocks, H.M.	113, 152
Gordon, C.J.	88	James, D.	150
Gorrepati, S.	139	Janssen, P.	52
Granade, H.R.	33, 63, 155	Jayroe, D.	153
Granhholm, A.	133	Jensen, S.	34, 43
Grant, K.	42	Jerez, E.	39

Jester, E.L.E.	33, 63, 155	Libby, P.S.....	87
Jester, R.	49	Libera, K.	37, 145
Jin, D.	46, 127 , 156	Lidie, K.B.....	54 , 75
Johns, D.J.....	128	Lin, S.	30, 55 , 148
John, U.....	109	Lind, H.	131
Johnson, T.B.....	23	Litaker, R.W.	35, 56 , 74, 105, 147, 170
Jones, B.	69	Lobel, P.S.....	156
Jones, K.L.	34	Lona, B.....	36, 117
Juhl, A.R.....	50	Long, W.....	29
Kamykowski, D.	71	Loram, J.E.....	57
Kaveggia, S.	69	Love, R.....	140
Keafer, B.A.....	21, 58	Lovko, V.J.....	141
Keller, H.M.....	107	Luber, G.	76
Kelly, R.R.....	50	MacFadyen, A.....	45
Kelly, V.	129	MacIntyre, H.L.	154
Kieszak, S.M.	23	Mafra, L.....	27
King, K.L.	34, 177	Mangum, A.	116, 142
Kirkpatrick, B.	23, 39, 92, 130 , 140, 153, 156 180	Margot, K.L.	143
Kirkpatrick, G.	51 , 130, 140	Marin, R.	34, 43
Kiryu, Y.....	133	Massion, E.....	34
Kittredge, C.	146	Masura, J.	123
Klarer, D.M.	147	Mazzillo, F.....	111, 144
Kleindinst, J.....	131	McCauley, L.A.R.	37, 131, 145
Kotlewski, A.....	51	McGillicuddy, D.J.	21, 58
Kubanek, J.	64, 151	McGowan, J.....	94
Kudela, R.M.	102, 134, 146, 170	McManus, G.B.....	160
Kuehne, L.	178	Meritt, D.....	40
Kulis, D.M.	70, 131 , 145	Meseck, S.L.	189
Laine, E.....	132 , 173	Michalopoulou, Z.-H.....	186
Lambert, C.....	108	Michael, B.	29, 129
Lamp, L.L.....	25	Michelliza, S.	113, 152
Landsberg, J.H.	66, 133	Mikulski, C.M.....	34
Lane, J.Q.....	134 , 146	Milian, A.	153
Lange, T.....	133	Miller, M.	51, 139
Langer Atkinson, H.....	135	Miller, P.E.	69, 146
Langlois, G.W.	49, 146	Millie, D.F.....	105, 147
Larkin, S.L.....	25	Minnett, P.J.	95
Lasiter, A.D.	115	Miranda, L.....	148
Leblond, J.D.	115	Monroe, E.A.	59
Lee, J.	136	Mooney, B.D.....	149
LeEberhart, B.-T.	167	Morris, J.G.	42, 67
Lefebvre, K.....	52	Morrison, J.	68
Lehman, P.W.....	137	Morton, S.L.....	99 , 106, 150
Leighfield, T.....	150	Moustafa, A.	57, 85
Lekan, D.	138	Muir, C.....	181
Lenzi, A.	152	Myers, T.L.	64, 151
LePrell, R.....	139	Naar, J.....	64, 97, 151, 152 , 194
Lessard, E.J.....	31, 45, 53	Naylor, M.	29
Levin, E.D.	88	Neely, M.B.....	118
Li, Y.	189	Neller, K.....	150
		Nierenberg, K.	23, 39, 92, 130, 140, 153 , 177

Norton, K.....	21, 131	Rocap, G.....	19, 191
Novoveska, L.....	154	Roelke, D.L.....	104
O'Donnell, S.....	176	Roesler, C.....	132, 173
Odell, A.....	101	Rogers, Y.-H.....	55
Olson, R.J.....	90	Roman, B.....	34, 43
Orchard, E.D.....	192	Roth, P.B.....	75
Osborn, D.....	145	Rowe, J.....	67
Osborn, S.....	155	Ruberu, S.....	146
Ozbay, G.....	116, 142, 189	Ryan, J.....	75, 144
Paerl, H.....	56	Sarnelle, O.....	68
Paillard, C.....	108	Sarode, N.....	41
Palmer, L.....	69	Satchwell, M.F.....	47, 125, 137
Pargett, D.....	43	Schaffner, R.....	69
Parker, S.....	67	Scherer, C.W.....	25
Parrow, M.W.....	167	Schnetzer, A.....	69
Parsons, M.L.....	60	Schoener, D.....	160
Parsons, R.....	99	Scholin, C.A.....	34, 43
Patchen, R.....	73	Schultz, I.R.....	161
Paternoster, J.....	150	Schulz, K.L.....	168
Pederson, B.....	51	Schumacker, J.....	101
Perera, K.....	146	Scott, B.....	184
Peterson, T.....	102	Scott, P.....	63, 155
Pettit, C.....	25	Senft, C.....	162
Pickell, L.D.....	61	Sengco, M.R.....	25, 70
Pierce, R.....	63, 155	Settlemyer, C.J.....	60
Pigg, R.....	133, 147	Shehee, M.....	163
Pilskaln, C.H.....	21	Shone, M.....	133, 195
Pinckney, J.L.....	100, 104	Shost, S.J.....	135
Place, A.R.....	20, 55, 56, 62 , 89, 149, 189	Shumway, S.E.....	44, 164
Plakas, S.M.....	33, 63 , 155	Silver, M.W.....	24, 49, 111, 144, 146, 165 , 172
Plaky, S.....	66	Sinclair, G.A.....	71
Plumley, F.G.....	57	Sinclair, J.....	166
Polansky, L.....	156	Skelton, H.....	167
Poli, M.....	38	Skillman, A.....	161
Poorvin, L.....	41	Smayda, T.....	72
Postel, J.R.....	123	Smith, G.J.....	121
Prince, E.K.....	64 , 151	Smith, J.L.....	168
Radan, R.L.....	157	Smith, K.W.....	58
Radwan, F.....	65	Smith, W.L.....	154
Ramers, D.....	29	Smolowitz, R.....	44
Ramsdell, J.S.....	65, 88, 158	Song, G.....	69
Rediske, R.....	147	South, R.....	66
Reich, A.....	39, 66 , 130, 153	Southwell, B.....	166
Renshaw, M.A.....	93, 122	Srinouanprachanh, S.....	52
Rezvan, A.H.....	88	Stapleton, C.A.....	154
Richard, D. 1.....	33	Stauffer, B.A.....	169
Richlen, M.L.....	159	Stephan W.....	39
Rivera, V.....	38	Stevely, J.M.....	25
Roach, J.....	38	Stewart, T.N.....	170
Robbins, H.....	27	Still, A.....	150
Roberts, S.....	42, 67	Stoecker, D.K.....	40

