Organic matter cycling in anoxic (no oxygen) Marine sediments



There are many examples where oxygen becomes exhausted in sediments

How is organic matter oxidized without oxygen?

What is the origin of petroleum source rocks?

What biological communities can exist in anoxic sediments and how do they function?

How does biogeochemical cycling in anoxic marine sediments affect global C:N:S cycles? Dysaerobic and anoxic environments are devoid of higher heterotrophic organisms. Biogeochemical cycling, Including photosynthesis and C respiration are carried out by bacteria



Bacterial communities are much more metabolically specialized than higher heterotrophs. The communities are strictly segregated according to the environmental conditions. This leads to biogeochemical zoning of microbes and chemical species (nutrients, carbon dioxide, methane, etc.).



These communities are widespread in the Great Sippewissett Marsh in West Falmouth.

Oxidation of organic matter in marine sediments

Re	eactior	1	$\Delta E(KJ/mole)$	Capacity (mmoles/L sed)
O ₂	\rightarrow	CO ₂	-475	0.85
NO3⁻	\rightarrow	$N_2 + CO_2$	-448	0.05
Mn(IV) ->	Mn (II)	-349	2-22
Fe(III)	\rightarrow	Fe (II)	-114	14-28
SO4 ²⁻	\rightarrow	S ²⁻	-77	56
CO ₂	\rightarrow	CH ₄	-58	

Methanogenesis

Organic Matter

proteins 50% C, 80% N (soluble/structural) polysaccharides 20-30% C, 10% N (soluble/structural) nucleaic acids 10% C, 5% N. 50% P lipids 10% C, few % N

oxic degradation nitrate reduction

soluble constituents CO₂, NO₃⁻, HPO₃⁻

proteins (structural), peptides polysaccharides (structural), oligosaccharides, sugars, nucleic acids, lipids

Consumption of acetate by sulfate reducing and methanogenic bacteria

Acetate = CH_3COOH Volatile Fatty Acid (VFA) = CH_3RCOOH Competition for acetate between Desulfobacter postgatei and Methanosarcina bakerii

Double reciprocal plot of rate vs Acetate concentration

Competition for acetate between Desulfobacter postgatei and Methanosarcina bakerii

Competition for hydrogen between sulfate reducing and methanogenic bacteria

Changes in pore water concentrations during a 114 day incubation

Concentration

Anaerobic degradation of pectin

Degradation of pectin in Knaack Lake sediments

Sulfate Present

Sulfate Absent

Chemical thermodynamics and pure culture studies of anaerobic micro-organisms suggest that the oxidizing capacity of sediments is >>> than [O2]. We predict that sulfate and ultimately carbon dioxide will be used as electron acceptors during organic matter oxidation. We also predict that sulfate reduction will occur before methanogensis.

Can we observe the consequences of these factors And model their impact on organic matter oxidation?

Biogeochemical cycling in Cape Lookout Bight Sediments

Biogeochemical cycling in Cape Lookout Bight

Can we achieve chemical balance if the system is at steady state?

Seasonal distribution of sulfate and acetate In Cape Lookout Bight sediments

Note that sulfate is exhausted closer to the sediment/water interface in summer due to enhanced rate of sulfate reduction. Acetate accumulates only when sulfate reaches low concentrations

Measurement of sulfate Oxidation using ³⁴S

Moles of sulfate reduced (m⁻² yr⁻¹)

	tube	³⁵ S	
Jan	8.0	12.2	
Feb	7.7	13.3	
Mar	11.4	12.7	
Apr	15.2	7.4	
May	11.8	16.4	
Jun	16.8	20.9	
July	22.0	25.4	
Aug	27.2	30.0	
Sept	26.2	23.3	
Oct	17.5	16.2	
Nov	8.7	10.0	
Dec	6.4	11.1	
	15.1	16.6	

 $2CH_2O + SO_4^{=} \longrightarrow 2CO_2 + H_2S + H_2O$

Therefore 32 moles C by sulfate oxidation

Methane fluxes from CLB sediments

Methane diffusive fluxes

Date	J_{s}	J _{theor}	J _{excess}
16 October 76	84	31	53
6 November 76	30	26	4
25 March 77	51	27	24
16 May 77	48	50	0
27 June 77	82	69	13
13 July 77	130	78	52
5 August 77	274	115	159
22 August 77	393	129	264
20 September 77	186	160	25
17 November 77	66	46	20

Calculation of Methane flux from CLB sediments

Methane concentrations build up to saturation values and form bubbles that remove CO2 and methane from sediments

	ΣCO2 fluxes µm m ⁻² hr ⁻¹		CH4 fluxes µm m ⁻² hr ⁻¹			
Month	diff	bubble	Total	diff	bubble	Total
Jan	1900	0	1900	50	0	50
Feb	1900	0	1900	50	0	50
March	1900	0	1900	51	0	51
April	1900	0	1900	50	0	50
May	3825	0.6	3826	48	52	100
June	5400	5.6	5406	82	480	562
July	6000	20	6020	130	1740	1870
Aug	6300	26	6326	334	2250	2484
Sept	4900	18	4918	184	1500	1684
Oct	2700	6.5	2707	84	560	644
Nov	2200	0.8	2201	48	65	113
Dec	1900	0	1900	50	0	50
Average	3400	6.5	3407	97	544	651
mean monthly flux (mole m ⁻² yr ⁻¹)						
	29.8	0.057	29.9	0.85	4.85	5.70

Total C remineraliztion in CLB sediments

out of bottom		
$\Sigma CO_2 = 5.7$ $\Sigma CH_4 = 0.1$		
5.8		

remineralization = C flux

Total C remineralization 41.4 moles m⁻² yr⁻¹ Total Carbon flux = 41.4

Methane flux = 5.8

total C remineraliztion due to methanogenesis:

5.8 x 2 = 11.6 moles m-2 yr-1 or 28%

C reminerealization due to sulfate reduction is:

41.4 - <u>11.6</u> 29.8 moles C m-2 yr-1 or 72%

Sulfate reduction measured or calculated from tube/tracer incubations and sulfate gradient is 30-36 moles yr-1 Production of methane from acetate and CO₂ in CLB sediments. ¹⁴C tracer studies.

Integrated (0-35 cm)rates of methane production and sulfate reduction in CLB sediments

Date	Т ^о С	CH ₄	SO ₄
21 Feb.81	11	0.04	1.00
24 April 81	17	0.03	2.40
19 May 80	23	0.05	
29 May 80	25	0.56	0.96
2 Jul. 81	23	1.60	
2 Aug. 81	27	2.00	2.60
30 Aug. 81	26	1.00	2.90
5 Oct. 80	26	0.79	4.40
8 Oct. 79	22	1.10	3.60
9 Nov. 81	19	0.02	1.70

mmol m⁻² h⁻¹

Summary:

Terminal electron acceptors are used in the order of free E yields (O_2 , Fe/Mn, SO₄, CO₂)

Anoxic sediments are biogeochemically zoned according to e- acceptors

Organic matter is oxidized by microbial consortia - no single organism degrades complex organic matter to CO_2 . Instead, fermentation produces VFAs which are Used by acetophiles to yield CO_2 and methane.

Methane is produced by two reactions CO_2 redn with Hydrogen, and disproportionation of acetic acid.