

Competition for acetate and hydrogen between Desulfobacter postgatei and Methanosarcina bakerii



Molecular hydrogen as a control on organic matter oxidation in anoxic sediments

Is C oxidation in anoxic sediments under thermodynamic regulation?

 $(CH_2O)n + nH_2O --> nCO_2 + 2nH_2$

 $2nH_2 + mX_{ox} \rightarrow mX_{red} + zH_2O$

 $(e.g. X_{ox} = SO_4^{2-} X_{red} = S^{2-})$ $\Delta G_{rxn} = \Delta G_{(T)}^{o} + RT \ln \left(\left\{ X_{red} \right\}^m / \left\{ X_{ox} \right\}^m (P_{H2})^{2n} \right)$

and...

$$\mathsf{P}_{H2} = \left(\{\mathsf{X}_{red}\}^m / \{\mathsf{X}_{ox}\}^m \, \mathsf{e}(\Delta \mathsf{G}_{rxn} - \Delta \mathsf{G}_{(t)}^o / \mathsf{RT}) \right)^{1/2n}$$

Hoehler et al. (and others...) developed a headspace analysis To measure hydrogen in cores. Cores were manipulated By adding a TEA or Δ T, Δ pH, Δ [TEA], etc. and measured Steady state concentration of hydrogen

$$\mathsf{P}_{H2} = \left(\{\mathsf{X}_{\mathsf{red}}\}^{\mathsf{m}} / \{\mathsf{X}_{\mathsf{ox}}\}^{\mathsf{m}} \; \mathrm{e}^{(\Delta \mathsf{Grxn} - \Delta \mathsf{G}(\mathsf{t}) \mathsf{o} / \mathsf{RT})} \right)^{1/2\mathsf{n}}$$

$\Delta G (KJ/mol)$

NO_3^-	\rightarrow N ₂	-448

$Mn(IV) \longrightarrow Mn(II) -3$

Fe(III)	Fe(II)	-114
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$$SO_4^{2-} \longrightarrow S^{2-}$$
 -77

 $CO_2 \longrightarrow CH_4 -58$

Effect of TEA on H₂ concentrations



Effect of temperature on H₂ concentrations



Dependence of $[H_2]$ on $[SO_4^{2-}]$

 $2(CH_2O) + SO_4^{2-}$ $2CO_2 + 4H_2O + S^{2-}$

 $P_{H2} = ({X_{red}}^m / {X_{ox}}^m e(\Delta G_{rxn} - \Delta G_{(t)}^o / RT))^{1/2n}$



Fig. 5. Response of hydrogen concentrations to variations in porewater sulfate concentration. Error bars represent one standard deviation about the mean of triplicate sediment samples. A power function fit to the data indicates that hydrogen has an exponential dependence of -0.26 ± 0.01 on sulfate (compare to theoretical value of -0.25).

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Sulfate Concentration (mM)

-O- Hydrogen Concentration (nM)

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Sulfate Concentration (mM)

-O- Hydrogen Concentration (nM)



Effect of temperature on H₂ concentrations



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Sulfate Concentration (mM)

Effect of sulfate on H₂ in CLB sediments



Fig. 8. The dependence of hydrogen concentrations on sulfate concentrations in the November core from Cape Lookout Bight (Fig. 7b). (a) blow-up of the 12–16 cm depth interval. Note that sulfate concentrations only reach threshold values below 16 cm; (b) plot of hydrogen concentration vs. sulfate concentration over the 12–16 cm interval. A power function fit to the data indicates that hydrogen has an exponential dependence of 0.30 ± 0.04 on sulfate (compared to a lab value of 0.26 ± 0.01 and a theoretical value of 0.25).

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Sulfate Concentration (mM)

Hydrogen as a control on organic matter oxidation In anoxic sediments (fresh and marine)

Hydrogen is a by-product of fermentation and is essential for sulfate reduction and methanogenesis.

Hydrogen concentrations respond to T, [X], pH. laboratory changes correspond well to field observations.

Variations in $\rm H_2$ suggest maintenance of constant ΔG values of -10 to -15 kJ mol^-1 .

 H_2 has a very short lifetime in sediments- makes an excellent E regulator. Small changes in H_2 concentration results in large changes in ΔG .

Intense competition by bacteria/archea regulate [H₂]

It was also recognized very soon after sedimentary methane profiles were compiled that methane consumption appeared to be occurring in marine systems



Anaerobic methane oxidation...where has all the methane gone?

Oceans have a huge reservoir of methane in sediments, but Contribute only 2% of the global atmospheric flux of methane.