Strojsova, A.	131	Wandell, H.	68
Strutton, P.	171 , 184	Wang, B.	186 , 187
Stumbaugh, M.	73	Wang, Z.	65, 106
Stumpf, R.P.	73 , 92, 107, 130, 140, 180 , 193	Wanner, A.	153
Sullivan, J.	74	Ward, J.E.	27
Sutherland, C.	172	Warner, M.E.	116
Tang, Y.Z.	91, 173	Watabayashi, G.	73
Tango, P.J.	29, 174	Watkins, S.	39, 66
Tatters, A.	175	Wecker, M.S.	101
Teegarden, G.	132, 176	Wei, L.	186, 187
Terlizzi, D.E.	96	Weisberg, S.	69
Terry, A.	41	Weisman, R.	39
Terzagian, R.	66	Wekell, J.C.	170
Tester, P.A.	25, 74 , 147, 170	Wells, M.L.	31, 45, 53, 61, 84, 86, 181, 182
Thessen, A.E.	177	Welschmeyer, N.A.	121
Thunberg, E.	46	Westrick, J.	166
Tilton, s.	52	White, Kevin	38
Tobin, E.D.	178	White, Kristy V.	188
Tomas, C.R.	152, 175	Wiggert, J.	29
Tomlinson, M.C.	73, 92 , 107, 140, 180 , 193	Wikfors, G.H.	44, 112, 142, 164, 189
Torrance, L.	133	Wilhelm, S.W.	41, 77
Tracy, K.	42, 67	Willard, E.	145
Trainer, V.L.	19, 31, 42, 45, 53, 83, 84, 86, 101, 170, 181, 182	Williams, C.	23
Tran, B.	55	Williams, J.	133
Trick, C.G.	31, 45, 53, 61, 84, 86, 181, 182	Wolf, D.	133
Truxal, L.T.	183	Wolny, J.L.	133
Turner, J.T.	87	Wood, M.	171, 184
Turonis, M.	150	Woodruff, D.	161, 185
Tweddle, J.	171, 184	Wrabel, M.L.	191
Twiner, M.J.	75	Wright, E.M.H.	188
Vanden Berghe, E.	81	Wright, J.L.C.	188
Van Dolah, F.M.	54, 59, 75	Wurch, L.L.	192
Vanderploeg, H.A.	35	Wycka, G.	132
Venrick, E.	94	Wynne, T.T.	73, 193
Vigilant, V.	49	Xu, J.	29
Villareal, T.A.	76	Yang, X.	152, 194
Vincent, M.S.	107	Younan, L.	102
Vogelbein, W.K.	141	Yranski, E.	145
Walker, E.A.	185	Zhang, H.	30, 55
Walker, H.W.	136	Zhang, Y.	81
Walker, N.D.	100	Zhao, H.	112
Waller, S.	137	Zhou, Y.	23
		Zimmermann, L.	195

LIST OF PARTICIPANTS

Nicolaus G. Adams

NOAA-Fisheries
2725 Montlake Blvd. E.
Seattle, WA 98112
Tel: (206) 860-6787
Fax: (206) 860-3335
E-mail: Nicolaus.Adams@noaa.gov

Jason E. Adolph

University of Maryland Biotechnology
Institute
Center of Marine Biotechnology
701 E. Pratt Street, Ste 236
Baltimore, MD 21202
Tel: (410) 234-8830
Fax: (410) 234-8839
E-mail: adolph@umbi.umd.edu

James W. Ammerman

Rutgers University
Institute of Marine and Coastal Sciences
71 Dudley Road
New Brunswick, NJ 08901-8521
Tel: (732) 932-6555 x339
Fax: (732) 932-8578
E-mail: ammerman@imcs.rutgers.edu

Don Anderson

Biology Department, MS #32
Woods Hole Oceanographic Institution
Woods Hole, MA 02543
Tel: (508) 289-2351
Fax: (508) 457-2027
E-mail: danderson@whoi.edu

Jon T. Anderson

Morgan State University
Estuarine Research Center
10545 Mackall Road
St. Leonard, MD 20685
Tel: (410) 586-9711
Fax: (410) 586-9705
E-mail: janderson@moac.morgan.edu

Clarissa R. Anderson

ESSIC-NOAA ESSIC/CICS
University of Maryland
College Park, MD 20742
Tel: (805) 729-0697
Fax: (301) 405-8468
E-mail: clarissa@umd.edu

Maureen E. Auro

Romberg Tiburon Center for Environmental
Studies
San Francisco State University
Tiburon, CA 94920-1205
Tel: (415) 317-5359
E-mail: maureen_auro@yahoo.com

David E. Avery

Department of Marine Sciences
University of Connecticut
Groton, CT 06340-6048
Tel: (860) 405-9066
E-mail: davery@uconn.edu

Dan L. Ayres

Washington Dept. of Fish and Wildlife
48 Devonshire Road
Montesano, WA 98563
Tel: (360) 249-4628 ext. 209
Fax: (360) 664-0689
E-mail: ayresdla@dfw.wa.gov

Lorraine C. Backer

Centers for Disease Control and Prevention
4770 Buford Highway NE
MS F-46
Chamblee, GA 30341
Tel: (770) 488-3426
Fax: (770) 488-3450
E-mail: lfb9@cdc.gov

Sibel Bargu Ates

Louisiana State University
Department of Oceanography and Coastal
Science
1235 Energy, Coast and Environment Bldg.
Baton Rouge LA 70803
Tel: (225) 578-0029
Fax: (225) 578-6326
E-mail: sbargu@lsu.edu

Benjamin F. N. Beall

University of Western Ontario
Department of Biology
Biological & Geological Sciences Building
London, Ontario
N6A 5B7 Canada
Tel: (519) 661-2111 x86470
Fax: (519) 661-3935
E-mail: bbeall2@uwo.ca

Michelle L. Berman

Santa Barbara Museum of Natural History
2559 Puesta del Sol
Santa Barbara, CA 93105
Tel: (805) 682-4711
E-mail: mberman@sbnature2.org

Debashish Bhattacharya

University of Iowa
446 Biology Building
Iowa City, IA 52242
Tel: (319) 335-1977
Fax: (319) 335-1977
E-mail: debashi-bhattacharya@uiowa.edu

Brian D. Bill

NOAA Fisheries
2725 Montlake Blvd E.
Seattle WA 98112
Tel: (206) 860-3387
E-mail: brian.d.bill@noaa.gov

Hannah Blossom

Biology Department, MS #32
Woods Hole Oceanographic Institution
Woods Hole, MA 02543
Tel: (508) 289-4908
Fax: (508) 457-2027
E-mail: blossom.hannah@gmail.com

Birgit Bolton

National Center for Environmental Health
4770 Buford Highway, NE
Building 101, Room 1162, MS F-46
Chamblee, GA 30341
Tel: (770) 488-2425
Fax: (770) 488-3450
E-mail: cyu2@cdc.gov

Jerry A. Borchert

Washington State Department of Health
111 Israel Road SE
Tumwater, WA 98501
Tel: (360) 236-3328
Fax: (360) 236-2257
E-mail: Jerry.Borchert@doh.wa.gov

David G. Borkman

Graduate School of Oceanography
University of Rhode Island
Narragansett, RI 02882
Tel: (401) 874-6686
E-mail: dborkman@gso.uri.edu

Marie-Yasmine Bottein

NOAA
219 Fort Johnson Road
Charleston, SC 29412
Tel: (843) 762-8589
Fax: (843) 762-8700
E-mail: marie-yasmine.bottein@noaa.gov

Holly Bowers

University of Maryland
Center of Marine Biotechnology
701 E. Pratt Street, Ste 236
Baltimore, MD 21202
Tel: (410) 234-8830
Fax: (410) 234-8896
E-mail: bowers@umbi.umd.edu

Gregory L. Boyer

State University of New York
College of Environmental Science
1 Forestry Drive
Syracuse, NY 13210 USA
Tel: (315) 470-6825
Fax: (315) 470-6856
E-mail: glboyer@esf.edu

Paul B. Bradley

Virginia Institute of Marine Science
P.O. Box 1346
Gloucester Point, VA 23062
Tel: (202) 482-6026
E-mail: pbradley@vims.edu

Ellen B. Braun-Howland

NYSDOH
Wadsworth Center
Empire State Plaza
Albany, NY 12201-0509
Tel: (518) 473-7925
Fax: (518) -402-5683
E-mail: bhowland@wadsworth.org

V. Monica Bricelj

National Research Council - NRC/Canada
1411 Oxford St.
Halifax, Nova Scotia
B3H 3Z1 Canada
Tel: (902) 426 8005
Fax: (902) 426 9413
E-mail: monica.bricelj@nrc-cnrc.gc.ca

Michael Brosnahan

Biology Department, MS #32
Woods Hole Oceanographic Institution
Woods Hole, MA 02543
Tel: (508) 289-3633
Fax: (508) 457-2027
E-mail: mbrosnahan@whoi.edu

Christopher W. Brown

NOAA
2207 Computer & Space Sciences Bldg. #22
University of Maryland
College Park, MD 20742
Tel: (301) 405-8031
Fax: (301) 314-1876
E-mail: christopher.w.brown@noaa.gov

Amanda M. Burson

SoMAS, Stony Brook University
239 Montauk Hwy
Southampton, NY 11703
Tel: (520) 591-7606
E-mail: Amanda.Burson@sunysb.edu

Meridith D. Byrd

Texas Parks and Wildlife Department
3000 South IH-35, Suite 320
Austin, TX 78704 USA
Tel: (512) 912-7068
Fax: (512) 707-1358
E-mail: meridith.byrd@tpwd.state.tx.us

Wendy C. Callendar

Institute of Ocean Sciences
2521 Graham Street
Victoria, BC
V8T 3Y6 Canada
Tel: (250) 386-7756
E-mail: wigginsw@pac.dfo-mpo.gc.ca

Lisa Campbell

Texas A&M University
3146 TAMU
Department of Oceanography
College Station, TX 77843
Tel: (979) 845-5706
Fax: (979) 845-6331
E-mail: lcampbell@ocean.tamu.edu

Melissa L. Carter

Scripps Institution of Oceanography
9500 Gilman Drive, Department 0227
Ritter Hall, Room 221
La Jolla, CA 92093-0227
Tel: (858) 534-6304
Fax: (858) 534-8675
E-mail: mlcarter@ucsd.edu

Gustavo Carvalho

University of Miami
RSMAS-MPO 4600
Rickenbacker Causeway
Miami, Florida 33149-1098
Tel: (305) 421-4628
Fax: (305) 421-4622
E-mail: gcarvalho@rsmas.miami.edu

Claudia Cenedese

Physical Oceanography Department, MS#21
Woods Hole Oceanographic Institution
360 Woods Hole Road
Woods Hole, MA 02543
Tel: (508) 289-2696
Fax: (508) 457-2181
E-mail: ccenedese@whoi.edu

Leo Chan

School of Biological Sciences
The University of Hong Kong
Pokfulam Road
Hong Kong SAR
E-mail: llchan@hkucc.hku.hk

Lihua Chen

University of Connecticut
Marine Sciences Building
1080 Shennecossett Road
Groton, CT 06340
Tel: (860) 514-8056
E-mail: lihua.chen@huskymail.uconn.edu

Francis J. Chrest

University of Maryland
701 East Pratt Street
Baltimore, MD 21202
Tel: (410) 952-3212
Fax: (410) 234-8896
E-mail: chrestflow@yahoo.com

Wayne Coats

Smithsonian Environmental Research
Center
P.O. Box 28
647 Contees Wharf Road
Edgewater, MD 21037
Tel: (443) 482-2271
E-mail: coatsw@si.edu

Francie E. Coblenz

UNCW
5600 Marvin K. Moss Lane
Wilmington, NC 28409
Tel: (910) 962-2387
Fax: (910) 962-2410
E-mail: coblenzfc@uncw.edu

William P. Cochlan

Romberg Tiburon Center
San Francisco State University
3152 Paradise Drive
Tiburon, CA 94920
Tel: (415) 338-3541
Fax: (415) 435-7120
E-mail: cochlan@sfsu.edu

Laurie B. Connell

University of Maine
School of Marine Sciences
Orono, ME 04469
Tel: (207) 581-2470
Fax: (207) 581-2801
E-mail: laurie.connell@umit.maine.edu

Darcie A. Couture

Maine Department of Marine Resources
194 McKown Pt Road
West Boothbay Harbor, ME 04575
Tel: (207) 633-9570
Fax: (207) 633-9579
E-mail: darcie.couture@maine.gov

Jennifer Cucksey

DOC/NOAA/NMFS
One Blackburn Drive
Gloucester, MA 01930
Tel: (978) 281-9300, ext. 6515
E-mail: jennifer.cucksey@noaa.gov

Hans G. Dam

University of Connecticut
Department of Marine Sciences
1080 Shennecossett Road
Groton, CT 06340-6098
Tel: (860) 405-9098
Fax: (860) 4059153
E-mail: hans.dam@uconn.edu

Padmanava Dash

Louisiana State University
Baton Rouge, LA 70802
Tel: (225) 772-6588
Fax: (225) 578-2520
E-mail: pdash@lsu.edu

Timothy W. Davis

SoMAS Stony Brook University
Stony Brook, NY 11794
Tel: (631) 655-5318
E-mail: timothy.walter.davis@gmail.com

Sheryl A. Day

Northwest Fisheries Science Center
2725 Montlake Boulevard East
Seattle, WA 98112
Tel: (206) 302-2410
E-mail: sheryl.day@noaa.gov

Robert W. Dickey

Division of Seafood Science and
Technology
1 Iberville Drive
P.O. Box 158
Dauphin Island, AL 36528
Tel: (251) 690-3388
Fax: (251) 694-4477
E-mail: robert.dickey@fda.hhs.gov

Kellie Dixon

Mote Marine Laboratory
1600 Ken Thompson Parkway
Sarasota, FL 34236
Tel: (941) 388-4441
E-mail: lkdxon@mote.org

Carol P. Dorsey

Alabama Department of Public Health
Mobile Division Laboratory
63 Demouy Avenue
Mobile, AL 36608
Tel: (251) 344-6049
E-mail: caroldorsey@aph.state.al.us

Quay Dortch

NOAA/NOS/NCCOS/CSCOR
10283 Green Holly Terrace
Silver Spring, MD 20902
Tel: (301) 593-6067
Fax: (301) 713-4044
E-mail: Quay.Dortch@noaa.gov

Gregory J. Doucette

NOAA/National Ocean Service
219 Fort Johnson Road
Charleston, SC 29412
Tel: (843) 762-8528
Fax: (843) 762-8700
E-mail: greg.doucette@noaa.gov

Martin P. Dowgert

U.S. Food and Drug Administration
One Montvale Avenue
Stoneham, MA 02180
Tel: (781) 596-7801
Fax: (781) 596-7894
E-mail: martin.dowgert@fda.hhs.gov

Janice Duy

University of Maine
Orono, ME 04473
E-mail: janice.duy@umit.maine.edu

Juli Dyble

NOAA
Great Lakes Environmental Research
Laboratory
2205 Commonwealth Boulevard
Ann Arbor, MI 48105
Tel: (734) 741-2444
Fax: (734) 741-2055
E-mail: juli.dyble@noaa.gov

Sonya Dyhrman

Woods Hole Oceanographic Institution
Biology Department, MS#33
Woods Hole Oceanographic Inst.
Woods Hole, MA 02543
Tel: (508) 289-3608
Fax: (508) 289-3608
E-mail: sdyhrman@whoi.edu

Deana Erdner

University of Texas
Marine Science Institute
750 Channel View Drive
Port Aransas, TX 78373
Tel: (361) 749-6719
Fax: (361) 749-6777
E-mail: derdner@utmsi.utexas.edu

Reagan M. Errera

NOAA SSMC-3 Room 11306
1315 East West Highway
Silver Spring, MD 20910
Tel: (301) 734-1067
Fax: (301) 713-1459
E-mail: Reagan.Errera@noaa.gov

Richard H. Evans

Pacific Marine Mammal Center
20612 Laguna Canyon Road
Laguna Beach, CA 92651
Tel: (949) 494-3050
Fax: (949) 494-2802
E-mail: revans@pacificmmc.org

Gary Fahnenstiel

LMFS/GLERL/NOAA
1431 Beach Street
Muskegon, MI 49441
Tel: (231) 759-7824
Fax: (231) 759-7906
E-mail: gary.fahnenstiel@noaa.gov

Kirsten Feifel

University of Washington
School of Oceanography
Box 357940
University of Washington
Seattle, WA 98195-7040
Tel: (206) 543-5098
e-mail: kfei04@u.washington.edu

Luciano Fernandes

Woods Hole Oceanographic Institution
Biology Department, MS #32
Woods Hole, MA 02543
Tel: (508) 289-4838
Fax: (508) 457-2027
E-mail: lfernandes@whoi.edu

Michael Finiguerra

University of Connecticut
Marine Sciences
1080 Shennecossett Road
Groton, CT 06340
Tel: (845) 641-3978
E-mail: michael.finiguerra@gmail.com