CH ₄ + SO ₄ ²⁻	\longrightarrow	$HCO_{3}^{-} + HS^{-} + H_{2}O - 25 \text{ kJ/mol}$
CH ₄ +2H ₂ O	\longrightarrow	$HCO_{3}^{-} + 4H_{2}$
$SO_4^{2-} + 4H_2$	\longrightarrow	$HS^{-} + 2H_2O$

Energetically favorable, but depends critically on the concentration of hydrogen. Only favorable in CLB sediments when $[H_2] < 0.29 \text{ nM!}$

Anaerobic methane oxidation probably occurs as a consortia between SRB and MO Archea

Coupled methane oxidation and sulfate reduction in CLB sediments



Hydrogen concentrations are < nM

Coupled methane oxidation and sulfate reduction in CLB sediments

summertime



Hydrogen concentrations are a few nM

We can use molecular probes to identify sulfate reducing baceria (green) and methane oxidizing Archea (red) to visually examine the consortia



FIG. 5. Whole-cell fluorescent in situ hybridization of methane-oxidizing consortia found in Monterey Canyon seep sediments (A) and in Monterey Canyon nonseep sediments after incubation on AMIS (B). Sediments were fixed in 4% formalin and transferred into a 1:1 ethanolphosphate-buffered saline solution for storage. Green-stained cells correspond to *Desulfosarcina-Desulfococcus* bacteria (DSS658), and red-stained cells correspond to the archaeal ANME-2 group (EelMSMX932). Separate images were overlaid to represent the structure of the aggregate. Scale bar, 10 μm. No consortia were detectable in Monterey Canyon nonseep sediments before incubation on the AMIS.

AOM consortia of different shapes

<text>

aggregate-structure

AOM consortium above gas hydrates at Hydrate Ridge, Cascadia Margin. "tissue"-structure



AOM consortium above gas seeps in the anoxic Black Sea.



The History of AOM

In 2001 Walter Michaelis et al. found out, that such AOM-consortia are able to build up a huge biomass above methane seeps in the anoxic part of the Black Sea. These reeflike structures are up to 1 m in diameter and 4 m high.



photos: GHOSTDABS, Jago-Team



Molecular identification reveal that a microbial consortium is responsible for AOM in the reef







Carbon isotopic changes in atmospheric methane



Methane production rate profiles



Date	Тетр	CO ₂	acetate	total	%total production
7/21/83	26.5	0.70	0.18	0.88	20
8/7/83	27	1.02	0.33	1.35	24
8/29/83	27.5	1.10	0.46	1.56	29

Differences in carbon isotope fractionation between freshwater and marine systems





Differences in carbon isotope fractionation between freshwater and marine systems



Freshwater

Marine



Carbon isotope fractionation with methanogenesis



Carbon and Hydrogen isotopes fractionation with methanogenesis



Production of methane from acetate and CO₂ in CLB sediments. ¹⁴C tracer studies.



Seasonal changes in ¹³C for methane and CO₂

Table 1. Cape Lookout Bight sediment gas bubble composition and δ^{13} C data. Values listed are means \pm SD for the number of sample bottles listed. Superscripts indicate the number of samples for which compositional data were obtained when different from the number of sample bottles listed.

Date	Methane sample bottles (no.)	Methane content (%)	δ ¹³ C-CH₄ (per mil)	Carbon dioxide sample bottles (no.)	Carbon dioxide content (%)	δ ¹³ C-CO ₂ (per mil)
6 June 1983	5	97 ± 2	-64.5 ± 0.7	5	2.5 ± 0.1	-6.8 ± 1.1
19 July 1983	6	95 ± 4	-62.2 ± 0.4	6	3.4 ± 0.2^3	-8.6 ± 1.2
3 August 1983	5	96 ± 4	-61.7 ± 0.9	5	2.4 ± 0.3	-8.8 ± 1.0
19 August 1983	5	94 ± 2	-57.5 ± 0.3	4	2.4 ± 0.2	-9.4 ± 0.3
15 September 1983	5	97 ± 2	-60.3 ± 0.4	5	2.5 ± 0.1	-8.3 ± 0.5
16 October 1983	6	95 ± 3	-60.0 ± 0.5	5	2.4 ± 0.5^4	-7.2 ± 0.6
20 November 1983	4	93 ± 2	-62.2 ± 0.4	4	2.4 ± 0.6	-8.0 ± 0.2
2 February 1984	4	98 ± 3	-63.4 ± 0.6	4	1.6 ± 0.5^{3}	-6.0 ± 1.2
7 April 1984	4	94 ± 3^{3}	-63.8 ± 0.2	4	1.0 ± 0.2^{3}	-5.1 ± 0.7
6 May 1984	4	90 ± 6	-63.8 ± 0.4	. 3	1.5 ± 0.2	-3.0 ± 0.8
31 May 1984	5	94 ± 5	-68.5 ± 0.7	3	1.8 ± 0.6	-7.0 ± 2.0
14 June 1984	5	94 ± 3	-64.1 ± 0.6	4	2.9 ± 1.0	-6.2 ± 2.4
2 July 1984	. 4	97 ± 4^2	-59.4 ± 1.2	2	2.1 ± 0.1	-10.0 ± 0.7
18 July 1984	4	98 ± 2^2	-60.6 ± 1.6	2	2.2 ± 0.2	-10.6 ± 3.2
11 August 1984	5	98 ± 3⁴	-57.3 ± 0.6	5	2.3 ± 0.2	-7.6 ± 1.2
30 August 1984	4	94 ± 1	-57.9 ± 1.0	3	3.8 ± 1.1	-8.9 ± 1.1
22 September 1984	5	99 ± 0^2	-58.0 ± 0.3	5	2.4 ± 1.3	-8.1 ± 1.0

Changes in $\Delta^{13}\mathrm{C}$ in CLB methane



Changes in $\Delta^{13}\mathrm{C}$ in CLB methane