Spencer E. Fire

NOAA-NOS Charleston
219 Fort Johnson Road
Charleston, SC 29412
Tel: (843) 762-8574
Fax: (843) 762-8700
E-mail: spencer.fire@noaa.gov

Katie M. Fisher

NOAA 1305 East West Highway
SSMCIV N/OPS3
Silver Spring, MD 20910
Tel: (301) 713-2890 x174
Fax: (301) 713-4437
E-mail: kathleen.fisher@noaa.gov

Lora E. Fleming

University of Miami
4600 Rickenbacker Causeway
E. Grosvenor #E211
Key Biscayne, FL 33149
Tel: (305) 421-4609
Fax: (305) 421-4833
E-mail: LFleming@med.miami.edu

Michael F. Freitag

Alfred Wegener Institute
Am Handelshafen 12
Bremerhaven 27570 Germany
E-mail: Michael.freitag@awi.de

Ana C. Garcia

Louisiana State University
Baton Rouge, LA 70820
Tel: (404) 202-2666
E-mail: Agarci3@lsu.edu

Suzanne M. Garcia

University of California, Santa Cruz
528 Western Drive
Santa Cruz, CA 95060
Tel: (650) 279-2431
E-mail: smgarcia@ucsc.edu

David L. Garrison

National Science Foundation (OCE)
4201 Wilson Boulevard, Room 725
Arlington, VA 22230
Tel: (703) 292-7588
E-mail: dgarriso@nsf.gov

Cheryl Gilpin

Texas A&M University
Tel: (830) 620-0167
E-mail: seacheyl@neo.tamu.edu

José L. Giner

SUNY-ESF
307 Stadium Place
Syracuse, NY 13210
Tel: (315) 470-6895
Fax: (315) 470-6856
E-mail: jlginer@syr.edu

Pat Glibert

Univ. of Maryland Center for Environmental
Science
Horn Point Laboratory
P.O. Box 775
Cambridge, MD 21613
Tel: (410) 221-8422
Fax: (410) 221-8490
E-mail: glibert@hpl.umces.edu

Christopher J. Gobler

Stony Brook University
School of Marine and Atmospheric Science
239 Montauk Highway
Southampton, NY 11968
Tel: (631) 632-5053
E-mail: christopher.gobler@stonybrook.edu

Elena P. Gold

University of North Carolina Wilmington
Center for Marine Science
5600 Marvin K. Moss Lane
Wilmington, NC 28409
Tel: (910) 962-2366
Fax: (910) 962-2410
E-mail: emp4338@uncw.edu

Jennifer A. Goleski

Stony Brook University SoMAS
239 Montauk Highway
Southampton, NY 11968
Tel: (848) 702-5346
E-mail: jgoleski@ic.sunysb.edu

Lynn M. Grattan

University of Maryland
22 S. Greene Street, Room S12B10
Baltimore, MD 21201
Tel: (410) 328-6297
Fax: (410) 328-5874
E-mail: sroberts@som.umaryland.edu

Cynthia G. Gray

Middle Tennessee State University
800 Sawyer Bend Court
Franklin, TN 37069
Tel: (615) 478-5504
E-mail: cgg2h@mtsu.edu

Dianne I. Greenfield

MBARI
7700 Sandholdt Road
Moss Landing, CA 95039
Tel: (831) 775-1942
Fax: (831) 775-1620
E-mail: dianne@mbari.org

Cheryl L. Greengrove

University of Washington Tacoma
IAS/Environmental Science
Box 358436
Tacoma, WA 98402
Tel: (253) 692-4455
Fax: (253) 692-5718
E-mail: cgreen@u.washington.edu

Kristin Gribble

Marine Biological Laboratory
Lillie 319
7 MBL Street
Woods Hole, MA 02543
Tel: (508) 289-7194
E-mail: kgribble@mbl.edu

Jennifer Joy Griffith

University of Delaware
700 Pilottown Road
Cannon 119
Lewes, DE 19958
Tel: (302) 841-9602
E-mail: jen@udel.edu

Andrew Grimm

Marine Biological Laboratory
7 MBL Street
Woods Hole, MA 02543
Tel: (508) 289-7386
Fax: (508) 457-4727
E-mail: agrimm@mbl.edu

Jeremiah D. Hackett

University of Arizona
1014 E. Lowell Street
Tucson, AZ 85721
Tel: (520) 621-7514
Fax: (520) 621-9190
E-mail: hackettj@email.arizona.edu

Johannes A. Hagström

University of Texas Marine Science Institute
750 Channel View Drive
Port Aransas, TX 78373
Tel: (361) 749-6786
Fax: (361) 749-6777
E-mail: johannes@utmsi.utexas.edu

Sheean T. Haley

Biology Department, MS #33
Woods Hole Oceanographic Institution
Woods Hole, MA 02543
Tel: (508) 289-3683
Fax: (508) 457-2134
E-mail: shaley@whoi.edu

Emily R. Hall

Mote Marine Laboratory
1600 Ken Thompson Parkway
Sarasota, FL 34236
Tel: (941) 388-4441
E-mail: emily8@mote.org

Scott A. Hamilton

University of Maine
School of Marine Science
Orono, ME 04473
Tel: (216) 235-5854
E-mail: shamilto@gmail.com

Christina Louise Haska

University of Connecticut
Department of Marine Sciences
1080 Shennecossett Road
Groton, CT 06340
Tel: (860) 405-9089
christina.haska@uconn.edu

Theresa K. Hattenrath

Stony Brook University
239 Montauk Highway
Southampton, NY 11968
Tel: (718) 598-1129
E-mail: thattenr@ic.sunysb.edu

Kendra Hayashi

Moss Landing Marine Laboratories
8272 Moss Landing Road
Moss Landing, CA 95039
Tel: (831) 771-4125
Fax: (831) 633-7263
E-mail: khayashi@mml.calstate.edu

Allison J. Haywood

FIO/FFWCC Fish and Wildlife Research
Institute
E-mail: ajhaywood@xnet.co.nz

Hélène Hégarret

University of Connecticut
Department of Marine Sciences
1080 Shennecossett Road
Groton, CT 06340
Tel: (203) 882-6525
Fax: (203) 882-6517
E-mail: helene.hegarret@gmail.com

Lauren Heinen

Biology Department, MS #32
Woods Hole Oceanographic Institution
Woods Hole, MA 02543
Tel: (508) 289-4908
Fax: (508) 457-2027
E-mail: heinen.l@neu.edu

Darren W. Henrichs

Texas A&M University
Eller O&M Building, Room 1204
MS 3146 - Dept of Oceanography
College Station, Texas 77843-3146
Tel: (979) 845-5738
Fax: (979) 845-6331
E-mail: dhenrichs@bio.tamu.edu

Barbara M. Hickey

University of Washington
School of Oceanography
Box 357940
Seattle, WA 98195-7940
Tel: (360) 825-3911
E-mail: bhickey@u.washington.edu

Porter Hoagland

Marine Policy Center, MS #41
Woods Hole Oceanographic Institution
Woods Hole, MA 02543
Tel: (508) 289-2867
Fax: (508) 457-2184
E-mail: phoagland@whoi.edu

Rita A. Horner

University of Washington
School of Oceanography
Box 357940
Seattle, WA 98195-7940
Tel: (206) 543-8599
Fax: (206) 543-0275
E-mail: rita@ocean.washington.edu

Amber M. Hotto

SUNY-ESF
1 Forestry Drive
121 Jahn Laboratory
Syracuse, NY 13210
Tel: (315) 569-1749
Fax: (315) 470-6856
E-mail: ahotto@syr.edu

Katherine A. Hubbard

University of Washington
School of Oceanography
Box 357940
Seattle, WA 98195
Tel: (206) 349-5613
E-mail: hubbard@ocean.washington.edu

Matthew V. Hunter

Oregon Department of Fish and Wildlife
2001 Marine Drive, Room 120
Astoria, OR 97103
Tel: (503) 325-2462
Fax: (503) 325-8227
E-mail: matthew.v.hunter@state.or.us

James B. Hyde

New York Department of Health
547 River Street
Troy, NY 12180-2216
Tel: (518) 402-7711
Fax: (518) 402-7599
E-mail: jbh01@health.state.ny.us

Rachel Wisniewski Jakuba

AAAS Policy Fellow / EPA
Tel: (617) 510-9694
E-mail: Jakuba.Rachel@epa.gov

Rosalind J. Jester

University of California, Santa Cruz
Ocean Science Department
1156 High Street
Santa Cruz, CA 95064
Tel: (831) 459-2948
Fax: (831) 459-4882
E-mail: rosalind.jester@gmail.com

Desmond Johns

UMCES/ Horn Point Laboratory
P.O. Box 775
2020 Horns Point Road
Cambridge, MD 21613
Tel: (410) 221-8318
Fax: (410) 221-8190
E-mail: djohns@hpl.umces.edu

Andrew R. Juhl

Lamont-Doherty Earth Observatory
Columbia University
61 Route 9W
Palisades, NY 10964
Tel: (845) 365-8837
E-mail: andyjuhl@ldeo.columbia.edu

Bruce A. Keafer

Woods Hole Oceanographic Institution
Biology Department, MS #32
Woods Hole, MA 02543
Tel: (508) 289-2509
Fax: (508) 457-2027
E-mail: bkeafer@whoi.edu

Barbara Kirkpatrick

Mote Marine Laboratory
1600 Ken Thompson Parkway
Sarasota, FL 34236
Tel: (941) 388-4441
Fax: (941) 388-4312
E-mail: bkirkpat@mote.org

Gary J. Kirkpatrick

Mote Marine Laboratory
1600 Ken Thompson Parkway
Sarasota, FL 34236
Tel: (941) 388-4441
Fax: (941) 388-4312
E-mail: gkirkpat@mote.org

Judy Kleindinst

Biology Department, MS #32
Woods Hole Oceanographic Institution
Woods Hole, MA 02543
Tel: (508) 289-2745
Fax: (508) 457-2027
E-mail: jkleindinst@whoi.edu

Nicole Kohmescher

State of Oklahoma Water Resources Board
3800 N. Classen Boulevard
Oklahoma City, OK 73118
Tel: (405) 530-8800
Fax: (405) 530-8900
E-mail: ndkohmescher@owrb.ok.gov

Raphael M. Kudela

University of California Santa Cruz
Ocean Sciences Department
1156 High Street
Santa Cruz, CA 95064
Tel: (831) 459-3290
Fax: (831) 459-4882
E-mail: kudela@ucsc.edu

David Kulis

Biology Department, MS #32
Woods Hole Oceanographic Institution
Woods Hole, MA 02543
Tel: (508) 289-2859
Fax: (508) 457-2027
E-mail: dkulis@whoi.edu

Edward Laine

Bowdoin College
6800 College Station
Geology Department
Brunswick, ME 04011-3404
Tel: (207) 725-3621
Fax: (207) 798-7037
E-mail: edlaine@bowdoin.edu

Jan H. Landsberg

Florida Fish and Wildlife Conservation
Commission
100 Eighth Avenue SE
St. Petersburg FL 33701
Tel: (727) 896-8626
E-mail: jan.landsberg@myfwc.com

Jenny Q. Lane

University of California Santa Cruz
1156 High Street
Ocean Sciences Department
Santa Cruz, CA 95064
Tel: (831) 460-9875
E-mail: jqlane@gmail.com

Heidi W. Langer Atkinson

University at Albany
School of Public Health
245 County Route 312
Westerlo, NY 12193
Tel: (518) 474-0255
E-mail: hw101@health.state.ny.us

Gregg Langlois

California Department of Public Health
850 Marina Bay Parkway, G165
Richmond, CA 94804
Tel: (510) 412-4635
E-mail: Gregg.Langlois@cdph.ca.gov

Andrew D. Lasiter

Middle Tennessee State University
3833 Edwards Ave
Nashville, TN 37216
Tel: (615) 717-5660
E-mail: adl2s@mtsu.edu

Jungju Lee

The Ohio State University
422 Hitchcock Hall
2070 Neil Avenue
Columbus, OH 43210
Tel: (614) 292-7340
Fax: (614) 292-3780
E-mail: lee.2374@osu.edu

Kathi Ann Lefebvre

NOAA Fisheries/NWFSC
2725 Montlake Boulevard East
Seattle, WA 98112
Tel: (206) 302-2454
Fax: (206) 860-3335
E-mail: Kathi.Lefebvre@noaa.gov

Peggy W. Lehman

California Department of Water Resources
Division of Environmental Services
901 P Street
Sacramento, CA 95814
Tel: (916) 651-9546
E-mail: plehman@water.ca.gov

Danelle K. Lekan

University of North Carolina Wilmington
5600 Marvin K. Moss Lane
Wilmington, NC 28409
Tel: (910) 620-2928
Fax: (910) 962-2410
E-mail: dkl5447@uncw.edu

Rebecca V. LePrell

CDC
National Center for Environmental Health
4770 Buford Highway NE
MS F-46
Atlanta, GA 30341
Tel: (770) 488-3418
Fax: (770) 488-3450
E-mail: RLePrell@cdc.gov

Evelyn J. Lessard

University of Washington
Box 357940
Seattle, WA 98195
Tel: (206) 543-8795
E-mail: elessard@u.washington.edu

Alan Lewitus

NOAA CSCOR
1305 East West Highway, Room 8220
Silver Spring, MD 20910
Tel: (301) 713-3338 x178
Fax: (301) 713-4044
E-mail: Alan.Lewitus@noaa.gov

Katie M. Libera

Biology Department, MS #32
Woods Hole Oceanographic Institution
Woods Hole, MA 02543
Tel: (508) 289-2948
Fax: (508) 457-2027
E-mail: klibera@whoi.edu

Kristy B. Lidie

Center of Marine Biotechnology
701 Pratt Street
Baltimore, MD 21202
Tel: (410) 234-8830
E-mail: kristy.lidie@noaa.gov

Senjie Lin

Department of Marine Sciences
University of Connecticut
1080 Shennecossett Road
Groton, CT 06340
Tel: (860) 405-9168
Fax: (860) 405-9153
E-mail: senjie.lin@uconn.edu

R. Wayne Litaker

National Ocean Service, NOAA
101 Pivers Island Road
Beaufort, NC 28516
Tel: (252) 728-8791
Fax: (252) 728-8784
E-mail: Wayne.Litaker@noaa.gov

Cary B. Lopez

NOAA National Ocean Service
Center for Sponsored Coastal Ocean
Research
1305 E West Hwy N/SCI2
SSMC 4 RM 8217
Silver Spring, MD 20910
Tel: (301) 713-3338 x170
Fax: (301) 713-4044
E-mail: cary.lopez@noaa.gov

Rebecca Love

NOAA Coastal Services Center
2234 South Hobson Avenue
Charleston, SC 29405
Tel: (843) 740-1169
Fax: (843) 740-1224
E-mail: rebecca.love@noaa.gov

Vincent J. Lovko

Virginia Institute of Marine Science
Gloucester Point, VA 23062
Tel: (804) 684-7738
E-mail: vlovko@vims.edu

Danielle Luttenberg Meitiv

NOAA CSCOR
1305 East-West Hwy N/SC12
Silver Spring MD 20910
Tel: (301) 713-3338 x155
Fax: (301) 713-4044
E-mail: danielle.meitiv@noaa.gov

Trina N. Mackie

University of California
Berkeley & Klamath BGA Work Group
Berkeley, CA 94702
Tel: (510) 643-5141
Fax: (510) 642-5815
E-mail: tmackie@berkeley.edu

Luiz L. Mafra

National Research Council Canada
1411 Oxford Street
Halifax, Nova Scotia
B3H 3Z1 Canada
Tel: (902) 426-2239
Fax: (902) 426-9658
E-mail: luiz.mafra@nrc-cnrc.gc.ca

Robert Magnien

Department of Commerce/NOAA
1305 East West Highway
Silver Spring, MD 20910
Tel: (301) 713-3338
Fax: (301) 713-4044
E-mail: Rob.Magnien@noaa.gov

Ulrika Malone

DOC/NOAA/NMFS
One Blackburn Drive
Gloucester, MA 01930
Tel: (978) 281-9300 X 6511
E-mail: ulrika.malone@noaa.gov

Alicia R. Mangum

Delaware State University
6190 Steffland Drive
Seaford, DE 19973
Tel: (302) 448-6435
E-mail: alicia_revis@yahoo.com

Lucie Maranda

University of Rhode Island
Graduate School of Oceanography
Narragansett, RI 02882
Tel: (401) 874-6216
Fax: (401) 874-6240
E-mail: lmaranda@gso.uri.edu

Kelli L. Margot

Medical University of South Carolina
CRI Rm. 304D
173 Ashley Avenue
Charleston, SC 29425
Tel: (419) 566-4282
Fax: (843) 792-1844
E-mail: margot@musc.edu

Julie Elizabeth Masura

University of Washington – Tacoma
IAS/Environmental Science
Box 358436
1900 Commerce Street
Tacoma, WA 98402
Tel: (253) 692-4450
Fax: (253) 692-5718
E-mail: jmasura@u.washington.edu

Fernanda Mazzillo

University of California Santa Cruz
Santa Cruz, CA 95060
Tel: (858) 405-6684
E-mail: mazzillo@gmail.com

Dennis J. McGillicuddy

Applied Ocean Physics and Engineering
MS #11
Woods Hole Oceanographic Institution
Woods Hole, MA 02543
Tel: (508) 289-2638
Fax: (508) 457-2194
E-mail: dmcgillicuddy@whoi.edu

Tim McLean

University of Southern Mississippi
Department of Biological Sciences
118 College Drive, #5018
Hattiesburg, MS 39406
Tel: (601) 266-4753
E-mail: timothy.mclean@usm.edu

Peter E. Miller

University of California, Santa Cruz
Institute of Marine Sciences
1156 High Street
Santa Cruz, CA 95064
Tel: (831) 459-5005
Fax: (831) 459-4882
E-mail: pemiller@ucsc.edu

David F. Millie

Florida Institute of Oceanography
4645 Stone Ridge Trail
Sarasota, FL 34232
Tel: (941) 544-7926
E-mail: dmillie@comcast.net

Lilibeth N. Miranda

University of Connecticut
Dept of Marine Sciences
1080 Shennecossett Road
Groton, CT 06340
Tel: (860) 405-9089
E-mail: lilibeth.miranda@uconn.edu

Emily A. Monroe

NOAA/CCEHBR
219 Fort Johnson Road
Charleston, SC 29412
Tel: (843) 762-8593
Fax: (843) 762-8700
E-mail: monroe@musc.edu

Ben Mooney

University of Maryland
Center of Marine Biotechnology
701 E. Pratt Street, Suite 236
Baltimore, MD 21202
Tel: (410) 234-8830
Fax: (410) 234-8896
E-mail: mooney@umbi.umd.edu

Lindsay K. Moore

Medical College of Georgia
Dept. of Medical Illustration
1120 15th Street, CJ1101
Augusta, GA 30912-0300
Tel: (631) 807-9691
E-mail: lindsay.koza.moore@gmail.com

Steve L. Morton

NOAA/NOS
331 Fort Johnson Road
Charleston, SC 29412
Tel: (843) 762-8857
E-mail: steve.morton@noaa.gov

Ahmed Moustafa

University of Iowa
456 Biology Building
Iowa City, Iowa 52246
Tel: (319) 335-1214
Fax: (319) 335-1069
E-mail: ahmed-moustafa@uiowa.edu

Christine L. Muir

RTC, SFSU and Woodside Priory School
Woodside Priory School
302 Portola Road
Portola Valley, CA 94028
Tel: (415) 272-0453
Fax: (650) 851-2839
E-mail: cmuir@woodsidepriory.com

Tracey L. Myers

Georgia Institute of Technology
311 Ferst Drive
ES&T Building – Biology
Atlanta, GA 30332
Tel: (404) 385-4437
Fax: (404) 385-4440
E-mail: tm192@mail.gatech.edu

Jerome Naar

Center for Marine Science – UNCW
5600 Marvin K Moss Lane
Wilmington, NC 28409
Tel: (910) 962 2367
Fax: (910) 962 2410
E-mail: naarj@uncw.edu

Amy W. Nau

Brown University
Box 6106
Providence, RI 02912
Tel: (919) 414-6155
E-mail: Amy_Nau@brown.edu

Kate Nierenberg

Mote Marine Laboratory
1600 Ken Thompson Parkway
Sarasota, FL 34236
Tel: (941) 388-4441
Fax: (941) 388-4312
E-mail: knierenberg@mote.org

Ron K. Nilsen

Bellarmino Preparatory School
2300 South Washington Street
Tacoma, WA 98405
E-mail: nilsenr@bellarminerep.org

Lucie Novoveska

Dauphin Island Sea Laboratory
101 Bienville Boulevard
Dauphin Island, AL 36528
Tel: (251) 861-7502
E-mail: lnovoveska@disl.org

Anthony M. Odell

University of Washington
ONRC, ORHAB
14 North Shore Place
Hoquiam, WA 98550
Tel: (360) 533-0384
E-mail: odellamo@u.washington.edu

Michael L. Parsons

Florida Gulf Coast University
Department of Marine and Ecological
Science
10501 FGCU Blvd S
Fort Myers, FL 33965
Tel: (239) 590-7526
Fax: (239) 590-7200
E-mail: mparsons@fgcu.edu

Chris Pettit

Mote Marine Laboratory
850 South Tamiami Trail, #204
Sarasota, FL 34236
Tel: (941) 284-6733
E-mail: chrispettit@hotmail.com

Lisa D. Pickell

University of Maine
Tel: (207) 563-3146
Fax: (207) 563-3119
E-mail: babydill_@hotmail.com

Richard H. Pierce

Mote Marine Laboratory
1600 Ken Thompson Parkway
Sarasota, FL 34236
Tel: (941) 388-4441 x 342
Fax: (941) 388-4312
E-mail: rich@mote.org

Allen Place

University of Maryland
Center of Marine Biotechnology
701 E. Pratt Street, Ste 236
Baltimore, MD 21202
Tel: (410) 234-8828
Fax: (410) 234-8896
E-mail: place@umbi.umd.edu

Steven M. Plakas

U.S. Food and Drug Administration
1 Iberville Drive
P.O. Box 158
Dauphin Island, AL 36528
Tel: (251) 690-3403
Fax: (251) 694-4477
E-mail: Steven.Plakas@fda.hhs.gov

F. Gerald Plumley

Bermuda Institute of Ocean Sciences
17 Biological Lane
St. George's
GE01 Bermuda
Tel: (441) 297-1880
Fax: (441) 297-8143
E-mail: gerry.plumley@bios.edu

Lara Yael Polansky

Marine Policy Center, MS #41
Woods Hole Oceanographic Institution
Woods Hole, MA 02543
Tel: (630) 697-0396
E-mail: Larayael@aol.com

Mark A. Poli

USAMRIID
Fort Detrick, MD 21702-5011
Tel: (301)-619-4801
Fax: (301)-619-2348
E-mail: mark.poli@det.amedd.army.mil

James R. Postel

University of Washington
School of Oceanography
Box 357940
Seattle, WA 98195-7940
Tel: (206) 543-4485
Fax: (206) 543-0275
E-mail: jrpostel@u.washington.edu

Emily K. Prince

Georgia Institute of Technology
311 Ferst Drive
Atlanta, GA 30332
Tel: (404) 385-4437
Fax: (404) 385-4440
E-mail: gte982j@mail.gatech.edu

Regina L. Radan

San Francisco State University/RTC
708 Bay Road
Mill Valley, CA 94941
Tel: (415) 516-0285
E-mail: rlradan@hotmail.com

Faisal F. Y. Radwan

NOAA/National Ocean Service
219 Fort Johnson Road
Charleston, SC 29412
Tel: (843) 762-8637
Fax: (843) 762-8700
E-mail: faisal.radwan@noaa.gov

John S. Ramsdell

NOAA/National Ocean Service
219 Fort Johnson Road
Charleston, SC 29412
Tel: (843) 762-8637
Fax: (843) 762-8700
E-mail: faisal.radwan@noaa.gov

Andrew Reich

Aquatic Toxins Program
Florida Dept of Health
4052 Bald Cypress Way, Bin A08
Tallahassee, FL 32399-1712
Tel: (850) 245-4444 x 2295
E-mail: andy_reich@doh.state.fl.us

Tammi L. Richardson

University of South Carolina
Department of Biological Sciences
Sumter Street
Columbia, SC 29208
Tel: (803) 777-2269
E-mail: richardson@biol.sc.edu

Mindy Richlen

Biology Department, MS #32
Woods Hole Oceanographic Institution
Woods Hole, MA 02543
Tel: (508) 289-2552
Fax: (508) 457-2027
E-mail: mrichlen@bu.edu

Sparkle M. Roberts

University of Maryland
22 S. Greene St., Rm. S12B10
Baltimore, MD 21201
Tel: (410) 328-6297
Fax: (410) 328-5874
E-mail: sroberts@som.umaryland.edu

Juliette N. Rooney-Varga

University of Massachusetts Lowell
Biological Sciences Olsen Hall
198 Riverside Street
Lowell, MA 01854
Tel: (978) 934-4715
Fax: (978) 934-3044
E-mail: juliette_rooneyvarga@uml.edu

Jessica Rowe

University of Maryland
22 S. Greene St., Rm. S12B10
Baltimore, MD 21201
Tel: (410) 328-6297
Fax: (410) 328-5874
E-mail: jrowe@som.umaryland.edu

Lauren R. Salvitti

University of Delaware
16003-2 Bowman Drive
Lewes, DE 19958
Tel: (610) 405-3959
E-mail: LSalvitt@udel.edu

Orlando Sarnelle

Michigan State University
Department of Fisheries and Wildlife
Michigan State University
East Lansing, MI 48824
Tel: (517) 353-4819
Fax: (517) 432-1699
E-mail: sarnelle@mu.edu

Clifford W. Scherer

Department of Communication
Social and Behavioral Research Unit
Cornell University
Ithaca, NY 14853
Tel: (607) 255-7498
Fax: (607) 254-1322
E-mail: CWS4@Cornell.edu

Astrid Schnetzer

University of Southern California
3616 Trousdale Parkway, AHF 301
Los Angeles CA 90089
Tel: (213) 821-1800
Fax: (213) 740-8123
E-mail: astrids@usc.edu

Donald M. Schoener

University of Connecticut Marine Sciences
1080 Shennecossett Road
Groton, CT 06340
Tel: (860) 405-9099
E-mail: Donald.Schoener@uconn.edu

Chris A. Scholin

Monterey Bay Aquarium Research Institute
7700 Sandholdt Road
Moss Landing CA 95039
Tel: (831) 775-1779
Fax: (831) 775-1620
E-mail: scholin@mbari.org

Irvin Schultz

Battelle PNNL-Marine Sciences Laboratory
1529 West Sequim Bay Road
Sequim, WA 98352
Tel: (360) 681-4566
Fax: (360) 681-3681
E-mail: irv.schultz@pnl.gov

Ervin Joe Schumacker

Quinault Indian Nation
P.O. Box 424
Taholah, WA 98587
Tel: (360) 276-8215 x 327
Fax: (360) 276-4682
E-mail: jschumacker@quinault.org

Kevin G. Sellner

Chesapeake Research Consortium
645 Contees Wharf Road
Edgewater, MD 21037
Tel: (410) 798-1283
Fax: (410) 798-0816
E-mail: sellnerk@si.edu

Stella G. Sellner

1422 Snug Harbor Road
Shady Side, MD 20764
Tel: (410) 867-7114
E-mail: sgsellner@moac.morgan.edu

Christina Danelle Senft

University of Connecticut, Avery Point
1080 Shennecossett Road
Marine Science Building
Groton, CT 06340
Tel: (860) 405-9152
Fax: (860) 405-9153
E-mail: christina.senft@uconn.edu

Mario R. Sengco

Smithsonian Environmental Research
Center
PO Box 28
647 Contees Wharf Road
Edgewater, MD 21037
Tel: (443) 482-2362
Fax: (443) 482-2380
E-mail: sengcom@si.edu

Valerie I. Shearn-Bochsler

USGS
National Wildlife Health Center
6006 Schroeder Road
Madison, WI 53711
Tel: (608) 270-2457
Fax: (608) 270-2415
E-mail: vbochsler@usgs.gov

Mina Shehee

North Carolina Dept Health & Human
Service
Raleigh, NC 27699
Tel: (919) 707-5920
E-mail: mina.shehee@ncmail.ne

Sandra E. Shumway

University of Connecticut
Dept of Marine Sciences
1080 Shennecossett Road
Groton, CT 06340
Tel: (860) 405-9282
Fax: (860) 405-9153
E-mail: sandra.shumway@uconn.edu

Mary Wilcox Silver

University of California, Santa Cruz
1156 High Street
Santa Cruz, CA 95064
Tel: (831) 459-2908
Fax: (831) 459-4882
E-mail: msilver@ucsc.edu

Geoffrey A. Sinclair

North Carolina State University
1800 Faucette Drive
Jordan Hall Room 1125
Raleigh, NC 27695
Tel: (919) 515-3396
E-mail: gasincla@ncsu.edu

James L. Sinclair

U.S. EPA
26 W. Martin Luther King Drive (MS-140)
Cincinnati, OH 45268
Tel: (513) 569-7970
Fax: (513) 569-7191
E-mail: sinclair.james@epa.gov

Alison Sirois

Maine Department of Marine Resources
194 McKown Point Road
West Boothbay Harbor, Maine 04575
Tel: (207) 633-9401
Fax: (207) 633-9579
E-mail: alison.sirois@maine.gov

Hayley M. Skelton

NCSU
Center for Applied Aquatic Ecology
620 Hutton Street, Suite 104
Campus Box 7510
Raleigh, NC 27606
Tel: (919) 515-3421
Fax: (919) 513-3194
E-mail: hmskelto@unity.ncsu.edu

Theodore J. Smayda

University of Rhode Island
Graduate School of Oceanography
Narragansett, RI 02882-1197
Tel: (401) 874-6171
Fax: (401) 874-6682
E-mail: tsmayda@gso.uri.edu

Juliette L. Smith

SUNY-ESF
Environmental and Forest Biology
1 Forestry Drive
Syracuse, NY 13210
Tel: (315) 470-6844
Fax: (315) 470-6934
E-mail: jlsmith19@syr.edu

G. Jason Smith

Moss Landing Marine Laboratories / ACT
8272 Moss Landing Rd
Moss Landing, CA 95039
Tel: (831) 771-4126
Fax: (831) 633-7263
E-mail: jsmith@mlml.calstate.edu

Beth A. Stauffer

University of Southern California
3616 Trousdale Parkway, AHF 301
Los Angeles, CA 90089-0371
Tel: (213) 821-2123
Fax: (213) 740-8123
E-mail: stauffer@usc.edu

Thomas N. Stewart

Mercury Science Inc.
2801 Blue Ridge Road, Suite G70
Raleigh, NC 27607
Tel: (866) 861-5836
Fax: (206) 202-2387
E-mail: tom@mercuryscience.com

Pete Strutton

Oregon State University
104 COAS Admin Building
Corvallis, OR 97330
Tel: (541) 737-2065
Fax: (541) 737-2064
E-mail: strutton@coas.oregonstate.edu

Richard P. Stumpf

NOAA National Ocean Service
1305 East-West Highway
Room 9115, Code N/SCI
Silver Spring, MD 20910
Tel: (301) 713-3028 x173
Fax: (301) 713-4384
E-mail: richard.stumpf@noaa.gov

Marc Suddleson

Department of Commerce/NOAA
1305 East West Highway
Silver Spring, MD 20910
Tel: (301) 713-3338
Fax: (301) 713-4044
E-mail: Marc.Suddleson@noaa.gov

Cristy M. Sutherland

University of California, Santa Cruz
65 Grandview Street Unit F
Santa Cruz, CA 95060-3082
Tel: (831-) 459-3469
Fax: (831) 459-3520
E-mail: cristym@ucsc.edu

Daniel Terlizzi

University of Maryland
701 E. Pratt Street
Baltimore, MD 21202
Tel: (410) 234-8837
Fax: (410) 234-8896
E-mail: dterlizz@umd.edu

Ying Zhong Tang

Stony Brook University
Marine Sciences Research Center
239 Montauk Highway
Southampton, NY 11968
Tel: (757) 201-1977
Fax: (631) 632-5070
E-mail: yittang@notes.cc.sunysb.edu

Peter J. Tango

U.S. Geological Survey
Chesapeake Bay Program
410 Severn Ave. Suite 109
Annapolis, MD 21403
Tel: (410) 267-9875
E-mail: ptango@chesapeakebay.net

Avery O. Tatters

University of North Carolina Wilmington
Center for Marine Science
5600 Marvin K. Moss Lane
Wilmington, NC 28409
Tel: (910) 962-2370
E-mail: aot2642@uncw.edu

Gregory J. Teegarden

Saint Joseph's College
278 Whites Bridge Road
Standish, ME 04084
Tel: (207) 893-7979
E-mail: gteegarden@sjcme.edu

Daniel E. Terlizzi

University of Maryland
701 E. Pratt Street
Baltimore, MD 21202
Tel: (410) 234-8837
Fax: (410) 234-8896
E-mail: dterlizz@umd.edu

Patricia A. Tester

National Ocean Service, NOAA
101 Pivers Island Road
Beaufort, NC 28516
Tel: (252) 728-8792
Fax: (252) 728-8784
E-mail: pat.testler@noaa.gov

Anne E. Thessen

Horn Point Laboratory
P.O. Box 775
Cambridge, MD 21613
Tel: (410) 221-8253
Fax: (410) 221-8290
E-mail: athessen@hpl.umces.edu

Elizabeth D. Tobin

University of Washington
Marine Sciences Building, Room G
1501 NE Boat St
Seattle, WA 98195
Tel: (206) 543-5098
E-mail: etobin@u.washington.edu

Carmelo R. Tomas

University of North Carolina Wilmington
Center for Marine Science
5600 Marvin K. Moss Lane
Wilmington, NC 28409
Tel: (910) 962-2385
Fax: (910) 962-2340
E-mail: tomasc@uncw.edu

Michelle C. Tomlinson

NOAA
Center for Coastal Monitoring and
Assessment
1305 East-West Highway
Silver Spring, MD 20910
Tel: (301) 713-3028 x225
Fax: (301) 713-4338
E-mail: Michelle.Tomlinson@noaa.gov

Jessie Tong

Biology Department, MS #32
Woods Hole Oceanographic Institution
Woods Hole, MA 02543
Tel: (508) 289-4838
Fax: (508) 457-2027
E-mail: mtong@whoi.edu

Dwight Trueblood

CICEET
University of New Hampshire
Gregg Hall, Suite 130
350 Colovos Road
Durham, NH 03824-3534
Tel: (603) 862-3580
Fax: (603) 862-2940
E-mail: Dwight.Trueblood@noaa.gov

Laura T. Truxal

UNCW
Center for Marine Science
5600 Marvin K. Moss Lane
Wilmington, NC 28409
Tel: (412) 554-5093
E-mail: ltt5269@uncw.edu

Jefferson T. Turner

University of Massachusetts Dartmouth
Biology Department
285 Old Westport Road
North Dartmouth, MA 02747
Tel: (508) 999-8229
Fax: (508) 999-8196
E-mail: jturner@umassd.edu

Jacqueline F. Tweddle

Oregon State University
104 COAS Admin. Building
Corvallis, OR 97331
Tel: (541) 737-6560
Fax: (541) 737-2064
E-mail: jtweddle@coas.oregonstate.edu

Mike J. Twiner

Marine Biotoxins Program, NOAA
219 Fort Johnson Road
Charleston, SC 29412
Tel: (843) 762-8649
E-mail: mike.twiner@noaa.gov

Cristina Urizar

NOAA 1305 East West Highway
SSMCIV N/OPS 3
Silver Spring, MD 20910
Tel: (301) 713-2890 x114
Fax: (301) 713-4437
E-mail: Cristina.Urizar@noaa.gov

Frances M. Van Dolah

NOAA 219 Fort Johnson Road
Charleston, SC 29412
Tel: (843) 762-8529
Fax: (843) 762-8700
E-mail: fran.vandolah@noaa.gov

Tracy A. Villareal

The University of Texas at Austin
Marine Science Institute
750 Channel View Drive
Port Aransas, TX 78373
Tel: (361) 749-6832
Fax: (361) 749-6777
E-mail: tracy@utmsi.utexas.edu

Wolfgang K. Vogelbein

Virginia Institute of Marine Science
Route 1208
Gloucester Point, VA 23062
Tel: (804) 684-7261
Fax: (804) 684-7186
E-mail: wolf@vims.edu

Elyse A. Walker

University of South Carolina
Marine Science Program
EWS 603, 712 Main Street
Columbia, SC 29208
Tel: (360) 731-7296
Fax: (803) 777-3922
E-mail: walkerea@mailbox.sc.edu

Harold W. Walker

Ohio State University
Department of Civil
& Environmental Engineering
470 Hitchcock Hall
2070 Neil Avenue
Columbus, OH 43210
Tel: (614) 292-8263
E-mail: walker.455@osu.edu

Da-Zhi Wang

Xiamen University
Xiamen
Fujian 361005 China
Tel: 86-592-2186016
Fax: 86-592-2180655
E-mail: dzwang@xmu.edu.cn

Bin Wang

New Jersey Institute of Technology
Newark, NJ 07102
Tel: (973) 596-6077
E-mail: bw36@njit.edu

Liping Wei

New Jersey Institute of Technology
138 Warren Street
Newark, NJ 07102
Tel: (973) 596-5389
Fax: (973) 596-3586
E-mail: wei@adm.njit.edu

Mark L. Wells

University of Maine
School of Marine Sciences
Orono, ME 04469
Tel: (207) 581-4322
Fax: (207) 581-4388
E-mail: mlwells@maine.edu

Kristy Van Etten White

University of North Carolina at Wilmington
Center for Marine Science
5600 Marvin K. Moss Lane
Wilmington, NC 28409
Tel: (910) 962-2398
Fax: (910) 962-2410

Dave Whittaker

Massachusetts Division of Marine Fisheries
1213 Purchase Street
New Bedford, MA 02740
Centrex #: (617)727-0349
Tel: (508) 990-2860 x 126
Fax: (508) 990-0449
E-mail: david.whittaker@state.ma.us

Gary H. Wikfors

NOAA Fisheries Service
212 Rogers Avenue
Milford, CT 06460
Tel: (203) 882-6525
Fax: (203) 882-6517
E-mail: Gary.Wikfors@noaa.gov

Steven W. Wilhelm

The University of Tennessee
1414 West Cumberland Avenue
Knoxville, TN 37996
Tel: (865) 974-0665
Fax: (865) 974-4007
E-mail: wilhelm@utk.edu

Season Wong

Lynntech, Inc.
7610 Eastmark Drive
College Station, TX 77840
Tel: (979) 693-0017
E-mail: seasonwong@yahoo.com

Michele L. Wrabel

University of Washington
School of Oceanography
Box 357940
Seattle, WA 98195-7940
Tel: (206) 897-1871
Fax: (206) 685-6651
E-mail: mlw22@u.washington.edu

Louie L. Wurch

Biology Department, MS #33
Woods Hole Oceanographic Institution
Woods Hole, MA 02543
Tel: (508) 289-3565
E-mail: lwurch@whoi.edu

Xingye Yang

University of North Carolina at Wilmington
Center for Marine Science
5600 Marvin K. Moss Lane
Wilmington, NC 28409
Tel: (910) 962-2368
E-mail: yangx@uncw.edu

Lawrence Y. Younan

Turner Designs, Inc.
Sunnyvale, CA 94087
Tel: (209) 605-3484
E-mail: lyounan@turnerdesigns.com

Leigh Zimmermann

Solutions To Avoid Red Tide (START)
1001 3rd Ave. W, Ste. 500
Bradenton, FL 34205
Tel: (941) 747-0324
Fax: (941) 747-0143
E-mail: leigh@start1.com